

Removal of lead from aqueous solutions using an immobilized biomaterial derived from a plant biomass

K. Chandra Sekhar^{a,*}, C.T. Kamala^a, N.S. Chary^a, A.R.K. Sastry^b,
T. Nageswara Rao^a, M. Vairamani^a

^a Analytical Chemistry and Environmental Sciences Division, Discovery Lab (D-120), Indian Institute of Chemical Technology, Hyderabad 500 007, AP, India

^b Organic Coatings and Polymers Division, Indian Institute of Chemical Technology, Hyderabad 500 007, AP, India

Received 26 August 2003; received in revised form 29 January 2004; accepted 29 January 2004

Abstract

Because of the severity of heavy metal contamination and potential adverse health impact on the public, a tremendous effort has taken place to purify waters containing toxic metal ions. Traditional methods which have been employed prove to be costly and prohibitive for low level waste remediation. Biosorption is presented as an alternative to traditional physicochemical means for removing toxic metals from ground and wastewaters. Most recently, plant based biomaterials have been of interest.

The bark of *Hemidesmus indicus*, an extensively available plant biomass commonly called as Indian sarsaparilla was used as biomaterial for removal of lead from aqueous streams. Batch experiments were carried out with immobilized biomass of *H. indicus* (IPBFIX) to optimize the experimental parameters like effect of contact time, initial metal concentration, initial IPBFIX concentration and co-metal ion effect on biosorption of lead from contaminated waters. Column experiments were performed under flow conditions for regeneration and recycle efficiency of IPBFIX and was found to be effective for three cycles. Elution experiments were carried out to remove lead ions from loaded IPBFIX and 100% elution was achieved with a 0.1 M HNO₃ solution. The effectiveness of the IPBFIX for biosorption of lead ions was demonstrated using the wastewater samples emanating from a non-ferrous metal industry and the results are presented in this paper. The results from these studies will be useful for a novel phytofiltration technology to remove and recover lead from wastewaters and this can also be well adapted for secondary treatment or polishing of wastewaters. An attempt has been made to remove lead from the lead polluted waters (both ground and surface) from an industrially contaminated sites.

© 2004 Elsevier B.V. All rights reserved.

Keywords: Lead; Immobilization; Biomass; Beads; Phytofiltration; Regeneration

1. Introduction

As today's technology progresses, the natural environment suffers from the detrimental effects of industrial pollution. The natural process of transportation of metal ions between the soil and water consolidates metal contamination in high concentrations that affect the areas of natural ecosystems [1]. Industries such as smelters, metal refineries, and mining operations have been indicated as major sources of metal release into the environment [2–6]. Heavy metal contamination that does get into the environment could cause permanent negative ecological effects [7]. Many attempts have been made throughout the globe to clean heavy metal

polluted groundwaters and soils. However, recent studies have shown that many of the treated polluted groundwater sites have not been restored back to drinking water standards [8]. Therefore, heavy metal contamination of the environment has become an area of increasing concern. The limitations of conventional groundwater remediation have spurred many investigations into alternative methods that are more cost effective. With the increase in environmental awareness and governmental policies, there has been a push toward development of new environmentally friendly ways to clean contamination by using low cost methods and materials.

Biological methods for remediation may provide the answer [9–11]. Among the various biological systems, dead systems offer many advantages over live systems because they do not fall prey to toxicological effects of high concentrations of contaminants and can be obtained inexpensively [12]. However, recovery of metal from loaded biomass is

* Corresponding author. Fax: +91-40-7160387.

E-mail addresses: kavi223@yahoo.com, kavi@iict.ap.nic.in
(K. Chandra Sekhar).

still cumbersome. Recently, plants have been studied for their ability to remove contaminants from the environment [13–19]. Researchers have recognized that immobilizing biomass in a granular or polymeric matrix may improve biomass performance and facilitate separation of biomass from solution [20,21]. The American Bureau of Mines has investigated the use of biomass immobilized in porous poly-sulfonate beads (BIOFIX) for extracting toxic and heavy metals from dilute waste streams [22]. Gardea-Torresdey et al. [23] have removed copper ions from aqueous solutions by silica immobilized *Medicago sativa* (alfalfa). Immobilized biomass appears to have a greater potential than raw biomass in packed bed or fluidized bed reactors with benefits such as control of particle size, better capabilities of regeneration, reuse and recovery without destruction of biomass beads [24–26].

