

Cyto-Genic Relationship Among Cytoplasmic-Genetic Male Sterile, Maintainer and Restorer Lines of Rice

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The cyto-genic relationship was determined among six of the cytoplasmic-genetic male sterility (cms) rice lines: Zhen Shan 97A, V20A (possessing wild rice *Oryza sativa* f. *spontanea* or *O. perennis* cytoplasm?), Yar Ai Zhao A (possessing Gambiaca cytoplasm), Pankhari 203A (possessing Taichung Native 1 cytoplasm), Wu 10A (possessing Chinsurah Boro II or BT cytoplasm), and MS 577A (possessing *O. sativa* f. *spontanea* cytoplasm). Based on the results of detailed crosses, the six cms lines were classified into four different cyto-sterility systems as follows: Wu 10A and Pankhari 203A (S_1), Zhen Shan 97A/V20A (S_2), Yar Ai Zhao A (S_3) and MS577A (S_4). The corresponding nuclear genes interacting with a specific cytoplasm to induce sterility in these lines were designated as S_1 -rf (for S_1), S_2 -rf (for S_2), S_3 -rf (for S_3) and S_4 -rf (for S_4).

Key words: cytoplasmic male sterility, hybrid rice, nuclear genes, maintainer lines, restorer lines.

During the past decade hybrid rice technology has been developed and used successfully in China (Yuan, 1977) where about 6 million ha (17% of the total rice area) were planted to hybrid rice varieties (Lin and Yuan, 1980; Virmani et al., 1981). These varieties have 20 to 30% yield advantage and wider adaptability in comparison to conventionally-bred rice varieties.

Development of hybrid rice varieties involves primarily the use of cytoplasmic genetic male sterility and fertility restoration system. The role of cytoplasm in causing male sterility in rice was first reported in 1954 (Weeraratne, 1954; Sampath and Mohanty, 1954). Later, Katsuo and Mizushima (1958) and Kitamura (1962a, b) also observed this phenomenon in inter-specific and inter-varietal crosses. Shinjyo and Omura (1966) developed the first cytoplasmic male sterile line in cultivated rice by substituting nuclear genes of a japonica variety, Taichung 65 into the cytoplasm of indica variety Chinsurah Boro II. This line was designated as BT cyto-sterile and the cyto-sterility system was designated as BT. Erickson (1969) and Carnahan et al. (1972) developed cytoplasmic male sterile lines from crosses of an indica variety Birco (PI 279120), with japonica rice varieties of California viz., Calrose, Caloro and Colusa. Watanabe (1971) and Carnahan et al. (1972) developed also cytoplasmic genetic male sterile lines from indica/japonica crosses and *Oryza glaberrima*/Colusa cross respectively. Moreover, Athwal and Virmani (1972) bred a cytoplasmic male sterile line from an indica/indica cross, Taichung Native 1/Pankhari 203.

The first cytoplasmic male sterile line used in developing commercial F_1 rice hybrids was developed in China in 1973 from a sterile plant (wild aborted) naturally occurring in a wild rice population (*O. sativa* f. *spontanea* or *O. perennis*?) on Hainan island (Hunan Prov. Agr. Res. Inst., 1977; Yuan, 1977). Subsequently, cytoplasmic male sterile lines have been developed from various accessions of *O. sativa* f. *spontanea*, indica variety Gambiaca (from Africa), and a Chinese variety O-Shan-Tao-Bai (Lin and Yuan, 1980). Currently, about 95% of the area allotted to hybrid rice in China is planted to the hybrids, which are derived from cyto-sterile lines designated as WA (wild aborted) cyto-steriles. However, this situation makes F_1 rice hybrids potentially susceptible to a disease or insect that could be associated with the cytoplasmic factor. It is, therefore, desirable to have diverse sources of cytoplasmic male sterility in rice. The International Rice Research Institute (IRRI) has a collection of cytoplasmic and sterile lines, some of which are derived from different cytoplasmic sources (Virmani et al., 1981). This study was conducted to determine cyto-genic relationship among six of these cms lines to identify different sources of cytoplasmic male sterility.

