

A Framework for Classification of Prokaryotic Protein Kinases

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Abstract

Background: Overwhelming majority of the Serine/Threonine protein kinases identified by gleaning archaeal and eubacterial genomes could not be classified into any of the well known Hanks and Hunter subfamilies of protein kinases. This is owing to the development of Hanks and Hunter classification scheme based on eukaryotic protein kinases which are highly divergent from their prokaryotic homologues. A large dataset of prokaryotic Serine/Threonine protein kinases recognized from genomes of prokaryotes have been used to develop a classification framework for prokaryotic Ser/Thr protein kinases.

Methodology/Principal Findings: We have used traditional sequence alignment and phylogenetic approaches and clustered the prokaryotic kinases which represent 72 subfamilies with at least 4 members in each. Such a clustering enables classification of prokaryotic Ser/Thr kinases and it can be used as a framework to classify newly identified prokaryotic Ser/Thr kinases. After series of searches in a comprehensive sequence database we recognized that 38 subfamilies of prokaryotic protein kinases are associated to a specific taxonomic level. For example 4, 6 and 3 subfamilies have been identified that are currently specific to phylum proteobacteria, cyanobacteria and actinobacteria respectively. Similarly subfamilies which are specific to an order, sub-order, class, family and genus have also been identified. In addition to these, we also identify organism-diverse subfamilies. Members of these clusters are from organisms of different taxonomic levels, such as archaea, bacteria, eukaryotes and viruses.

Conclusion/Significance: Interestingly, occurrence of several taxonomic level specific subfamilies of prokaryotic kinases contrasts with classification of eukaryotic protein kinases in which most of the popular subfamilies of eukaryotic protein kinases occur diversely in several eukaryotes. Many prokaryotic Ser/Thr kinases exhibit a wide variety of modular organization which indicates a degree of complexity and protein-protein interactions in the signaling pathways in these microbes.

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Introduction

Archaea and eubacteria respond to a wide array of environmental stimuli including alterations in nutrient availability, temperature, osmolarity and host proximity. The primary sensing machinery present in eubacteria and archaea, which senses temperature, light, chemical concentration, viscosity, osmolarity etc, is the Two-component system. Serine/Threonine or Tyrosine phosphorylation by prokaryotic Ser/Thr or Tyrosine protein kinases is also recognized as another signaling mechanism in prokaryotes [1,2,3,4,5,6,7,8,9]. The well known signal transduction systems in prokaryotes are: i) the Two-component system [10] also referred as His-Asp phosphorelay system consisting of a sensor protein (histidine kinase) and response regulator [11,12] ii) the phosphoenolpyruvate transferase system [13] iii) the bacterial Tyrosine kinase system [7] and iv) the eukaryotic Ser/Thr or tyrosine kinase-like system [14]. The main focus of the current

study is detailed analysis of prokaryotic Serine/Threonine protein kinases involving clustering on the basis of amino acid sequence similarity of catalytic regions leading to identification of subfamilies.

Pkn1 from *Myxococcus xanthus* (*M. xanthus*) was the first prokaryotic Serine/Threonine protein kinase identified in prokaryotes which is known to autophosphorylate serine when incubated with radiolabelled ATP (Adenosine Tri Phosphate) [15]. Later phosphotransfer to an exogenous protein substrate by prokaryotic Serine/Threonine protein kinase, AfsK from *Streptomyces coelicolor* was demonstrated [16]. Sequencing of eubacterial and archaeal genomes later facilitated identification of prokaryotic Serine/Threonine protein kinases encoded in various eubacterial and archaeal genomes [3,5,15,17,18,19,20,21,22,23,24] which suggested that phosphorylation-dephosphorylation of proteins on hydroxyl group plays important roles in prokaryotes also. This mechanism of regulation is quite ancient than previously assumed

suggesting high complexity of the system. Prokaryotic Serine/Threonine protein kinases identified in eubacteria and archaea exhibit moderate to low sequence similarities to their eukaryotic counterparts. Recently, analysis of 619 prokaryotic genomes revealed presence of Serine/Threonine protein kinases in approximately two-thirds of the prokaryotes analysed [22].

Biological roles of some of the Ser/Thr kinases in prokaryotes have been investigated by laboratory experiments. Several Ser/Thr kinases from prokaryotes have been characterized biochemically which has revealed roles of these kinases in sugar transport [25], adaptation to light [26], flagellin phosphorylation and export [27], aggregation and sporulation [28], cell division and differentiation [29], morphological differentiation, secondary metabolism [30], oxidative stress response [31], sporulation and biofilm formation [32], glucose metabolism and glycogen consumption [33,34], carbon catabolite repression [35], glucose transport and cell division [36], purine biosynthesis [37]. Ser/Thr kinases also play crucial role in the virulence of pathogenic prokaryotes [19,38,39,40].

Little was known about the structures and functions of the prokaryotic Serine/Threonine protein kinases until the three-dimensional crystal structure of *Mycobacterium tuberculosis* protein, PknB in complex with a nucleotide triphosphate analog was solved [41,42] which revealed that tertiary structure of PknB has similarity to eukaryotic Ser/Thr and Tyr kinases (STYKs). This study supports a universal activation mechanism of Ser/Thr kinases in prokaryotes and eukaryotes [41,42]. Recently, crystal structure of one more prokaryotic protein kinase, YihE from *Escherichia coli* [43] has been solved which has significant similarity with choline kinase which shares the fold with eukaryotic STYKs. YihE is a protein of Cpx signaling system of *Escherichia coli* and *Salmonella enterica* which senses extra-cytoplasmic stress, and further, controls expression of factors that allow bacteria to adapt to stress. Functional analysis revealed that this protein kinase is most abundant in stationary phase and is important for long-term cell survival. The autophosphorylation and phosphorylation of protein substrates at Ser/Thr residues *in vitro* by YihE has been observed suggesting that it is a novel Ser/Thr kinase in prokaryotic cells [43].

Database of Kinases in Genomes (KinG) version 1.5 (<http://hodgkin.mbu.iisc.ernet.in/~king>) contains information on protein kinases encoded in hundreds of genomes and is updated periodically. Current release of KinG contains information on kinases from 47 archaeal and 256 eubacterial organisms. Sequence analysis of these prokaryotic Ser/Thr kinases depicts that these are distantly related to eukaryotic protein kinase superfamily [44]. Apart from KinG, SENTRA [45,46,47] which is a manually curated database provides information on signal transduction proteins. SENTRA has information for Two-component histidine kinases and response regulators, Serine/Threonine protein kinases and protein phosphatases, as well as adenylate and diguanylate cyclases and c-di-GMP phosphodiesterases from 202 completely sequenced prokaryotic genomes. However the present work is confined only to Ser/Thr kinases of prokaryotes.

With the advent of genome sequencing projects, several prokaryotic protein kinases have been identified and many more prokaryotic kinases are likely to be identified in the future. From the genome analysis of prokaryotic kinases and from the work described in KinG database, it is clear that many prokaryotic kinases can not be classified into one of the known groups or subfamilies of eukaryotic protein kinases originally defined by Hanks et al [44]. Hence, there is a need for the classification of prokaryotic Ser/Thr protein kinases as there is no classification

scheme available for these kinases similar to the classification framework proposed by Hanks et al [44] for eukaryotic protein kinases. In the present analysis, an attempt has been made to develop a classification scheme for prokaryotic Ser/Thr kinases and to study these kinases further for their potential biological roles based on tethered domains.

Results and Discussion

Classification of prokaryotic serine/threonine protein kinases

Based upon sequence similarity over the length of kinase domains (File S1), 993 Ser/Thr kinases from 303 prokaryotic genomes have been clustered into 270 clusters (File S2) with minimum sequence identity of 40% within a cluster. The members of these 270 clusters (sub-families) are quite divergent from each other as can be seen from multiple sequence alignment which is presented in File S3. Seventy two clusters out of total 270 clusters, which have four or more members, have been used for the classification of prokaryotic protein kinases into various sub-families based on their common properties. It should be noted that the sole consideration for the classification of these prokaryotic protein kinases into various families is the sequence similarity in the catalytic kinase domain region as this has been proved to be a good indicator in the classification of eukaryotic protein kinases [44].