Lead contamination of drinking water occurs as a result of corrosion and leaching from Pb/Sn soldered joints associated with the copper service lines commonly used in household plumbing [27]. Lead poisoning in human causes severe damage to the kidney, nervous system, reproductive system, liver and brain and causes sickness or death [28]. Severe exposure to lead has been associated with sterility, abortion, stillbirth and neonatal deaths [29]. The permissible limits for lead in drinking water given by US Environmental Protection Agency (USEPA) is 0.015 mg dm^{-3} [30] and for wastewaters is 0.1 mg dm^{-3} given by both USEPA and Bureau of Indian Standards (BIS) [31,32]. Thus it becomes mandatory for the removal of lead from drinking and wastewaters.

We have selected *Hemidesmus indicus*, a plant biomass as a source for biomaterial because it has a higher tolerance to lead contamination [33]. The objective of our study is to investigate the binding of Pb from representative wastewaters by immobilized *H. indicus* biomass beads and to indicate factors that impact the sorption and elution characteristics of the beads. Laboratory batch experiments were performed with biomass beads to determine optimal time required for lead ion binding to the biomass, effect of varying IPBFIX loadings, initial lead ion concentration and effect of co-metal ions such as Zn and Cr. Column experiments were performed under flow conditions for regeneration of IPBFIX beads for their re-use. The elution experiments were carried out with varying concentration (0.025–0.2 M) of eluants such as HCl, HNO_3 , EDTA, Na_2CO_3 and NaOH for elution of lead ions from the loaded IPBFIX. Biosorption experiments with IPBFIX for real-life samples from non-ferrous metal processing wastes was also carried out.

2. Experimental

2.1. Materials and methods

2.1.1. Chemicals and reagents

The stock solutions of Pb and other co-ions Zn and Cr were prepared by dissolving appropriate quantities of

$\text{Pb}(\text{NO}_3)_2$ (Drug houses [INDIA] Pvt. Ltd., Graham Road, Mumbai), $\text{Zn}(\text{CH}_3\text{COO})_2$ (Glaxo Laboratories IND Ltd., Mumbai) and $\text{K}_2\text{Cr}_2\text{O}_7$ (Ranbaxy Laboratories Ltd., Punjab) salts in 5% HNO_3 , respectively. The pH of the working solutions was maintained with 0.01 M HNO_3 or NH_3 using a Elico pH meter (Elico Pvt. Ltd., India). All the chemicals used were of AR grade and deionized distilled water was used for all the experiments.

2.1.2. Preparation of biomass beads

Our previous experimental studies with raw biomass of *H. indicus* have proved that the root bark of the plant is a potential biosorbent for lead ions [33]. *H. indicus* commonly known as Indian sarsaparilla is taxonomically classified as a member of family Asclepiadaceae (Warrier, 1995). The biomass was oven dried at 100°C for 24 h and powdered to pass through a 1 mm mesh and used for immobilization. Immobilized biomass beads (IPBFIX) were prepared using a method developed by Indian Institute of Chemical Technology (IICT) (experimental details not given).

2.1.3. Equilibrium experimental set-up

Equilibrium experiments were conducted with stoppered flasks in a mechanical shaker (Techno India Ltd.). One gram of IPBFIX was washed twice with 50 ml of 0.01 M HCl to remove any debris or soluble biomolecules that might interact with metal ions. Washings were collected, and IPBFIX was dried and weighed to account for any weight loss of the IPBFIX. The IPBFIX was then stirred with 100 ml of 100 ppm lead ion solutions at room temperature ($\pm 30^\circ\text{C}$) at 300 rpm for 5 min. The pH of the lead ion solution was taken as it is (pH 1.8–2.0) as our earlier batch experimental studies with raw biomass of *H. indicus* revealed that lead ion removal from aqueous solutions was independent of pH variation and hence no experiments were carried out with IPBFIX to study the effect of pH. After biosorption the samples were filtered and analyzed three times along with a control sample by AAS for lead [34].

Batch experiments were conducted to study the effect of various parameters on the biosorption of lead by the IPBFIX such as metal to IPBFIX contact period, IPBFIX loadings, initial metal ion concentration and co-metal ions.