MATERIALS AND METHODS

Six cytoplasmic-genetic male sterile and their maintainer lines (Table 1) found stable for pollen sterility were used. The following sets of crosses were made: (1) the six cms (A) lines crossed with the six maintainer lines in all possible A/B crosses; (2) the six B lines crossed in all possible B/B crosses; (3) the six A lines crossed with 31 elite lines; and (4) the six B lines crossed with 31 elite lines.

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Table 1. The six cytoplasmic male sterile and their maintainer lines used in the study.

CMS	Maintainer line	Origin	Source of cytoplasm	Remark
Wu 10A	Wu 10B	China	Chinsurah Boro II	Japonica, semidwarf
Pankhari 203A (P 203A)	Pankhari 203B (P 203B)	IRRI	TN1	Indica, photosensitive
Zhen Shan 97A (97A)	Zhen Shan 97B (97B)	China	Wild rice with aborted pollen (<i>O. sativa</i> f. <i>spontanea</i> ?)	Indica, semidwarf
V20A	V20B	China	Wild rice with aborted pollen (<i>O. sativa</i> f. <i>spontanea</i> ?)	Indica, semidwarf
Yar-Ai-Zhao A 577A	Yar-Ai-Zhao B 577B	China Korea	Gambiaca <i>O. sativa</i> f. <i>spontanea</i>	Indica, semidwarf Indica, semidwarf

The parents and F_1 s of these crosses were grown during 1982 wet season at the rate of 15 plants each in single row plots (3.0 m long). Seedlings were transplanted at the rate of one seedling per hill with a spacing of 30 x 20 cm. Fertilizer was applied at the rate of 80-30-40 (N, P, K) kg per ha. The recommended cultural practices with regard to insect, and weed control were followed.

From each F_1 and parental lines between 40 to 50 spikelets per plant were collected a day before anthesis. Five anthers were collected randomly from the spikelets of each plant to determine pollen fertility. Pollen grains were stained in 1% I_2KI solution and observed under light microscope. Based on their staining behavior and shape they were classified into four categories viz., unstained withered (UW), unstained spherical (US), partially stained round (PSR) and completely stained round (CSR). The first three categories of pollen grains were considered inviable, however, judgment on the viability for the completely stained round pollen grains was based on the spikelet fertility of the bagged panicles.

Spikelet fertility was determined by counting the total number of seed set in proportion to the total number of spikelets in the two primary panicles which had been bagged before flowering.

Mean pollen and spikelet fertility in the F_1 s in relation to the parents were used to determine cyto-genetic relationships among various cms, maintainer and restorer lines. Linear correlation coefficients were calculated for spikelet fertility in A/B crosses (involving the six cms lines and all the maintainer lines) and A/elite line crosses (involving the cms lines with all the elite lines). The correlation coefficient values for each cms line in relation to the other cms line were used also to determine the cyto-genetic relationship among the cms and maintainer lines.

RESULTS AND DISCUSSION

Pollen and Spikelet Fertility of CMS and Maintainer Lines. Frequency of different categories of pollen grains in the selected cms and maintainer lines indicated that the majority of pollen grains of Zhen Shan 97A (97A), V20A,

Yar Ai Zhao A (YAZ A) and Pankhari 203A (P 203A) were unstained withered while majority of pollen grains of Wu 10A and MS 577A were partially stained round to completely stained round (Table 2). P 203A and YAZ A possessed pollen grains of all the four categories but 86 and 90%, respectively, were unstained withered. Pollen grains of the maintainer lines were mostly (96 to 99%) completely stained and round.

Spikelet fertility was zero in 97A, V20A, YAZ A and P 203A possessing mostly unstained pollen grains indicating that unstained pollen grains were inviable. Wu 10A and MS 577A possessing stained and round pollen grains also did not show any seed set on bagged panicles. Apparently, stained round pollen grains of these cyto-sterile lines were also inviable and did not affect fertilization. The cyto-sterile Wu 10A, is derived from BT cyto-sterility system which is known to be gametophytic in nature (Shinryo, 1969, 1975). Pollen grains of this cms line are known to abort at two nuclear stages of development as compared to 'WA' cyto-steriles in which case the pollen abortion takes place at uni-nuclear stage (Xu, 1982). It appears that stainability reaction and shape of pollen grains in male sterile lines is determined by the stage at which pollen grains abort. The pollen grains aborting at uni-nucleate development stage would be unstained, withered spherical but those aborting at bi-nucleate or later stage would be partially to completely stained and round. By implication, therefore, pollen grains of MS 577A, which are stained normally may be aborting at or after bi-nuclear stage of pollen development. Chaudhary et al. (1981) also derived similar conclusion from their studies on pattern of pollen abortion on some cms lines of rice.