In this analysis, there are different types of sub-families which have been observed based on common feature/s shared by the members within a cluster:

I) Sub-families which show specificity or predominance at taxonomic level

- a) Phylum specific/predominant subfamilies
- b) Order specific/predominant subfamilies
- c) Sub-order specific/predominant subfamilies
- d) Class specific/predominant subfamilies
- e) Family specific/predominant subfamilies
- f) Genus specific/predominant subfamilies

II) Subfamilies which show organism diversity

However these initially derived taxonomic specific/predominant clusters are only tentative as they have been derived from a limited dataset of 303 prokaryotic organisms. So, each of these (tentative) taxonomy specific or predominant clusters were probed, using PSI_BLAST, in the Uniref90 sequence database which is a comprehensive collection of protein sequences from diverse organisms. A tentative taxonomy-specific cluster is confirmed only in case the close homologues (indicated by $\geq 40\%$ sequence identity for catalytic kinase region) from Uniref90 are in the same taxonomic classification as the cluster in question.

Thus, finally, 38 subfamilies of prokaryotic Ser/Thr kinases have been identified which are specific at certain taxonomic level. For example, genus specific cluster contains members and close homologues from a particular genus of the eubacteria or archaea and so on. Table 1 lists these 38 subfamilies. Subfamilies which are organism diverse, share high sequence similarities with homologues from various organisms belonging to different taxonomic levels such as bacteria, eukaryotes, viruses and archaea.

A dendrogram depicting protein kinases specific to certain taxonomic levels are represented schematically in figure 1. The distance matrices representing extent of sequence dissimilarities at the catalytic kinase domains in every cluster are provided in File S4.

Table 1. Brief description of 38 subfamilies of prokaryotic Ser/Thr kinases which show specificity/predominance at certain taxonomic level.

S. No.	Cluster number(s)	Specificity/predominance at taxonomic level
1	6, 23, 38, 63	Phylum Proteobacteria specific
2	3, 4, 25, 29, 44, 59, 62	Phylum Proteobacteria predominant
3	15, 19, 24, 37, 54, 67	Phylum Cyanobacteria specific
4	60	Phylum Cyanobacteria predominant
5	10, 14, 26	Phylum Actinobacteria predominant
6	32	Phylum Euryarchaeota predominant
7	5	Class Gammaproteobacteria specific
8	8, 27, 34, 43, 46	Order Actinomycetales specific
9	22	Order Actinomycetales predominant
10	49	Order Chlamydiales specific
11	31, 72	Suborder Cystobacterineae specific
12	28	Family Thermoproteaceae specific
13	7	Genus Sulfolobus specific
14	45	Genus Bacillus and Geobacillus specific
15	52, 56	Genus Metallosphaera and Sulfolobus specific
16	48	Genus Chlamydia and Chlamydophila specific

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1a) Phylum specific subfamilies of prokaryotic Serine/Threonine protein kinases

In our analysis we find certain subfamilies which are phylum specific. There are four clusters (numbered as 6, 23, 38, 63), with members exclusively from phylum Proteobacteria. Bacteria belonging to this phylum are Gram-negative phototrophic and heterotrophic which are generally referred as “purple bacteria and their relatives” [48]. There are 7 other clusters (numbered as 3, 4, 25, 29, 44, 59 and 62) with members predominantly from the same phylum. Clusters 62 and 3 contain substantial representation of members from *Myxococcus xanthus*. *M. xanthus* whose developmental cycle and multicellular morphogenesis resemble those of eukaryotic slime molds such as *Dictyostelium discoideum* is reported to contain a large family of Ser/Thr kinases [49,50]. Transmembrane protein kinase pkn6 from *Myxococcus Xanthus* shows close similarity with a predominantly proteobacterial cluster. This kinase is expressed constitutively and is responsible for growth and development of the bacterium and it also has been speculated to sense the external signals for developmental process [51].

We also report here Tyrosine-protein kinase masK which happens to be a member of cluster 62 which belongs predominantly to phylum Proteobacteria. Protein kinase masK interacts with GTPase MglA to control social gliding motility of bacterium. masK is also reported to be essential for growth of the bacterium [52].

Another category in phylum specific clusters belongs to phylum Cyanobacteria. This phylum comprises oxygenic photosynthetic prokaryotes [53]. We report here 6 clusters (numbered as 15, 19, 24, 37, 54 and 67) members exclusively from Cyanobacteria. In addition, another cluster (numbered as 60), contains members mainly from cyanobacterial species.

Synechocystis sp. protein Ser/Thr kinases, spkA belonging to cluster 60 and spkB from very specific cyanobacterial cluster 54 are required for the normal motility of this unicellular cyanobacterium [54,55]. Another protein kinase spkD from same organism

is apparently essential for survival and protein spkF from *Synechocystis sp.* which is member of cyanobacteria specific cluster 67 has also been reported as a functional eukaryotic-like protein kinase [56].

Filamentous cyanobacterium *Anabaena* has differentiated cells called heterocysts which have a specialized function of nitrogen fixation. Protein kinase pknA has been shown to be important for normal cellular growth as disrupted pknA leads to formation of light green and rough colonies when adequate amount of nitrogen is not supplied. Moreover, a mutant results in formation of lower number of heterocysts as compared to wild-type filaments [57].

There are three clusters (numbered as 10, 14 and 26) with members predominantly from phylum Actinobacteria. This phylum comprises of Gram-positive bacteria with generally high G+C content in their DNA [58]. The phylum includes pathogens (*Mycobacterium spp.*, *Nocardia spp.*, *Corynebacterium spp.*, and *Propionibacterium spp.*), nitrogen fixing symbionts (*Frankia*).

Cluster number 32 has members predominantly from phylum Euryarchaeota of archaea, which comprises of methane producing methanogens and their phenotypically diverse relatives [59]. However, this subfamily members share close similarity with some of the kinases from eubacteria and hence this subfamily has members both from eubacteria and archaea.

1b) Order specific subfamilies of prokaryotic Serine/Threonine protein kinases

There are six clusters which are Order specific. Cluster 49 is Chlamydiales specific, member organisms of which are exclusively obligate intracellular parasite. For example, *Chlamydia trachomatis* belonging to same order causes trachoma which leads to blindness and sexually transmitted disease in human beings [60,61,62,63]. Other members include *Chlamydia pneumoniae* which causes pneumonia and bronchitis in human beings [64].

Serine/threonine-protein kinase pknD from bacterium *Chlamydia trachomatis* (Chlamydiales specific cluster 49) is a functional kinase and is expressed at early mid-phase of developmental cycle. This protein also has been predicted to have transmembrane domain and might serve as a receptor to sense environmental stimuli to regulate cellular functions. Protein kinase, pknD has been shown to interact with another protein kinase pkn1. All these factors may help the pathogen to exploit the host signaling pathways and supporting its own growth [65].