2.1.4. Column set-up

Experiments using column set-up (Fig. 1) as per Reddy et al. [15] were conducted to determine the IPBFIXs retention capacity for lead ions, elution of metal ions from loaded IPBFIX, and regeneration of the IPBFIX for its reuse. The results of such a study are useful for determining the basic engineering data, which is necessary for scaling up the process. Ten grams of IPBFIX (bed height 7 cm) was packed into the column having an internal diameter of 1.6 cm and a column height of 20 cm. One liter of solution containing 100 ppm of lead was passed through the column containing IPBFIX at a flow rate of 0.8 l/h for 24 h at the predetermined pH. The pH of the inlet metal solution was

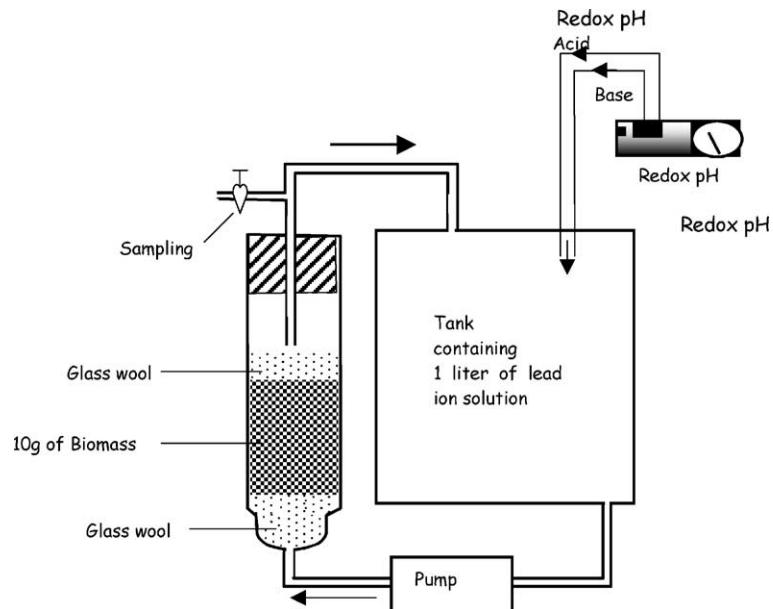


Fig. 1. Column experimental set-up.

continuously monitored and maintained constant automatically by the addition of 0.01 M NH₄OH or HCl. After the experiment, the concentration of metal ions in the solution was analyzed to determine the retention capacity of the IPBFIIX. Elution of metal from loaded IPBFIIX was carried out with 100 ml (at a flow rate of 1 ml/min) each of different concentrations (0.025–0.2 M) of HCl, HNO₃, EDTA, Na₂CO₃ and NaOH eluants. The eluted IPBFIIX beads were washed twice with 500 ml of deionized water for regeneration. The regenerated IPBFIIX beads were tested for lead uptake to determine the evolution of its retention capacity.

3. Results and discussion

3.1. Batch studies

3.1.1. Kinetic study for lead binding to the IPBFIIX

The time dependency batch experiments were performed with varying the contact time of IPBFIIX to lead ions from 5 to 120 min (5, 15, 30, 60 and 120 min). All other parameters such as IPBFIIX concentration (1 g), volume of lead ion solution (100 ml) was kept constant. The experiments were conducted to determine the time required for IPBFIIX beads to bind the lead ions. It can be seen from Fig. 2 that 99.2% lead ions are bound to the IPBFIIX within 5 min of contact after which it increased marginally in 30 min and stabilized thereafter. The rapid binding of metal ions from solution compared to 15 min by pure biomass [33] could mean that the binding sites for lead are located on the *H. indicus* cell walls and also the immobilization did not alter the binding sites and instead enhanced the binding capac-

ity of IPBFIIX. Similar conclusions were obtained by Zhang and Majidi [35].

3.1.2. Effect of IPBFIIX concentration

The percentage lead removal was studied with varying IPBFIIX loadings from 0.1 to 2.5 g (0.1, 0.5, 1.0, 1.5, 2.0 and 2.5 g) and taking the pH of the metal solution as it is (pH ~ 2). The parameters such as contact time (5 min), volume of lead ion solution (100 ml), and initial lead ion concentration (100 ppm) were kept constant. The results of the experiment are presented in Fig. 3a and b. From the figure it is clear that by using 1 g IPBFIIX, 100% of the metal can be removed whereas 2 g of the non-immobilized raw biomass required for the same amount of metal removal [33]. This may be explained due to the increase in surface area of the biomass due to immobilization. Experiments with control (the polymer used for immobilization) revealed no metal removal indicating the metal removal is only due to plant biomass. All these data obtained with simple experiments can be of great interest in scale-up process to optimize

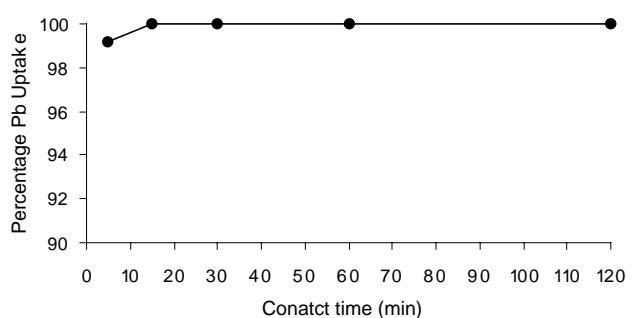


Fig. 2. Effect of kinetics on biosorption.