All the six maintainer lines possessing 96 to 99% completely stained round pollen grains showed 85 to 94% spikelet fertility. Their pollen grains were, therefore, viable.

Pollen and Spikelet Fertility in A/B and B/B Crosses. The pollen and spikelet fertility of diallel crosses involving the six cms and their maintainer lines (Table 3) indicated that: (a) the male sterility of Wu 10A was maintained by

Wu 10B and P 203B, the other maintainers acted as partial restorers; (b) the male sterility of 97A and V20A was maintained by all the maintainer lines except YAZ B, the latter behaved as partial restorer; (c) the male sterility of YAZ A was maintained by YAZ B, P 203B, and Wu 10B, the other three maintainer lines behaved as partial restorers; and (d) the male sterility of MS 577A was maintained only by MS 577B, the crosses with other maintainers showed high frequency of completely stained round pollens (like MS 577A) but low spikelet fertility.

These results imply that: (a) the cytoplasmic male sterility system in Wu 10A and P 203A is identical; (b) the cytoplasmic male sterility systems in 97A and V20A is identical; (c) the cytoplasmic male sterility system in YAZ A is different from that of Wu 10A/P 203A, 97A/V20A, and MS 577A; and (d) the cytoplasmic male sterility system of MS 577A is different from Wu 10A/P 203A, 97A/V20A, and YAZ A.

Pollen and spikelet fertility of all possible B/B crosses (Table 4) indicated that: (a) crosses involving 97B, V20B, YAZ B and 577B were normally fertile, without showing reciprocal differences and (b) crosses involving Wu 10B and P 203B used either as maternal or as paternal parent among themselves and with the other maintainer lines showed partial fertility, without showing large and consistent differences between the reciprocal crosses.

Table 2. Frequency of different categories of pollen grains in six cytoplasmic male sterile and maintainer lines of rice.

Line	% Frequency of pollen grain in categories [†]				Spikelet fertility (%)
	UW	US	PSR	CSR	
Wu 10A	22.8	7.1	25.1	45.0	0
Wu 10B	1.9	2.1	0.1	95.9	87
P 203A	86.5	1.8	4.6	7.1	0
P 203B	0.6	2.0		97.4	94
97A	97.2	2.8			0
97B	0.6	0.5		98.9	92
V20A	97.6	2.4			0
V20B	0.8	0.8		98.4	92
YAZ A	90.4	8.0	0.8	0.8	0
YAZ B	1.3	0.8		97.9	85
577A	1.6	5.0	23.4	70.0	0
577B	0.5	1.5		98.0	91

† UW = Unstained withered.
 US = Unstained spherical.
 PSR = Partially stained round.
 CSR = Completely stained round.

Table 3. Frequency of completely stained round (CSR) pollen (%) and spikelet fertility (%) of F₁s derived from the six cms lines with the maintainer lines in all possible crosses.

CMS Lines	Percent CSR pollen and spikelet fertility [†] of F ₁ s involving						Mean
	Wu 10B	P 203	97B	V20B	YAZ B	577B	
Wu 10A	45 (0)	33 (1)	49 (21)	56 (12)	84 (27)	53 (11)	53 (12.1)
P 203A	56 (2)	7 (0)	59 (21)	56 (22)	55 (29)	54 (20)	47.6 (15.6)
97A	0 (0)	0 (0)	0 (0)	0 (0)	33 (13)	0 (0)	5.5 (2.2)
V20A	0 (0)	0 (0)	0 (0)	0 (0)	11 (10)	0 (0)	1.9 (1.7)
YAZ A	0 (0)	2 (1)	50 (25)	43 (26)	1 (0)	27 (16)	28.7 (11.2)
577A	50 (6)	74 (5)	76 (6)	70 (4)	82 (7)	70 (0)	70 (4.8)
Mean	25.2 (1.3)	19.4 (1.2)	38.8 (12.4)	37.5 (10.7)	44.3 (14.2)	42.2 (7.8)	

† Spikelet fertility in parenthesis.