Other 5 clusters are exclusively Actinomycetales specific. Members of cluster number 22 predominantly contain homologues from order Actinomycetales. One of the members of this order is *Arthrobacter aurescens* which is a soil dwelling aerobe capable of surviving in extreme conditions like starvation, temperature changes, ionizing radiation, oxygen radicals, and toxic chemicals etc and also has the ability to degrade pollutants [66]. Other members from genus *Corynebacterium* belonging to same order are *Corynebacterium diphtheriae* which produces diphtheria toxin and causes the symptoms of diphtheria [67], *Corynebacterium urealyticum* which causes urinary tract infection [68], multiresistant nosocomial pathogen *Corynebacterium jeikeium* [69], *Mycobacterium abscessus* which causes skin, soft tissue and pulmonary infections [70], *Mycobacterium tuberculosis* causing tuberculosis [71] and an unculturable obligate pathogen *Mycobacterium leprae* which is responsible for causing leprosy [72]. Soil dwelling bacteria corresponding to genus *Streptomyces* also belongs to Order Actinomycetales which produces over two-thirds of naturally derived antibiotics [73]. Ser/thr protein kinase afsK (member of Actinomycetales specific cluster) is reported to phosphorylate AfsR, a transcription factor which is involved in regulation of production of secondary metabolites such as actinorhodin and undecylprodigiosin in *Streptomyces coelicolor* [16]. It also has been shown to regulate aerial mycelium

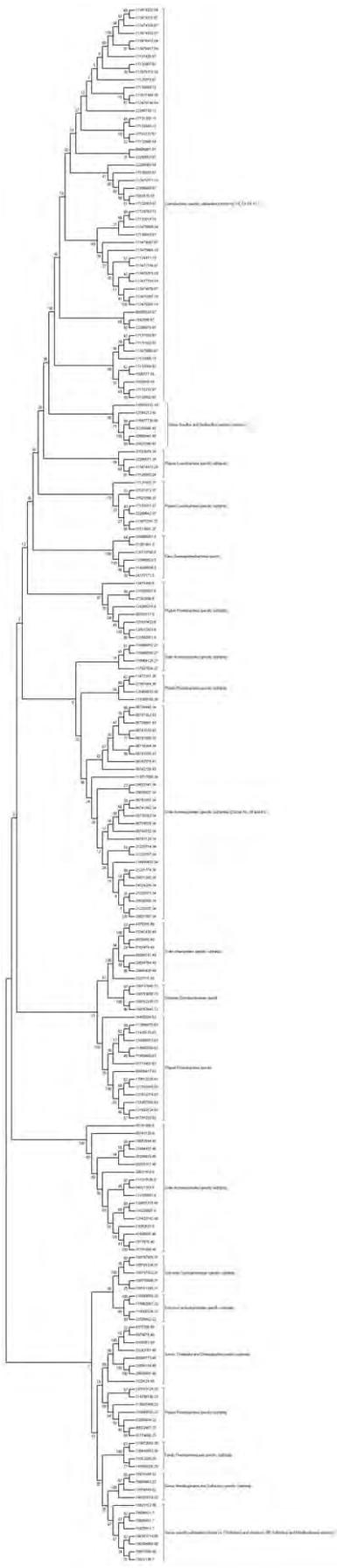


Figure 1. Dendrogram representation of prokaryotic protein kinase catalytic domains which are specific to certain taxonomic levels. Bootstrap values are provided at the main branches. Each protein kinase is represented as GI number and serial number of that particular cluster after dot. Taxonomic specificity of cluster is written at right hand side. For example, 120553124.23 represents GI number 120553124, it belongs to cluster number 23, which is a phylum Proteobacteria specific cluster and bootstrap value for the branch which encompasses this cluster is 99.

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formation and spore formation, thus morphological differentiation in *S. griseus* [74].

Protein kinase, pknG from organism *Mycobacterium tuberculosis* (Actinomycetales specific cluster 46) has been shown to regulate glutamate/glutamine level in the cell as pknG deletion results in accumulation of these amino acids and is also important for the growth of bacterium [75]. Another study suggests that Mycobacterial pknG is secreted in macrophage phagosome and inhibits phagosome-lysosome fusion and thus enable the pathogen to survive in the host cell [76].

lc) Sub-order specific subfamilies of prokaryotic Serine/Threonine protein kinases

There are two clusters under this category (numbered as 31, 72). Based on the analysis of our main dataset of kinases these two clusters are comprised of members only from *Myxococcus xanthus*. But when kinase sequences from this subfamily were searched in a large sequence database Uniref90 (see Materials and methods section) close homologues, which can be considered to be members of this subfamily, were identified from prokaryotes *Angiococcus disciformis*, *Myxococcus xanthus* and *Stigmatella aurantiaca*. Interestingly all of these organisms belong to sub-order Cystobacterineae of order Myxococcales under phylum Proteobacteria. *Angiococcus disciformis* and *Stigmatella aurantiaca* produce antibiotics angiolam A and stigmatellin, respectively [77,78]. Importance of *Myxococcus* is described in phylum specific cluster and organism diverse cluster section.

ld) Class specific subfamily of prokaryotic Serine/Threonine protein kinases

We report here a class specific cluster. Cluster number 5 is Gammaproteobacteria specific. Members are included from genus *Shewanella* (bacteria belonging to this genus are exclusively marine) such as metal reducing *Shewanella amazonensis* [79], denitrifying *Shewanella denitrificans* [80] and respiratory luminous bacterium *Shewanella woodyi* [81]. Other members come from genus *Marinobacter*, *Pseudoalteromonas* and *Alteromonas*.

le) Family specific subfamily of prokaryotic Serine/Threonine protein kinases

This subset corresponds to family Thermoproteaceae under archaea (Cluster number: 28). This family is characterized by hyperthermophilic archaeans. This includes members from genus *Pyrobaculum* such as *Pyrobaculum aerophilum*, *Pyrobaculum arsenaticum*, *Pyrobaculum calidifontis*, *Pyrobaculum islandicum* and also from *Thermoproteus tenax*. Protein kinases falling into this category are all single domain proteins.

lf) Genus specific subfamilies of prokaryotic Serine/Threonine protein kinases

There are five clusters (numbered as 7, 45, 48, 52 and 56) under this category. Cluster 7 is specific to genus *Sulfolobus* from archaea. Genus *Sulfolobus* represents sulfur-oxidizing microorganisms living

at low pH and high temperature. Different thermoacidophilic species belonging to this genus are *S. acidocaldarius* [82], *S. solfataricus* [83] and *S. Tokodaii*. Protein phosphorylation studies have been carried out and Ser/Thr protein kinase has been reported from this archaeon [23,84]. Protein kinase domains appearing in this cluster are not tethered with any other domain.

Clusters 52 and 56 are specific to genus Metallosphaera and Sulfolobus from family sulfolobaceae under archaea. Genus Metallosphaera comprises aerobic, metal-mobilizing, thermoacidophilic microorganisms [85].

One of the clusters (No. 45) is specific to genus *Bacillus* and *Geobacillus* of family bacillaceae which comprises of all Gram-positive bacteria. Interaction of phosphatase, BA-Stop1 with Ser/Thr protein kinase BA-Stk1 from *Bacillus anthracis*, has been shown to be responsible for the virulence of the bacterium [86].

Above mentioned clusters in this section have members which correspond to eukaryotic like protein kinase (epk) mentioned by Kannan et al [6].

Another cluster (numbered 48) is specific to genus *Chlamydophila* and *Chlamydia* from family chlamydiaceae. Genus Chlamydia comprises bacteria which are intracellular obligate parasites and causes diseases such as sexually transmitted diseases, blindness, pneumonia and bronchitis as mentioned above. A member of this cluster is present in pknB (bacterial specific) subfamily according to Kannan et al [6].

II) Organism diverse subfamilies of prokaryotic Serine/Threonine protein kinases

There are 34 clusters falling under this category. Members of these clusters show close similarity to members from diverse organisms. Search, in UniRef90, for closely related kinases of one of the clusters (number 68) resulted in recognition of serine/threonine-protein kinase pkn1 of Gram-negative soil bacterium *Myxococcus xanthus*. This protein kinase is required for normal development of this bacterium and deletion of pkn1 gene results in premature differentiation and poor spore formation [87].

Cluster number 71 has pknD, a protein kinase member from *Mycobacterium tuberculosis*. It has extracellular highly symmetric six-bladed β -propeller structure which could bind a multivalent ligand and can act as a sensor domain [88]. PknD has been reported to phosphorylate MmpL7 which is associated with the formation of cell wall of bacterium and serves as virulence factor [89]. Studies also suggest role of this class of kinase in regulation of transcription of numerous genes in bacterium [90]. Same cluster has pknF, a protein kinase from *M. tuberculosis* which interacts with ABC transporter containing a Forkhead-associated domain to play role in virulence and cell growth of bacterium [91]. PknF involvement has been suggested in glucose uptake, cellular growth and septum formation also [36]. PknE, a transmembrane protein kinase, which phosphorylates multiple FHA domain is also a member of organism diverse cluster 61 [92].