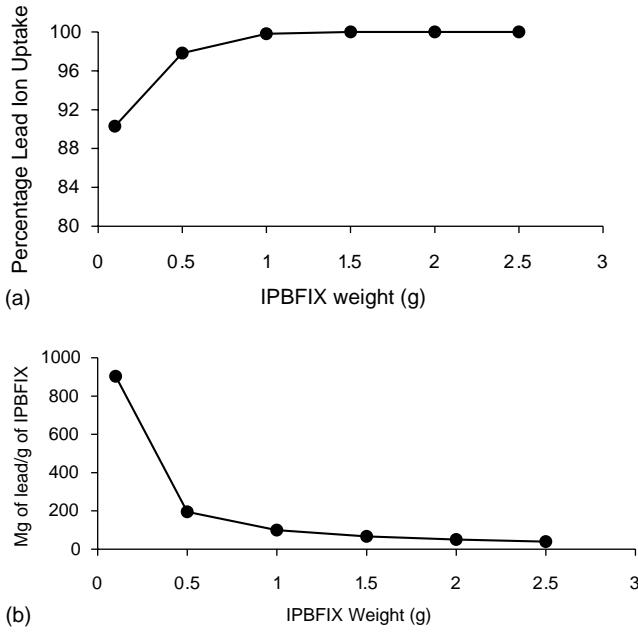


Fig. 3. Effect of IPBFIX loadings on biosorption of lead ions.

industrial effluent purification [36]. These results show that by immobilization, using smaller amount of IPBFIX, higher concentration of metal can be removed compared to raw biomass.

3.1.3. Effect of initial metal ion concentration

The metal removal by IPBFIX corresponding to different initial metal concentrations (25, 50, 100, 250, 500, 1000, 2000, 4000, 5000 and 6000 ppm) are shown in Fig. 4a and b. The parameters such as contact time (15 min), volume of solution (100 ml) and IPBFIX loading (1 g) were

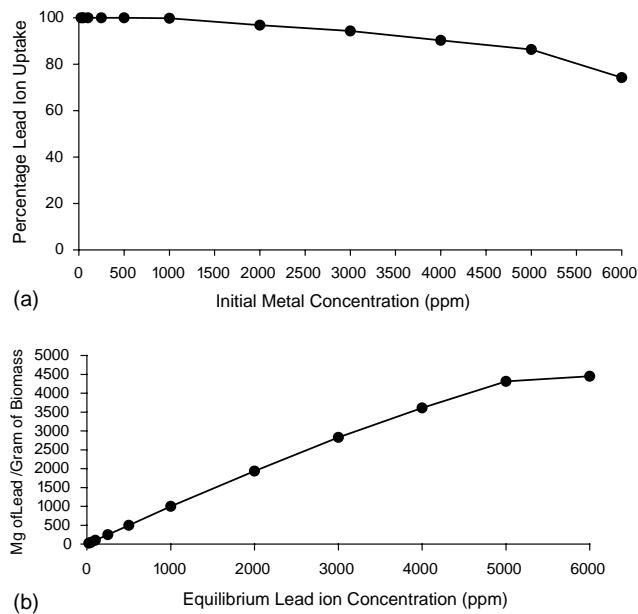


Fig. 4. Effect of initial metal concentrations on biosorption of lead.