Table 4. Frequency of CSR pollen and spikelet fertility (%) in F₁s involving the six maintainer lines in all possible combinations.

CMS Lines	Percent CSR pollen and spikelet fertility [†] of F ₁ s involving					
	Wu 10B	P 203B	97B	V20B	YAZ B	577B
Wu 10B	96 (87)	57 (27)	48 (47)	19 (18)	92 (25)	60 (38)
P 203B	49 (28)	97 (94)	45 (46)	31 (25)	37 (26)	44 (20)
97B	76 (41)	47 (49)	99 (92)	99 (89)	99 (84)	97 (70)
V20B	20 (16)	47 (25)	99 (94)	98 (92)	99 (84)	99 (86)
YAZ B	99 (26)	40 (24)	99 (72)	99 (65)	98 (85)	94 (91)
577B	50 (29)	36 (23)	95 (94)	96 (90)	98 (91)	98 (91)

[†] Spikelet fertility in parenthesis.

Table 5. Linear correlation coefficients for spikelet fertility of sets of A/B crosses involving different cms lines.

CMS Line	P 203A	97A	V20A	YAZ A	577A
Wu 10A	0.92*	0.69	0.69	0.41	0.24
P 203A		0.55	0.55	0.47	-0.07
97A			1.00**	-0.44	0.46
V20A				-0.44	0.46
YAZ A					0.24

** Significant at 1% level; * Significant at 5% level.

Partial fertility of crosses involving P 203B and Wu 10B with the other four maintainer lines may be due to gametic development genes identified by Oka (1953, 1954). Therefore, it may be concluded that cytoplasm of the six maintainer lines do not possess factor(s) inducing male sterility.

The values of correlation coefficient for spikelet fertility of A/B crosses involving the six cms lines and all the maintainer lines are given in Table 5. These values indicated that behavior of Wu 10A and P 203A in crosses with the maintainer lines was highly correlated ($r = 0.92$); 97A and V20A also showed similar behavior ($r = 1$). These results further confirmed that cyto-sterility systems in cyto-sterile W 10A and P 203A, and 97A and V20A, were the same. All other possible comparisons among cms lines showed non-significant correlations, hence, the cyto-sterility systems possessed by them should be different.

Spikelet Fertility in Crosses of CMS Lines with Elite Lines. Values of correlation coefficients on spikelet ferti-

Table 6. Linear correlation coefficient for spikelet fertility of sets of A/elite line crosses involving different cms lines.

CMS Line	P 203A	97A	V20A	YAZ A	577A
Wu 10A	0.29	0.06	0.10	0.26	-0.12
P 203A		0.06	0.04	0.33	0.19
97A			0.99**	0.55**	0.16
V20A				0.48*	0.01
YAZ A					0.03

** Significant at 1% level; * Significant at 5% level.

lity of cross involving the cms lines with elite lines are given in Table 6. The results confirmed that the behavior of 97A and V20A in crosses with elite lines was highly correlated ($r = 0.99$), however, behavior of crosses involving Wu 10A and P 203A did not show significant correlation ($r = 0.29$). Wu 10A is a japonica line and P 203A is derived from Pankhari 203, the latter is known for giving less fertile hybrids regardless of other parent involved (Engle, 1970; Parmar et al., 1981), therefore, the confounding effect of inter-varietal hybrid sterility due to nuclear genes (Oka, 1953; 1954) can not be ruled out. The relationship between Wu 10A and P 203A may be studied further by following up the crosses showing differential behavior in backcross generations where the effects of inter-varietal hybrid sterility are reduced. Correlated behavior of Yar Ai Zhao A with 97A ($r = 0.55$) and V20A ($r = 0.41$) may be due to the ability of some elite lines to restore fertility in both cms lines.

Table 7. Proposed cytoplasmic and genotypic constitution of the six cms and their maintainer lines.