PknH from *Mycobacterium tuberculosis*, is a member of a subfamily (cluster number: 71) and it has been shown to phosphorylate EmbR, which is mediated by FHA (forkhead-associated) domain [93]. Protein EmbR is associated with regulation of activity of enzyme arabinosyltransferase involved in arabinan biosynthesis of arabinogalactan which is an important molecule of the *Mycobacterial* cell wall. PknH deletion also leads to survival and higher bacillary loads in BALB/c mice, suggesting a role of the protein in regulating the growth profile of the bacterium [94].

A member of cluster number 17 protein kinase pknB from *Mycobacterium tuberculosis* phosphorylates PBPA, a penicillin binding protein and regulates the growth and cell division of the bacterium [95].

Serine/threonine-protein kinase prkC from Gram-positive bacterium *Bacillus subtilis* is a member of organism diverse cluster (cluster 33). PrkC participates in developmental processes like spore formation and biofilm formation as mutant of this gene show decreased efficiency of both the processes [32].

Members of cluster number 35 show high similarity to StoPK-1, serine/threonine protein kinase from *Streptomyces toyocaensis*. Disruption of StoPK-1 leads to unusual mycelium morphology. StoPK-1 is also associated with signal transduction pathways which are sensitive to oxidative stress as inactivation of same gene results in increased sensitivity towards oxygen radical-generating compound [31].

Clusters 11 and 55 have protein kinases pknA and pknB respectively from soil-borne, non-pathogenic Gram-positive bacterium *Corynebacterium glutamicum*, which is used for production of L-lysine and L-glutamic acid on commercial scale. These kinases are absolutely essential for *Corynebacterium* growth. Partial depletion of both kinases results in defect in cell division and formation of elongated cell [96].

Cluster 13 has transmembrane protein Ser/Thr kinase pkn2 from *Myxococcus xanthus* which has been shown to phosphorylate beta-lactamase and restrict its secretion across the membrane in *E. coli*. Enzyme beta lactamase are produced by certain bacteria and are responsible for bacterial resistance against beta-lactam antibiotics such as penicillins. Thus pkn2 is speculated to regulate the activity of penicillin binding proteins. Disruption of pkn2 also results in low yield of myxospores [97].

There are two organism diverse subfamilies (numbered 30 and 9) with members from both archaea and eubacteria. However most of the members in these two clusters are from eubacteria with only a minor representation from archaeal organisms (~1% and ~2% in subfamilies 30 and 9 respectively).

Domains associated with prokaryotic Ser/Thr protein kinases

It is well known that many of the prokaryotic protein kinases are not multi-modular in nature. However, there are few prokaryotic Ser/Thr kinases identified which have other domains tethered to the protein kinase domain which adds complexity to the type of function they are performing. Domain architectures of all the prokaryotic kinases in each of the 72 clusters are provided in File S5.

The most commonly tethered domains to the protein kinase domain are Tetratricopeptide (TPR) repeats, PASTA, WD40 repeats, GAF, PD40 repeats and APH domains. TPR repeats are involved in variety of functions such as extensive protein-protein interaction in the assembly of multiprotein complexes [98]. WD40 repeats containing proteins are involved in wide range of functions like signal transduction, RNA processing, gene regulation and regulation of cell cycle [99]. WD40 repeats help in coordinating multi-protein complex assemblies, where the repeating units of WD40 serve as a rigid scaffold for protein-protein interactions [100]. PASTA domain is found in both archaea and bacteria, occurs at C-terminus of several penicillin-binding proteins and bacterial serine/threonine kinases [101]. While the GAF domain is known to participate in photo transduction in plants and vertebrates [102], it has been reported to have role in change of pigment-protein composition according to light color changes in cyanobacteria [103]. This domain has also been speculated to participate in regulation of various signalling events in non-photosynthetic bacteria. PD40 protein domain family is related to WD40 domain family and is a cell surface protein [104]. Another most commonly tethered domain is APH (aminoglycoside phosphotransferase) domain. Aminoglycoside phosphotransferases

are proteins which inactivate aminoglycoside antibiotic substrate by phosphorylating the same in prokaryotes [105].

Domains which are generally tethered to Ser/Thr kinases specific to Proteobacteria are SAF domain which is found in antifreeze proteins, flagellar proteins and pilus proteins [106], Universal stress protein (Usp) family which is expressed in response to stress agents in bacteria [107] and APH domain which is reported to inactivate the antibiotics in prokaryotes.

Domains generally tethered to Cyanobacteria specific Ser/Thr kinase are Pentapeptide repeats which are most commonly found in Cyanobacteria and speculated to be involved in Cyanobacteria-specific metabolism [108], WD40 repeats, APH domain, TPR repeats, CHASE2, which is an extracellular sensory domain present in various classes of transmembrane receptors that takes part in signal transduction pathways in bacteria and archaea [109], SH3_3 (src Homology-3) domain which is involved in signal transduction and cytoskeletal organization [110] and FHA (forkhead-associated) domain which is a phosphopeptide binding motif [111].

In addition to supporting information files with this paper details of prokaryotic Ser/Thr kinases identified in this study can be found at KinG database (<http://hodgkin.mbu.iisc.ernet.in/~king/>) in the link “A Framework for Classification of Prokaryotic Protein Kinases”.

Materials and Methods

From 303 prokaryotic genomes, 993 non redundant eubacterial and archaeal Ser/Thr protein kinases have been retrieved from KinG (database version 1.5) [112]. Briefly, protein kinases are identified using a combination of profile-based search methods such as PSI-BLAST [113] and RPS-BLAST [114] using multiple profiles (MulPSSM) [115,116] and HMMER search [117], which have been previously benchmarked and has been used in our earlier kinome analysis for several other genomes [118,119,120,121].

Multiple Position Specific Scoring Matrices (MulPSSM) from 2810 sequence profiles have been generated from different groups of kinases as mentioned in www.kinase.com. In case of single profile approach a reference sequence is chosen arbitrarily for building a PSSM and the query sequence is searched in database of PSSMs of various protein families. But in the case of multiple profile approach every sequence from a given multiple sequence alignment of a protein domain family is used for building PSSMs which increases the search space as well as removes bias toward the reference sequence. Protein sequences from prokaryotic genomes have been searched in database of Multiple PSSMs using RPS-BLAST. Conditions for hit in RPS-BLAST searches include an e-value cut-off of 10^{-4} and more than 70% of profile should be covered by the query in the alignment. Ser/Thr kinases have also been identified using HMMER against Pfam [122] (release 23) protein kinase (Pfam code: PF00069) profiles. E-value cut off used in HMM search is 0.01. Amino acid sequences from prokaryotes have been searched, using PSI-BLAST, in a database of kinases procured from Pfam. E-value cut-off used in this method is 0.0001. Query should cover greater than or equal to 70% length of the sequence in the database in the alignment for considering the database entry as a hit.

CD-HIT [123], a program for clustering large protein database at specific sequence identity threshold has been used to make the prokaryotic protein kinase domain dataset non-redundant at the sequence identity cut off 40%. Hence no two sequences have more than 40% sequence identity to each other across any two clusters. The number of clusters generated by the CD-HIT program is 270. There are 126 clusters with only one member in each suggesting

their high sequence divergence. There are 72 clusters which have four or more members in each cluster. These 72 clusters are considered as prominent subfamilies of prokaryotic Ser/Thr kinases.