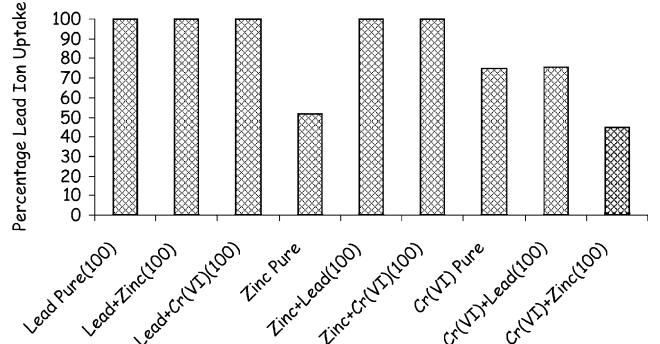


Fig. 5. Effect of multi-component system on metal removal.

kept constant. As can be seen from Fig. 4a, removal of lead ions is 99.8% till 1000 ppm concentration of lead ions in aqueous solutions. The binding sites are limited keeping IPBFIX loadings as constant. With increasing metal concentration, the percentage of metal removed stabilized and then decrease to 70 for 6000 ppm of lead ions in solution. In our earlier experiments with raw *H. indicus* biomass the percentage lead removal above 1000 ppm metal decreased due to slurry formation [33]. Thus by using IPBFIX the problem of slurry formation was avoided and the beads were stable for metal concentrations as high as 6000 ppm.

3.1.4. Co-ion effect

The co-ion experiments were conducted with 1.0 g of IPBFIX and a shaking time of 15 min and keeping the volume constant (100 ml). The results of the experiment are presented in Fig. 5. The results revealed that lead removal was not affected by any of the cations (Zn and Cr), however, the presence of co-ions enhanced Zn removal but decreased Cr removal. In the presence of co-ions in solution, chemical interactions between the ions themselves as well as with the biomass take place resulting in site competition [37,38]. Many of the functional groups present on the cell wall and the membrane are non-specific and different cations compete for the binding sites. It has been reported that metal removal is increased as the ionic radii of metal cations affect the ion-exchange and adsorption process [39]. The differences in the sorption affinities may also be attributed to differences in the electrode potentials of the various ions. The greater the electrode potential, the greater is the affinity for biomass [40,41]. The ionic radii and the electrode potentials of the metal ions are summarized in Table 1. In multicomponent systems, the complex interactions of

Table 1
Ionic charge, ionic radii and electrode potentials of metal ions

Metal ion	Ionic charge	Ionic radii (Å)	Electrode potential (V)
Pb ²⁺	+2	1.32	0.126
Zn ²⁺	+2	0.74	0.763
Cr ⁶⁺	+6	0.64	0.744

several factors such as ionic charge, ionic radii and electrode potential will account for the differences in the metal removal capacity of the biomass. As a result, ordering of the metal ions based on a single factor is very difficult.

3.2. Column studies

3.2.1. Elution tests

Efficient removal of loaded metal from the IPBFIX biomass beads was necessary to ensure their long-term use for repeated extraction–elution cycles. A review of literature indicated that dilute mineral acids, ethylenediamine tetraacetic acid (EDTA), and caustic solutions [41] have been utilized to remove sorbed metal ions from microorganisms. These reagents were therefore evaluated for eluting sorbed metals from the IPBFIX beads.

Mineral acids evaluated as potential eluants included HNO_3 and HCl . Experimental conditions were maintained as explained in column set-up. Among the mineral acids 0.1 M HNO_3 was very effective eluant than HCl (Fig. 6). In the first 10 min of contact, 90% of elution occurred and complete metal elution (100%) was achieved within half an hour. EDTA was also evaluated as a potential metal eluant but is not as effective as HNO_3 . The caustic solutions studied were NaOH and Na_2CO_3 . Sodium carbonate was the most effective of the caustic eluants tested. A 0.1 M sodium carbonate eluted 75% of the metal. Among all the eluants used 0.1 M nitric acid proved to be the best eluant.

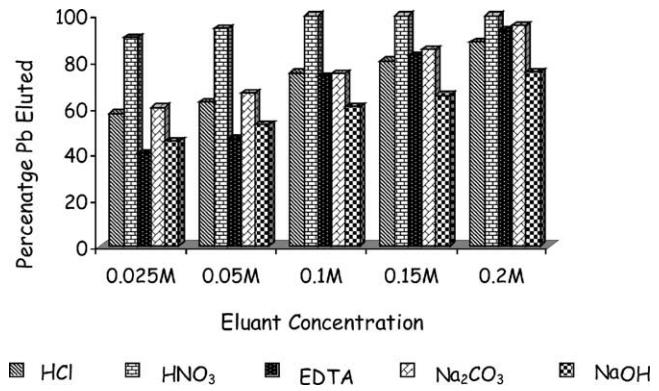


Fig. 6. Elution tests.

3.2.2. Regeneration of loaded beads for its reuse

The batch laboratory experiments showed that IPBFIX has ability to bind lead and remove it from aqueous lead ion solutions, but batch system may not be practicable for removing lead ions from contaminated waters. Therefore, column experiments were performed to study the binding of lead ions to the IPBFIX beads under flow conditions.