CMS/maintainer line	Cytoplasmic constitution	Genotype							
Wu 10A	S ₁	S ₁ -rf	S ₁ -rf	S ₂ -rf	S ₂ -rf	S ₃ -rf	S ₃ -rf	S ₄ -Rf	S ₄ -Rf
Wu 10B	N ₁	S ₁ -rf	S ₁ -rf	S ₂ -rf	S ₂ -rf	S ₃ -rf	S ₃ -rf	S ₄ -Rf	S ₄ -Rf
P 203A	S ₁	S ₁ -rf	S ₁ -rf	S ₂ -rf	S ₂ -rf	S ₃ -rf	S ₃ -rf	S ₄ -Rf	S ₄ -Rf
P 203B	N ₁	S ₁ -rf	S ₁ -rf	S ₂ -rf	S ₂ -rf	S ₃ -rf	S ₃ -rf	S ₄ -Rf	S ₄ -Rf
97A	S ₂	S ₁ -Rf	S ₁ -Rf	S ₂ -rf	S ₂ -rf	S ₃ -Rf	S ₃ -Rf	S ₄ -Rf	S ₄ -Rf
97B	N ₂	S ₁ -Rf	S ₁ -Rf	S ₂ -rf	S ₂ -rf	S ₃ -Rf	S ₃ -Rf	S ₄ -Rf	S ₄ -Rf
V20A	S ₂	S ₁ -Rf	S ₁ -Rf	S ₂ -rf	S ₂ -rf	S ₃ -Rf	S ₃ -Rf	S ₄ -Rf	S ₄ -Rf
V20B	N ₂	S ₁ -Rf	S ₁ -Rf	S ₂ -rf	S ₂ -rf	S ₃ -Rf	S ₃ -Rf	S ₄ -Rf	S ₄ -Rf
YAZ A	S ₃	S ₁ -Rf	S ₁ -Rf	S ₂ -Rf	S ₂ -Rf	S ₃ -rf	S ₃ -rf	S ₄ -Rf	S ₄ -Rf
YAZ B	N ₃	S ₁ -Rf	S ₁ -Rf	S ₂ -Rf	S ₂ -Rf	S ₃ -rf	S ₃ -rf	S ₄ -Rf	S ₄ -Rf
577A	S ₄	S ₁ -Rf	S ₁ -Rf	S ₂ -rf	S ₂ -rf	S ₃ -Rf	S ₃ -Rf	S ₄ -rf	S ₄ -rf
577B	N ₄	S ₁ -Rf	S ₁ -Rf	S ₂ -rf	S ₂ -rf	S ₃ -Rf	S ₃ -Rf	S ₄ -rf	S ₄ -rf

On the basis of the foregoing results the six cms lines can be classified into four groups of cyto-steriles viz., Wu 10A/P 203A, 97A/V20A, YAZ A and MS 577A. The cytoplasmic constitution of these four groups may be designated tentatively as below:

Wu 10A/P 203A	S ₁	(designated as 'BT' by Shinjyo, 1969)
97A/V20A	S ₂	(designated as 'WA' by Yuan, 1977; Lin and Yuan, 1980)
Yar Ai Zhao	S ₃	(designated as 'Gam' by Lin and Yuan, 1980)
MS 577A	S ₄	(not designated earlier)

Shinjyo (1969) designated the nuclear gene interacting with the 'BT' cytoplasm as 'rf'. Since a number of sterility inducing cytoplasm have been identified, the corresponding nuclear genes interacting with the specific cytoplasmic factor inducing male sterility in these lines may be designated as S₁-rf (for S₁), S₂-rf (for S₂), S₃-rf (for S₃) and S₄-rf (for S₄). Based on these assumptions and the results presented in Table 3 cytoplasmic and genotypic constitution of the six cms and maintainer lines are proposed in Table 7. Studies conducted in China (Zhu, 1979) have also shown that 'WA' cyto-sterility system is different from 'BT' system.

The use of diverse sources of cytoplasmic male sterility in hybrid rice development should help reduce the possibilities of repeating 'Southern corn blight situation' in rice.

LITERATURE CITED

Athwal, D.S. and S.S. Virmani. 1972. Cytoplasmic male sterile and hybrid breeding in rice. *In* Rice Breeding.

International Rice Research Institute, Los Banos, Laguna, Philippines. pp. 615-620.

Carnahan, H.L., J.R. Erickson, S.T. Tseng, and J. N. Rutger. 1972. Outlook for hybrid rice in the U.S.A. *In* Rice Breeding. International Rice Research Institute, Los Banos, Laguna, Philippines. pp. 603-607.

Chaudhary, R.C., Virmani, S.S. and Khush, G.S. 1981. Patterns of pollen abortion in some cytoplasmic-genetic male sterile lines of rice. *Oryza* 18:140-142.