A randomly chosen member from each of these 72 clusters was searched, using PSI-BLAST, in Uniref90 (www.ebi.ac.uk/uniref/) dataset which is a comprehensive collection of amino acid sequences of non-redundant proteins. Uniref90 represents the best current knowledge on amino acid sequences from diverse organisms. Our kinase dataset is derived from KinG (version 1.5) and has information for only for 303 prokaryotic genomes. However to classify any cluster as, for example, phylum Cyanobacteria specific, we have not only considered phylum of member organisms from that clusters, we also ensured by searching into comprehensive database Uniref90 that member of this cluster picks up homologues from Cyanobacteria phylum only. If proteins from other than Cyanobacteria are picked-up as close homologues of the query then the cluster concerned is not considered as Cyanobacteria specific. Close homologues have been identified with sequence identity of 40% or greater and greater than or equal to 70% query coverage. These conditions should be satisfied over and above the E-value cut-off of 0.00001.

Multiple sequence alignment program, CLUSTALW has been used to align kinase domain sequences from each of the 72 clusters [124]. The tree was generated using neighbor-joining (NJ) method [125]. NJ method provides topology as well as branch length of final tree. This method is based on principle of finding pairs of operational taxonomic units (OTUs), “neighbors” that minimizes the sum of branch lengths at each stage of clustering of OTUs. Tree is annotated with the bootstrap values (1000 iterations).

MEGA program (version 4.0) has been used to draw the tree [126]. Many of the prokaryotic protein kinases are not multi-modular in nature but some of them have domains tethered to the protein kinase domains. The domain architectures of these prokaryotic Ser/Thr kinases have been identified on the basis of searches using HMMER [117] against the Pfam (release 23) profiles [122] containing 10340 families. E-value cut -off used in this search is 0.01.

Conclusions

The present study involving identification and analysis of Ser/Thr kinases in prokaryotic genomes has provided insights into signal transduction and metabolic processes in prokaryotes. The extensive dataset of prokaryotic kinases obtained from KinG has given us the opportunity to classify these kinases into different categories based upon their occurrence in particular taxonomic group.

Specificity of Protein Ser/Thr kinases at particular taxonomic level suggest requirement of these Ser/Thr protein kinases for certain specific function which is lineage specific and not needed for all the prokaryotes. It is interesting to note that occurrence of several taxonomic specific subfamilies of prokaryotic kinases contrasts with classification of eukaryotic protein kinases in which most of the popular subfamilies of eukaryotic protein kinases occur diversely in several eukaryotes. Clusters representing prokaryotic protein kinase subfamilies which are taxonomic level specific suggest role of these Ser/Thr protein kinases in some specific function being carried out by limited sets of prokaryotes. Finally, organism diverse subfamilies of prokaryotes suggests wide spread occurrence of such Ser/Thr kinases. Almost 50% of the clusters obtained in this analysis have only one member suggesting their sequence and, probably, functional divergence. Genomic data of many more prokaryotes is not yet available. With the completion of genome sequencing of many more prokaryotes, some of these

clusters may have additional members. Ongoing efforts are directed towards development of profiles of clusters in the present classification scheme for prokaryotic Ser/Thr protein kinases. This should allow convenient classification of prokaryotic Ser/Thr kinases in the future.

Supporting Information

File S1 Amino acid sequences of prokaryotic protein kinases (catalytic domain).

Found at: doi:10.1371/journal.pone.0010608.s001 (0.29 MB TXT)

File S2 Taxonomic level specificity details of clusters.

Found at: doi:10.1371/journal.pone.0010608.s002 (0.08 MB XLS)