Experiments were carried out using the conditions in column set-up for retention capacity of IPBFIX for lead ions and its regeneration trials and the results are presented in Table 2. From the results it is very much clear that elution of lead ions from IPBFIX is stable till three cycles when compared to the raw biomass. This suggests that the IPBFIX could be regenerated for use in continuous operation of

Table 2
Regeneration of IPBFIX and raw biomass

Number of cycles	Raw biomass		IPBFIX	
	Lead uptake (mg/g)	Lead elution (mg/g)	Lead uptake (mg/g)	Lead elution (mg/g)
1	93.6 ± 3.9	90.2 ± 3.8	100 ± 4.5	100 ± 3.6
2	89.3 ± 1.8	85.3 ± 4.1	99.8 ± 3.1	99.6 ± 4.1
3	74.2 ± 3.2	70.2 ± 3.8	99.7 ± 2.4	99.5 ± 3.8

Table 3
Analytical application

Number	Sample label and composition	Pb (mg/g) removal	
		Raw biomass	IPBFIX beads
Model solutions			
1	Control composition (ppm): $\text{Pb} = 1000$	900 ± 10.8 (90)	1000 ± 10.5 (100)
2	Syn-1 composition (ppm): $\text{Pb} = 1000$; $\text{Zn} = 5000$; $\text{Cr} = 1000$	790 ± 8.6 (79)	1000 ± 9.2 (100)
3	Syn-2 composition (ppm): $\text{Pb} = 2500$; $\text{Zn} = 2500$; $\text{Cr} = 500$	1875 ± 14.3 (75)	1000 ± 11.5 (100)
4	Syn-3 composition (ppm): $\text{Pb} = 5000$; $\text{Zn} = 1000$; $\text{Cr} = 2500$	3300 ± 12.5 (66)	1000 ± 11.3 (100)
“Real-life” samples			
5	Real-1 composition (ppm): $\text{Pb} = 26.04$; $\text{Zn} = 0.41$	19.6 ± 0.02 (75.27)	25.7 ± 0.015 (99)
6	Real-2 composition (ppm): $\text{Pb} = 19.27$; $\text{Zn} = 0.94$	18.4 ± 0.03 (95.87)	19.27 ± 0.016 (100)
7	Real-1 + Cr Syn composition (ppm): $\text{Pb} = 26.04$; $\text{Zn} = 0.41$; $\text{Cr} = 100$	19.8 ± 0.013 (76.38)	25.51 ± 0.012 (98)
8	K1 (surface waters): $\text{Pb} = 20.3$ ppm	16.09 ± 0.012 (79.3)	20.09 ± 0.010 (99)
9	K2 (surface waters): $\text{Pb} = 16.7$ ppm	12.9 ± 0.013 (77.3)	16.7 ± 0.010 (100)
10	K3 (groundwater): $\text{Pb} = 33.2$ ppm	8.81 ± 0.016 (75.3)	33.2 ± 0.10 (100)
11	K4 (groundwater): $\text{Pb} = 30.7$ ppm	7.8 ± 0.015 (76.5)	30.7 ± 0.10 (100)

Values in parenthesis are % lead removal.

metal ion removal from aqueous solutions. The decreased efficiency of metal uptake and release by raw biomass could be due to physical damage to the biomass caused by the extended volumes of the eluant.

3.2.3. Analytical application

The studies reported here are conducted using model metal ion solutions. However, the metal ions at the range of concentrations chosen are representative of typical waste effluents emanating from various non-ferrous processing industries. Therefore results of this work have considerable practical implications. The results shown in Table 3 represent the comparison of metal removal in model co-ion solutions, real effluent collected from non-ferrous metal industry and also the ground and surface water samples collected from the lead contaminated sites of Kattedan Industrial Area, Hyderabad. As we did not get “real-life” effluent samples for Cr(VI) we have spiked Real-1 sample with Cr(VI). Table 3 clearly shows that IPBFIIX is effective in lead ion removal when compared to raw biomass. The results from these experiments prove that IPBFIIX can be used effectively for the removal of lead ions from aqueous systems.