Engle, L.M. 1970. The cytogenetics of sterility in F₁ hybrids of indica x indica and indica x japonica varieties of rice, *Oryza sativa* L. M.S. Thesis, University of the Philippines, Los Banos.

Erickson, J.R., 1969. Cytoplasmic male sterility in rice (*Oryza sativa* L.). *Agron. Abst.* 1969:6.

Hunan Provincial Rice Research Institute. 1977. Breeding success with hybrid rice a song of triumph composed according to Mao Tse Tung's idealistic score, *Zhongguo Nongye Kexue* (Chinese Agricultural Sciences) 1:21-26 (Original in Chinese).

Katsuo, K. and U. Mizushima. 1958. Studies on the cytoplasmic difference among rice varieties, *Oryza sativa* L. I. On the fertility of hybrids obtained reciprocally between cultivated and wild varieties. *Jap. J. Breed.* 8:1-5. (In Japanese).

Kitamura, E. 1962a. Studies on cytoplasmic sterility of hybrids in distantly related varieties of rice, *Oryza sativa* L. Fertility of F₁ hybrids between strains derived from a certain Philippine x Japanese variety crosses and Japanese varieties. *Japan. J. Breed.* 12:81-84.

Kitamura, E. 1962b. Studies on cytoplasmic sterility of hybrid in distantly related varieties of rice, *Oryza sativa* L. II. Analysis of nuclear genes in Japanese varieties

- controlling cytoplasmic sterility. Japan. J. Breed. 12: 166-168.
- Lin, S.C. and L.P. Yuan. 1980. Hybrid rice breeding in China. p. 35-51. *In Innovative Approaches to Rice Breeding*. IRRI, Los Banos, Laguna, Philippines.
- Oka, H.I. 1953. Influence of inter-varietal hybrid sterility on segregation ratios in rice (Phylogenetic differentiation of the cultivated rice plant IX). Jap. J. Breed. 3:31-39.
- Oka, H.I. 1954. Classification of rice varieties by inter-varietal hybrid sterility. Japan. J. Breed. 3(3-4):1-6. (In Japanese).
- Parmar, K.S., E.A. Siddiq, and M.S. Swaminathan, 1981. Evaluation of known and new sources of cytoplasmic male sterility-restorer systems in cultivated rice, *Oryza sativa* L. Z. Pflanzenzuchtg. 86:1-10.
- Sampath, S. and H.K. Mohanty. 1954. Cytology of semi-sterile rice hybrid. Cur. Sci, 23:182-183.
- Shinjo, C. 1969. Cytoplasmic-genetic male sterility in cultivated rice, *Oryza sativa* L. II. The inheritance of male sterility. Japan J. Genetics 44:149-159.
- Shinjo, C. 1975. Genetical studies of cytoplasmic male sterility and fertility restoration on rice, *Oryza sativa* L. Bulletin of the College of Agriculture, University of Kyukyus No. 22.
- Shinjo, C. and Omura. 1966. Cytoplasmic-genetic male sterility in cultivated rice, *Oryza sativa* L. I. Fertilities of F_1 , F_2 and offsprings obtained from their mutual reciprocal backcrosses; and segregation of completely male sterile plants. Japan. J. Breed. 16 (Suppl. 1):179-180. (In Japanese).
- Virmani, S.S., R.C. Chaudhary, and G.S. Khush. 1981. Current outlook on hybrid rice. *Oryza* 18 :67-84.
- Watanabe, Y. 1971. Establishment of cytoplasmic and genetic male sterile lines by means of indica-japonica cross. *Oryza* 8(2):9-16.
- Weeraratne, H. 1954. Hybridization technique in rice. Trop. Agric. (Colombo) 110:93-97.
- Xu Shuhua. 1982. Cytological observation on pollen development in the main male sterile types of rice (*Oryza sativa* L.) developed in China. *Scientia Agriculture Sinica* 2:9-14. (Chinese with English summary).
- Yuan, L.P. 1977. The execution and theory of developing hybrid rice. *Zhonggong Nongye Kexue* (Chinese Agricultural Sciences) 1:27-31. (Original in Chinese).
- Zhu, Y.G. 1979. Investigation of different male-sterile lines of rice with various cytoplasm. *Acta Genetica Sinica* 6 :9-11.