References

- Leonard CJ, Aravind L, Koonin EV (1998) Novel families of putative protein kinases in bacteria and archaea: evolution of the “eukaryotic” protein kinase superfamily. *Genome Res* 8: 1038–1047.
- Han G, Zhang CC (2001) On the origin of Ser/Thr kinases in a prokaryote. *FEMS Microbiol Lett* 200: 79–84.
- Kennelly PJ (2002) Protein kinases and protein phosphatases in prokaryotes: a genomic perspective. *FEMS Microbiol Lett* 206: 1–8.
- Krupa A, Srinivasan N (2002) Lipopolysaccharide phosphorylating enzymes encoded in the genomes of Gram-negative bacteria are related to the eukaryotic protein kinases. *Protein Sci* 11: 1580–1584.
- Krupa A, Srinivasan N (2005) Diversity in domain architectures of Ser/Thr kinases and their homologues in prokaryotes. *BMC Genomics* 6: 129.
- Kannan N, Taylor SS, Zhai Y, Venter JC, Manning G (2007) Structural and functional diversity of the microbial kinome. *PLoS Biol* 5: e17.
- Grangeasse C, Cozzone AJ, Deutscher J, Mijakovic I (2007) Tyrosine phosphorylation: an emerging regulatory device of bacterial physiology. *Trends Biochem Sci* 32: 86–94.
- Jadeau F, Bechet E, Cozzone AJ, Deleage G, Grangeasse C, et al. (2008) Identification of the idiosyncratic bacterial protein tyrosine kinase (BY-kinase) family signature. *Bioinformatics* 24: 2427–2430.
- Bach H, Wong D, Av-Gay Y (2009) Mycobacterium tuberculosis PtkA is a novel protein tyrosine kinase whose substrate is PtpA. *Biochem J* 420: 155–160.
- Bourret RB, Hess JF, Borkovich KA, Pakula AA, Simon MI (1989) Protein phosphorylation in chemotaxis and two-component regulatory systems of bacteria. *J Biol Chem* 264: 7085–7088.
- Bourret RB, Borkovich KA, Simon MI (1991) Signal transduction pathways involving protein phosphorylation in prokaryotes. *Annu Rev Biochem* 60: 401–441.
- Hoch JA (2000) Two-component and phosphorelay signal transduction. *Curr Opin Microbiol* 3: 165–170.
- Reizer J, Saier MH, Jr., Deutscher J, Grenier F, Thompson J, et al. (1988) The phosphoenolpyruvate:sugar phosphotransferase system in gram-positive bacteria: properties, mechanism, and regulation. *Crit Rev Microbiol* 15: 297–338.
- Bakal CJ, Davies JE (2000) No longer an exclusive club: eukaryotic signalling domains in bacteria. *Trends Cell Biol* 10: 32–38.
- Munoz-Dorado J, Inouye S, Inouye M (1993) Eukaryotic-like protein serine/threonine kinases in *Myxococcus xanthus*, a developmental bacterium exhibiting social behavior. *J Cell Biochem* 51: 29–33.
- Matsumoto A, Hong SK, Ishizuka H, Horinouchi S, Beppu T (1994) Phosphorylation of the AfsR protein involved in secondary metabolism in Streptomyces species by a eukaryotic-type protein kinase. *Gene* 146: 47–56.
- Kennelly PJ, Potts M (1996) Fancy meeting you here! A fresh look at “prokaryotic” protein phosphorylation. *J Bacteriol* 178: 4759–4764.
- Zhang CC, Gonzalez L, Phalip V (1998) Survey, analysis and genetic organization of genes encoding eukaryotic-like signaling proteins on a cyanobacterial genome. *Nucleic Acids Res* 26: 3619–3625.
- Av-Gay Y, Everett M (2000) The eukaryotic-like Ser/Thr protein kinases of Mycobacterium tuberculosis. *Trends Microbiol* 8: 238–244.
- Petricekova K, Petricek M (2003) Eukaryotic-type protein kinases in Streptomyces coelicolor: variations on a common theme. *Microbiology* 149: 1609–1621.
- Wehenkel A, Bellinzoni M, Grana M, Duran R, Villarino A, et al. (2008) Mycobacterial Ser/Thr protein kinases and phosphatases: physiological roles and therapeutic potential. *Biochim Biophys Acta* 1784: 193–202.
- Perez J, Castaneda-Garcia A, Jenke-Kodama H, Muller R, Munoz-Dorado J (2008) Eukaryotic-like protein kinases in the prokaryotes and the myxobacterial kinome. *Proc Natl Acad Sci U S A* 105: 15950–15955.
- Skorko R (1984) Protein phosphorylation in the archaeabacterium *Sulfolobus acidocaldarius*. *Eur J Biochem* 145: 617–622.
- Smith RF, King KY (1995) Identification of a eukaryotic-like protein kinase gene in Archaeabacteria. *Protein Sci* 4: 126–129.
- Deutscher J, Saier MH, Jr. (1983) ATP-dependent protein kinase-catalyzed phosphorylation of a seryl residue in HPr, a phosphate carrier protein of the phosphotransferase system in *Streptococcus pyogenes*. *Proc Natl Acad Sci U S A* 80: 6790–6794.
- Warner KM, Bullerjahn GS (1994) Light-Dependent Tyrosine Phosphorylation in the Cyanobacterium *Prochlorothrix hollandica*. *Plant Physiol* 105: 629–633.
- South SL, Nichols R, Montic TC (1994) Tyrosine kinase activity in *Pseudomonas aeruginosa*. *Mol Microbiol* 12: 903–910.
- Frasch SC, Dworkin M (1996) Tyrosine phosphorylation in *Myxococcus xanthus*, a multicellular prokaryote. *J Bacteriol* 178: 4084–4088.
- Wu J, Ohta N, Zhao JL, Newton A (1999) A novel bacterial tyrosine kinase essential for cell division and differentiation. *Proc Natl Acad Sci U S A* 96: 13068–13073.
- Umezawa T, Lee PC, Horinouchi S (2002) Protein serine/threonine kinases in signal transduction for secondary metabolism and morphogenesis in Streptomyces. *Appl Microbiol Biotechnol* 59: 419–425.
- Neu JM, MacMillan SV, Nodwell JR, Wright GD (2002) StoPK-1, a serine/threonine protein kinase from the glycopeptide antibiotic producer Streptomyces toyaensis NRRL 15009, affects oxidative stress response. *Mol Microbiol* 44: 417–430.
- Madec E, Laszkiewicz A, Iwanicki A, Obuchowski M, Seror S (2002) Characterization of a membrane-linked Ser/Thr protein kinase in *Bacillus subtilis*, implicated in developmental processes. *Mol Microbiol* 46: 571–586.
- Nariya H, Inouye S (2002) Activation of 6-phosphofructokinase via phosphorylation by Pkn4, a protein Ser/Thr kinase of *Myxococcus xanthus*. *Mol Microbiol* 46: 1353–1366.
- Nariya H, Inouye S (2003) An effective sporulation of *Myxococcus xanthus* requires glycogen consumption via Pkn4-activated 6-phosphofructokinase. *Mol Microbiol* 49: 517–528.
- Ponct S, Mijakovic I, Nessler S, Gueguen-Chaignon V, Chaptal V, et al. (2004) HPr kinase/phosphylase, a Walker motif A-containing bifunctional sensor enzyme controlling catabolite repression in Gram-positive bacteria. *Biochim Biophys Acta* 1697: 123–135.
- Deol P, Vohra R, Saini AK, Singh A, Chandra H, et al. (2005) Role of *Mycobacterium tuberculosis* Ser/Thr kinase PknF: implications in glucose transport and cell division. *J Bacteriol* 187: 3415–3420.
- Rajagopal L, Vo A, Silvestroni A, Rubens CE (2005) Regulation of purine biosynthesis by a eukaryotic-type kinase in *Streptococcus agalactiae*. *Mol Microbiol* 56: 1329–1346.
- Galyov EE, Hakansson S, Forsberg A, Wolf-Watz H (1993) A secreted protein kinase of *Yersinia pseudotuberculosis* is an indispensable virulence determinant. *Nature* 361: 730–732.
- Rajagopal L, Vo A, Silvestroni A, Rubens CE (2006) Regulation of cytotoxin expression by converging eukaryotic-type and two-component signalling mechanisms in *Streptococcus agalactiae*. *Mol Microbiol* 62: 941–957.
- Wiley DJ, Nordfeldth R, Rosenzweig J, DaFonseca CJ, Gustin R, et al. (2006) The Ser/Thr kinase activity of the *Yersinia* protein kinase A (YpkA) is necessary for full virulence in the mouse, mollifying phagocytes, and disrupting the eukaryotic cytoskeleton. *Microb Pathog* 40: 234–243.
- Ortiz-Lombardia M, Pompeo F, Boitel B, Alzari PM (2003) Crystal structure of the catalytic domain of the PknB serine/threonine kinase from *Mycobacterium tuberculosis*. *J Biol Chem* 278: 13094–13100.
- Young TA, Delagoutte B, Endrizzi JA, Falick AM, Alber T (2003) Structure of *Mycobacterium tuberculosis* PknB supports a universal activation mechanism for Ser/Thr protein kinases. *Nat Struct Biol* 10: 168–174.
- Zheng J, He C, Singh VK, Martin NL, Jia Z (2007) Crystal structure of a novel prokaryotic Ser/Thr kinase and its implication in the Cpx stress response pathway. *Mol Microbiol* 63: 1360–1371.
- Hanks SK, Quinn AM, Hunter T (1988) The protein kinase family: conserved features and deduced phylogeny of the catalytic domains. *Science* 241: 42–52.

45. Maltsev N, Marland E, Yu GX, Bhatnagar S, Lusk R (2002) Sentra, a database of signal transduction proteins. *Nucleic Acids Res* 30: 349–350.

46. D'Souza M, Romine MF, Maltsev N (2000) SENTRA, a database of signal transduction proteins. *Nucleic Acids Res* 28: 335–336.

47. D'Souza M, Glass EM, Syed MH, Zhang Y, Rodriguez A, et al. (2007) Sentra: a database of signal transduction proteins for comparative genome analysis. *Nucleic Acids Res* 35: D271–273.

48. Stachebrandt RGMaHGT E (1988) Proteobacteria classis nov., a Name for the Phylogenetic Taxon That Includes the “Purple Bacteria and Their Relatives”. *Int J Syst Bacteriol* 38: 321–325.

49. Zhang W, Munoz-Dorado J, Inouye M, Inouye S (1992) Identification of a putative eukaryotic-like protein kinase family in the developmental bacterium *Myxococcus xanthus*. *J Bacteriol* 174: 5450–5453.

50. Munoz-Dorado J, Inouye S, Inouye M (1991) A gene encoding a protein serine/threonine kinase is required for normal development of *M. xanthus*, a gram-negative bacterium. *Cell* 67: 995–1006.

51. Zhang W, Inouye M, Inouye S (1996) Reciprocal regulation of the differentiation of *Myxococcus xanthus* by Pkn5 and Pkn6, eukaryotic-like Ser/Thr protein kinases. *Mol Microbiol* 20: 435–447.

52. Thomasson B, Link J, Stassinopoulos AG, Burke N, Plamann L, et al. (2002) MgLA, a small GTPase, interacts with a tyrosine kinase to control type IV pilus-mediated motility and development of *Myxococcus xanthus*. *Mol Microbiol* 46: 1399–1413.

53. Oren A (2004) A proposal for further integration of the cyanobacteria under the Bacteriological Code. *Int J Syst Evol Microbiol* 54: 1895–1902.

54. Kamei A, Yuasa T, Orikawa K, Geng XX, Ikeuchi M (2001) A eukaryotic-type protein kinase, SpkA, is required for normal motility of the unicellular Cyanobacterium *synechocystis* sp. strain PCC 6803. *J Bacteriol* 183: 1505–1510.

55. Kamei A, Yoshihara S, Yuasa T, Geng X, Ikeuchi M (2003) Biochemical and functional characterization of a eukaryotic-type protein kinase, SpkB, in the cyanobacterium, *Synechocystis* sp. PCC 6803. *Curr Microbiol* 46: 296–301.

56. Kamei A, Yuasa T, Geng X, Ikeuchi M (2002) Biochemical examination of the potential eukaryotic-type protein kinase genes in the complete genome of the unicellular Cyanobacterium *synechocystis* sp. PCC 6803. *DNA Res* 9: 71–78.