4. Conclusions

The characterization of lead uptake by IPBFIIX showed that the binding kinetics is pH independent up to pH 5.0 and occurs in less than 5 min. The binding of the lead ions is on the cell wall of the plant biomass but not on the polymer material used for immobilization. The IPBFIIX that was saturated with lead ions shows the remarkable ability for lead recovery by treatment with low concentrated nitric acid. Through column experiments it was showed that the IPBFIIX was not only successful for effective lead removal but it was also reusable. These studies show that IPBFIIX has the potential to be used as a biofilter for fast removal and recovery of lead ions from contaminated waters. This innovative technology provides a reusable material, which is environmentally friendly.

Acknowledgements

The authors are grateful to Dr. K.V. Raghavan, Director, Indian Institute of Chemical Technology (IICT), and Dr. V. Balaram, Head, Geochemistry Department, National Geo-physical Research Institute (NGRI) for their encouragement and support in carrying out this work.

References

- [1] D.D. Runnels, T.A. Shepherd, E.E. Angino, Metals in determining natural background concentrations in mineralized areas waters, Environ. Sci. Technol. 26 (12) (1992) 2316–2323.
- [2] P.H. Kansenen, J. Venetvaara, Comparison of biological collectors of airborne heavy metals near ferrochrome and steel works, Water Air Soil Pollut. 60 (3–4) (1991) 337.
- [3] D.E. Kimbrough, I.H. Suffet, Off-site forensic determinations of airborne elemental emissions by multimedia analysis—a case study at 2 secondary lead smelters, Environ. Sci. Technol. 29 (1995) 2217.
- [4] M. Bergers, T. Harris, Pb and Cd speciation in smelter-contaminated soil, in: Proceedings of the 10th Annual Conference on Hazardous Waste Research, Kansas State University, Manhattan, KS, 1995, p. 324.
- [5] E. Pip, Cd, Cu and Pb in soils and garden produce near a metal smelter at Flin-Flon, Manitoba, Bull. Environ. Contam. Toxicol. 46 (5) (1991) 790–796.
- [6] C. Countryman, Arsenic and Old Lakes, The Texas Observer, February 6, 1995.
- [7] G. Micera, A. Densi, Chromium adsorption by plant roots and formation of long lived Cr(V) species—an ecological hazard, J. Inorg. Biochem. 34 (1988) 157–166.
- [8] R. Bruce, In Situ Bioremediation: When Does it Work? Committee on In Situ Bioremediation, Water Science and Technology Board, Commission on Engineering and Technical Systems National Research Council, National Academy Press, Washington, DC, 1993.
- [9] H. Tan, J.T. Champion, J.F. Artiola, M.L. Bruscan, R.M. Miller, Complexation of Cd by a Rhamnolipid biosurfactant, Environ. Sci. Technol. 28 (13) (1994) 2402–2406.
- [10] K. Chandra Sekhar, K.A. Natarajan, S. Subramaniam, J.M. Modak, Removal of metal ions using an industrial biomass with reference to environmental control, Int. J. Miner. Process. 53 (1998) 107–120.
- [11] R. Jalali, H. Ghafourian, Y. Asef, S.J. Davarpanah, S. Sepehr, Removal and recovery of lead using nonliving biomass of marine algae, J. Hazard. Mater. 92 (3) (2002) 253–262.
- [12] D. Kratochvil, B. Volesky, Advances in biosorption of heavy metals, Trends Biotechnol. 16 (7) (1997) 291–300.
- [13] V. Dushenkov, P.B.A.N. Kumar, H. Motto, I. Raskin, Rhizofiltration: the use of plants to remove heavy metals from aqueous streams, Environ. Sci. Technol. 29 (1995) 1232–1245.
- [14] C.D. Scott, Removal of dissolved metals by plant tissues, Biotechnol. Bioeng. 39 (10) (1992) 1064–1068.
- [15] B.R. Reddy, N. Mirghaffari, I. Gaballah, Removal and recycling of copper from aqueous solutions using treated Indian barks, Resour. Conserv. Recycl. 21 (1997) 227–245.
- [16] I. Villaescusa, M. Maria, M. Nuria, Heavy metal uptake from aqueous solutions by cork and yohimbe bark wastes, J. Chem. Technol. Biotechnol. 75 (9) (2000) 812–816.
- [17] P. Kumar, S.S. Dara, Modified barks for scavenging toxic heavy metal ions, Ind. J. Environ. Health 22 (3) (1980) 196–202.
- [18] G. Sun, W. Shi, Sunflower stalks as adsorbents for the removal of metal ions from wastewater, Ind. Eng. Chem. Res. 37 (1998) 1324–1328.
- [19] M.S. Osman, M. Bandyopadhyah, Cadmium removal from water environment by a fungus *Volvariella volvacea*, Bioprocess Eng. 14 (5) (1996) 249–254.
- [20] D.W. Darnall, B. Greene, M.T. Henzl, J.M. Hosea, R.A. McPherson, J. Sneddon, M.D. Alexander, Selective recovery of gold and other metal ions from an algal biomass, Environ. Sci. Technol. 20 (2) (1986) 206–208.
- [21] J.A. Brierley, C.L. Brierley, R.F. Decker, G.M. Goyak, Treatment of microorganisms with alkaline solution to enhance metal uptake properties, US Patent 4,690,894 (September 1, 1987).
- [22] T.H. Jeffers, C.R. Ferguson, Bennett, Biosorption of metal contaminants using immobilized biomass—a laboratory study, Report of Investigation No. 9340, United States Department of the Interior, Bureau of Mines, 1992.
- [23] J.L. Gardea-Torresdey, K.J. Tiemann, J.H. Gonzalez, J.A. Henning, M.S. Townsend, Ability of silica immobilized *Medicago sativa* (alfalfa) to remove copper ions from solution, J. Hazard. Mater. 48 (1996) 181–190.