57. Zhang CC (1993) A gene encoding a protein related to eukaryotic protein kinases from the filamentous heterocystous cyanobacterium *Anabaena* PCC 7120. *Proc Natl Acad Sci U S A* 90: 11840–11844.

58. Ventura M, Canchaya C, Tauch A, Chandra G, Fitzgerald GF, et al. (2007) Genomics of Actinobacteria: tracing the evolutionary history of an ancient phylum. *Microbiol Mol Biol Rev* 71: 495–548.

59. Woese CR, Kandler O, Wheelis ML (1990) Towards a natural system of organisms: proposal for the domains Archaea, Bacteria, and Eucarya. *Proc Natl Acad Sci U S A* 87: 4576–4579.

60. Horn M, Collingro A, Schmitz-Esser S, Beier CL, Purkhold U, et al. (2004) Illuminating the evolutionary history of chlamydiae. *Science* 304: 728–730.

61. Kari L, Whitmire WM, Carlson JH, Crane DD, Reveneau N, et al. (2008) Pathogenic diversity among Chlamydia trachomatis ocular strains in nonhuman primates is affected by subtle genomic variations. *J Infect Dis* 197: 449–456.

62. Carlson JH, Porcella SF, McClarty G, Caldwell HD (2005) Comparative genomic analysis of Chlamydia trachomatis oculotropic and genitotropic strains. *Infect Immun* 73: 6407–6418.

63. Thomson NR, Holden MT, Carder C, Lennard N, Lockey SJ, et al. (2008) Chlamydia trachomatis: genome sequence analysis of lymphogranuloma venereum isolates. *Genome Res* 18: 161–171.

64. Kalman S, Mitchell W, Marathe R, Lammel C, Fan J, et al. (1999) Comparative genomes of Chlamydia pneumoniae and C. trachomatis. *Nat Genet* 21: 385–389.

65. Verma A, Maurelli AT (2003) Identification of two eukaryote-like serine/threonine kinases encoded by Chlamydia trachomatis serovar L2 and characterization of interacting partners of Pkn1. *Infect Immun* 71: 5772–5784.

66. Mongodin EF, Shapir N, Daugherty SC, DeBoy RT, Emerson JB, et al. (2006) Secrets of soil survival revealed by the genome sequence of *Arthrobacter aurescens* TC1. *PLoS Genet* 2: e214.

67. Cerdeno-Tarraga AM, Efstratiou A, Dover LG, Holden MT, Pallen M, et al. (2003) The complete genome sequence and analysis of *Corynebacterium diphtheriae* NCTC13129. *Nucleic Acids Res* 31: 6516–6523.

68. Tauch A, Tross E, Tilker A, Ludewig U, Schneiker S, et al. (2008) The lifestyle of *Corynebacterium urealyticum* derived from its complete genome sequence established by pyrosequencing. *J Biotechnol* 136: 11–21.

69. Tauch A, Kaiser O, Hain T, Goessmann A, Weisshaar B, et al. (2005) Complete genome sequence and analysis of the multiresistant nosocomial pathogen *Corynebacterium jeikeium* K411, a lipid-requiring bacterium of the human skin flora. *J Bacteriol* 187: 4671–4682.

70. Brown-Elliott BA, Wallace RJ, Jr. (2002) Clinical and taxonomic status of pathogenic nonpigmented or late-pigmenting rapidly growing mycobacteria. *Clin Microbiol Rev* 15: 716–746.

71. Cole ST, Brosch R, Parkhill J, Garnier T, Churcher C, et al. (1998) Deciphering the biology of *Mycobacterium tuberculosis* from the complete genome sequence. *Nature* 393: 537–544.

72. Monot M, Honore N, Garnier T, Zidane N, Sherifi D, et al. (2009) Comparative genomic and phylogeographic analysis of *Mycobacterium leprae*. *Nat Genet* 41: 1282–1289.

73. Bentley SD, Chater KF, Cerdeno-Tarraga AM, Challis GL, Thomson NR, et al. (2002) Complete genome sequence of the model actinomycete *Streptomyces coelicolor* A3(2). *Nature* 417: 141–147.

74. Ueda K, Umeyama T, Beppu T, Horinouchi S (1996) The aerial mycelium-defective phenotype of *Streptomyces griseus* resulting from A-factor deficiency is suppressed by a Ser/Thr kinase of *S. coelicolor* A3(2). *Gene* 169: 91–95.

75. Cowley S, Ko M, Pick N, Chow R, Downing KJ, et al. (2004) The *Mycobacterium tuberculosis* protein serine/threonine kinase PknG is linked to cellular glutamate/glutamine levels and is important for growth in vivo. *Mol Microbiol* 52: 1691–1702.

76. Walburger A, Koul A, Ferrari G, Nguyen L, Prescianotto-Baschong C, et al. (2004) Protein kinase G from pathogenic mycobacteria promotes survival within macrophages. *Science* 304: 1800–1804.

77. Kunze B, Kohl W, Hofle G, Reichenbach H (1985) Production, isolation, physico-chemical and biological properties of angiolam A, a new antibiotic from *Angiococcus disciformis* (Myxobacterales). *J Antibiot (Tokyo)* 38: 1649–1654.

78. Kunze B, Kemmer T, Hofle G, Reichenbach H (1984) Stigmatellin, a new antibiotic from *Stigmatella aurantiaca* (Myxobacterales). I. Production, physico-chemical and biological properties. *J Antibiot (Tokyo)* 37: 454–461.

79. Venkateswaran K, Dollhopf ME, Aller R, Stachebrandt E, Nealon KH (1998) *Shewanella amazonensis* sp. nov., a novel metal-reducing facultative anaerobe from Amazonian shell muds. *Int J Syst Bacteriol* 48 Pt 3: 965–972.

80. Brettar I, Christen R, Hofle MG (2002) *Shewanella denitrificans* sp. nov., a vigorously denitrifying bacterium isolated from the oxic-anoxic interface of the Gotland Deep in the central Baltic Sea. *Int J Syst Evol Microbiol* 52: 2211–2217.

81. Makemson JC, Fulayfil NR, Landry W, Van Ert LM, Wimpie CF, et al. (1997) *Shewanella woodyi* sp. nov., an exclusively respiratory luminous bacterium isolated from the Alboran Sea. *Int J Syst Bacteriol* 47: 1034–1039.

82. Chen L, Brugger K, Skovgaard M, Redder P, She Q, et al. (2005) The genome of *Sulfolobus acidocaldarius*, a model organism of the Crenarchaeota. *J Bacteriol* 187: 4992–4999.

83. Sensen CW, Charlebois RL, Chow C, Clausen IG, Curtis B, et al. (1998) Completing the sequence of the *Sulfolobus solfataricus* P2 genome. *Extremophiles* 2: 305–312.

84. Lower BH, Potters MB, Kennelly PJ (2004) A phosphoprotein from the archaeon *Sulfolobus solfataricus* with protein-serine/threonine kinase activity. *J Bacteriol* 186: 463–472.

85. Huber G, Spinnler C, Gambacorta A, Stetter KO (1989) *Metallosphaera sedula* gen. nov. and sp. nov. represents a new genus of aerobic, metal-mobilizing, thermoacidophilic archaeabacteria. *Syst Appl Microbiol* 12: 38–47.

86. Shakir SM, Bryant KM, Larabee JL, Hamm EE, Lovchik J, et al. (2009) Regulatory Interactions of a Virulence-Associated Serine/Threonine Phosphatase-Kinase Pair in *Bacillus anthracis*. *J Bacteriol*.

87. Munoz-Dorado J, Inouye S, Inouye M (1991) A gene encoding a protein serine/threonine kinase is required for normal development of *M. xanthus*, a gram-negative bacterium. *Cell* 67: 995–1006.

88. Good MC, Greenstein AE, Young TA, Ng HL, Alber T (2004) Sensor domain of the *Mycobacterium tuberculosis* receptor Ser/Thr protein kinase, PknD, forms a highly symmetric beta propeller. *J Mol Biol* 339: 459–469.

89. Perez J, Garcia R, Bach H, de Waard JH, Jacobs WR, Jr, et al. (2006) *Mycobacterium tuberculosis* transporter MmpL7 is a potential substrate for kinase PknD. *Biochem Biophys Res Commun* 348: 6–12.

90. Greenstein AE, MacGurn JA, Baer CE, Falick AM, Cox JS, et al. (2007) *M. tuberculosis* Ser/Thr protein kinase D phosphorylates an anti-anti-sigma factor homolog. *PLoS Pathog* 3: e49.

91. Curry JM, Whalan R, Hunt DM, Gohil K, Strom M, et al. (2005) An ABC transporter containing a forkhead-associated domain interacts with a serine-threonine protein kinase and is required for growth of *Mycobacterium tuberculosis* in mice. *Infect Immun* 73: 4471–4477.

92. Grundner C, Gay LM, Alber T (2005) *Mycobacterium tuberculosis* serine/threonine kinases PknB, PknD, PknE, and PknF phosphorylate multiple FHA domains. *Protein Sci* 14: 1918–1921.

93. Molle V, Kremer L, Girard-Blanc C, Besra GS, Cozzzone AJ, et al. (2003) An FHA phosphoprotein recognition domain mediates protein EmbR phosphorylation by PknH, a Ser/Thr protein kinase from *Mycobacterium tuberculosis*. *Biochemistry* 42: 15300–15309.

94. Papavinasasundaram KG, Chan B, Chung JH, Colston MJ, Davis EO, et al. (2005) Deletion of the *Mycobacterium tuberculosis* pknH gene confers a higher bacillary load during the chronic phase of infection in BALB/c mice. *J Bacteriol* 187: 5751–5760.

95. Dasgupta A, Datta P, Kundu M, Basu J (2006) The serine/threonine kinase PknB of *Mycobacterium tuberculosis* phosphorylates PBPA, a penicillin-binding protein required for cell division. *Microbiology* 152: 493–504.

96. Fiuzza M, Canova MJ, Zanella-Cleon I, Becchi M, Cozzzone AJ, et al. (2008) From the characterization of the four serine/threonine protein kinases (PknA/B/G/L) of *Corynebacterium glutamicum* toward the role of PknA and PknB in cell division. *J Biol Chem* 283: 18099–18112.

97. Udo H, Munoz-Dorado J, Inouye M, Inouye S (1995) *Myxococcus xanthus*, a gram-negative bacterium, contains a transmembrane protein serine/threonine kinase that blocks the secretion of beta-lactamase by phosphorylation. *Genes Dev* 9: 972–983.

98. D'Andrea LD, Regan L (2003) TPR proteins: the versatile helix. *Trends Biochem Sci* 28: 655–662.

99. Neer EJ, Schmidt CJ, Nambudripad R, Smith TF (1994) The ancient regulatory-protein family of WD-repeat proteins. *Nature* 371: 297–300.

100. Smith TF, Gaitatzes C, Saxena K, Neer EJ (1999) The WD repeat: a common architecture for diverse functions. *Trends Biochem Sci* 24: 181–185.

101. Yeats C, Finn RD, Bateman A (2002) The PASTA domain: a beta-lactam-binding domain. *Trends Biochem Sci* 27: 438.

102. Aravind L, Ponting CP (1997) The GAF domain: an evolutionary link between diverse phototransducing proteins. *Trends Biochem Sci* 22: 458–459.

103. Kehoe DM, Grossman AR (1996) Similarity of a chromatic adaptation sensor to phytochrome and ethylene receptors. *Science* 273: 1409–1412.

104. Adindla S, Inampudi KK, Guruprasad K, Guruprasad L (2004) Identification and analysis of novel tandem repeats in the cell surface proteins of archaeal and bacterial genomes using computational tools. *Comp Funct Genomics* 5: 2–16.

105. Wright GD, Thompson PR (1999) Aminoglycoside phosphotransferases: proteins, structure, and mechanism. *Front Biosci* 4: D9–21.

106. Iyer LM, Aravind L (2004) The emergence of catalytic and structural diversity within the beta-clip fold. *Proteins* 55: 977–991.

107. Nystrom T, Neidhardt FC (1994) Expression and role of the universal stress protein, UspA, of *Escherichia coli* during growth arrest. *Mol Microbiol* 11: 537–544.

108. Bateman A, Murzin AG, Teichmann SA (1998) Structure and distribution of pentapeptide repeats in bacteria. *Protein Sci* 7: 1477–1480.

109. Zhulin IB, Nikolskaya AN, Galperin MY (2003) Common extracellular sensory domains in transmembrane receptors for diverse signal transduction pathways in bacteria and archaea. *J Bacteriol* 185: 285–294.

110. Kami K, Takeya R, Sumimoto H, Kohda D (2002) Diverse recognition of non-PxxP peptide ligands by the SH3 domains from p67(phox), Grb2 and Pex13p. *Embo J* 21: 4268–4276.

111. Durocher D, Henckel J, Fersht AR, Jackson SP (1999) The FHA domain is a modular phosphopeptide recognition motif. *Mol Cell* 4: 387–394.

112. Krupa A, Abhinandan KR, Srinivasan N (2004) KinG: a database of protein kinases in genomes. *Nucleic Acids Res* 32: D153–155.

113. Altschul SF, Madden TL, Schaffer AA, Zhang J, Zhang Z, et al. (1997) Gapped BLAST and PSI-BLAST: a new generation of protein database search programs. *Nucleic Acids Res* 25: 3389–3402.

114. Marchler-Bauer A, Anderson JB, DeWeese-Scott C, Fedorova ND, Geer LY, et al. (2003) CDD: a curated Entrez database of conserved domain alignments. *Nucleic Acids Res* 31: 383–387.

115. Gowri VS, Krishnadev O, Swamy CS, Srinivasan N (2006) MulPSSM: a database of multiple position-specific scoring matrices of protein domain families. *Nucleic Acids Res* 34: D243–246.

116. Gowri VS, Tina KG, Krishnadev O, Srinivasan N (2007) Strategies for the effective identification of remotely related sequences in multiple PSSM search approach. *Proteins* 67: 789–794.

117. Eddy SR (1998) Profile hidden Markov models. *Bioinformatics* 14: 755–763.

118. Anamika, Srinivasan N, Krupa A (2005) A genomic perspective of protein kinases in *Plasmodium falciparum*. *Proteins* 58: 180–189.

119. Anamika K, Srinivasan N (2007) Comparative kinomics of *Plasmodium* organisms: unity in diversity. *Protein Pept Lett* 14: 509–517.

120. Anamika K, Bhattacharya A, Srinivasan N (2008) Analysis of the protein kinome of *Entamoeba histolytica*. *Proteins* 71: 995–1006.

121. Anamika K, Martin J, Srinivasan N (2008) Comparative kinomics of human and chimpanzee reveal unique kinship and functional diversity generated by new domain combinations. *BMC Genomics* 9: 625.

122. Bateman A, Birney E, Cerruti L, Durbin R, Etwiller L, et al. (2002) The Pfam protein families database. *Nucleic Acids Res* 30: 276–280.

123. Li W, Jaroszewski L, Godzik A (2001) Clustering of highly homologous sequences to reduce the size of large protein databases. *Bioinformatics* 17: 282–283.

124. Larkin MA, Blackshields G, Brown NP, Chenna R, McGgettigan PA, et al. (2007) Clustal W and Clustal X version 2.0. *Bioinformatics* 23: 2947–2948.

125. Saitou N, Nei M (1987) The neighbor-joining method: a new method for reconstructing phylogenetic trees. *Mol Biol Evol* 4: 406–425.

126. Tamura K, Dudley J, Nei M, Kumar S (2007) MEGA4: Molecular Evolutionary Genetics Analysis (MEGA) software version 4.0. *Mol Biol Evol* 24: 1596–1599.