[24] B. Volesky, Z.R. Holan, Biosorption of heavy metals, *Biotechnol. Prog.* 11 (1996) 235–250.

[25] A.C. Texier, Y. Andres, B.C. Faur, C.P. Lee, Fixed bed study for lanthanide (La, Eu, Yb) ions removal from aqueous solutions by immobilized *Pseudomonas aeruginosa*: experimental data and modelisation, *Chemosphere* 47 (2002) 333–342.

[26] L. Shan, R.D. Lawrence, D.R. Gary, Affinity distributions of lead ion binding to an immobilized biomaterial derived from cultured cells of *Datura innoxia*, *Adv. Environ. Res.* 6 (2002) 523–532.

[27] K.S. Subramaniam, J.W. Coonor, Lead contamination of drinking water, *J. Environ. Sci. Health, Part A* 54 (1991) 29–33.

[28] S. Manahan, *Environmental Chemistry*, Brooks/Cole, California, USA, 1984.

[29] R.A. Goyer, I.J. Chisolm, Lead, in: D.H.K. Lee (Ed.), *Metallic Contaminations and Human Health*, Academic Press, New York, 1972, p. 57.

[30] US Environmental Protection Agency (USEPA), *Guidelines for Drinking Water*, March 2001.

[31] K.V.S.G. Muralikrishna, *Chemical Analysis of Water and Soil—A Laboratory Manual*, Environmental Protection Society, National Institute of Ecology and Environment, Kakinada, India, 1997.

[32] IS: 10500, *Drinking Water Standards*, Bureau of Indian Standards (BIS), 1995.

[33] K. Chandra Sekhar, C.T. Kamala, N.S. Chary, Y. Anjaneyulu, Removal of heavy metals using a plant biomass with reference to environmental control, *Int. J. Miner. Process.* 68 (2003) 37–45.

[34] *Analytical Methods for Atomic Absorption Spectrometry*, Perkin Elmer Instruments Manual, 2002, p. 96.

[35] W. Zhang, V. Majidi, Monitoring the cellular response of *Stichococcus bacillaris* to exposure of several different metals using *in vivo* ^{31}P NMR and other spectroscopy techniques, *Environ. Sci. Technol.* 28 (9) (1994) 1577–1581.

[36] E. Forest, J.C. Roux, Heavy metal biosorption by fungal mycelial byproducts—mechanisms and influence of pH, *Appl. Microbiol. Biotechnol.* 37 (1992) 399–403.

[37] S. Schiewer, B. Volesky, Modeling multi-metal exchange in biosorption, *Environ. Sci. Technol.* 30 (1996) 2921–2927.

[38] M.P. Khovrychev, T.Y. Mareev, V.F. Pomytkin, Ability of microorganisms to sorb radionuclides, *Microbiology* 63 (1) (1994) 83–86.

[39] Y. Marcus, A.S. Kertes, *Ion Exchange and Solvent Extraction of Metal Complexes*, Wiley, New York, 1969.

[40] B. Mattuschka, G. Straube, Biosorption of metals by waste biomass, *J. Chem. Technol. Biotechnol.* 58 (1993) 57–63.

[41] J.C. Jenette, J.E. Smith, J.M. Hassett, Factors influencing metal accumulation by algae, *NTIS PB 83-149377*, 1982, p. 124.