Prospecting for alternate sources of shikimic acid, a precursor of Tamiflu, a bird-flu drug

Shikimic acid, more commonly known by its anionic form, shikimate, is an important intermediate compound of the 'shikimate pathway' in plants and microorganisms1. It is the principal precursor for the synthesis of aromatic amino acids, phenylalanine, tryptophan and tyrosine and other compounds such as alkaloids, phenolics and phenyl propanoids². It is used extensively as a chiral building block for the synthesis of a number of compounds in both pharmaceutical and cosmetic industries3. In the recent past, the focus on shikimic acid has increased since it is the key precursor for the synthesis of Tamiflu, the only drug against avian flu caused by the H5N1 virus4,5. Shikimic acid is converted to a diethyl ketal intermediate, which is then reduced in two steps to an epoxide that is finally transformed to Tamiflu6.

Currently most of the world's demand for shikimic acid is met from fruits of Chinese star anise (Illicium verum), a small tree reaching up to about 10 m (ref. 7). The fruits of this tree have been traditionally used in China for culinary as well as medicinal purposes8, and are reported to contain about 2-7% of shikimic acid, one of the highest reported estimates in plants9. However, because the tree is not easily cultivable, attaining its first seedbearing stage after 6 years of growth, it is unlikely that the growing market demand would be met by the I. verum source alone. Consequently efforts are being made to explore alternate sources of shikimic acid to meet the world market. In the recent past, there have been attempts to prospect for alternate plant sources for shikimic acid10. Kramer et al.11 reported the fermentative production of shikimic acid by metabolic engineering of Escherichia coli. It is estimated that nearly two-thirds of the requirement of shikimic acid is still being sourced from plants, with the remaining one-third only obtained from genetically engineered E. coli12. In summary, with shikimic acid being used as a bulk chemical for various industrial and pharmaceutical uses, it is imperative that newer sources of this chemical are identified.

Against this background, we have explored for alternate sources of shikimic acid from plants of the Western Ghats, a megadiversity hotspot in South India¹³. Based on our analysis of 210 plant species, we have identified broad taxonomic patterns of shikimic acid richness in the Western Ghats and a few promising species that may serve as newer sources of shikimic acid.

Plant species were selected either randomly or were based on previous reports of a related species, genera or family accumulating shikimic acid. The latter followed the assumption that chemical accumulation in plants could be a function of the phylogenetic relatedness of species. A database was developed of previous reports on the occurrence of shikimic acid in plant species. Phylogenetically related genera and species occurring in the Western Ghats were then selected and sampled. A total of 193 angiosperms belonging to 59 families and 17 gymnosperms belonging to five families were collected for the study. Based on the consistency of results, we extracted the tissues following Harring et al.14. Following the extraction, shikimic acid was quantified using a HPLC system and the detection method reported by Lydon and Duke15. Shikimic acid levels were quantified in the plant samples using the regression of peak areas against the standard shikimic acid and expressed as per cent dry weight of tissue.

Shikimic acid content was estimated in 193 angiosperms belonging to 59 families. Only seven of the 193 plant species yielded shikimic acid in excess of 1%, while the rest yielded low or no shikimic acid. Among the 59 families, only Clusiaceae, Illiciaceae and Combretaceae accumulated higher levels of shikimic acid; the rest of the families accumulated little or no shikimic acid. The most promising species were Calophyllum apetalum (4.10% shikimic acid by dry weight), Pinanga dicksonii (1.05%), Garcinia morella (1.45%), Melia azedarach (1.40%) and Aegle marmelos (1.00%). The overall mean shikimic acid content over all the angiosperms analysed was $0.17 \pm 0.15\%$.

All the 17 gymnosperm species evaluated had detectable levels of shikimic acid, with six species accumulating greater than 1% by dry weight. High yields were obtained from species belonging to

Araucariaceae and Pinaceae. Araucaria excelsa R.Br. belonging to the Araucariaceae family, yielded the highest levels of shikimic acid (5.02%). The overall mean shikimic acid content of gymnosperm species $(X = 1.48 \pm 1.64\%)$ was significantly greater than that obtained from the angiosperm species (P < 0.001; Figure 1). These results confirm earlier reports that had indicated the higher and more widespread occurrence of shikimic acid in gymnosperms (Figure 1). For example, Bohm16 compared the occurrence of shikimic acid in the major plant taxa and showed that over 60% of all gymnosperms analysed were found to accumulate shikimic acid compared to just under 50% for angiosperms (dicots and monocots). Hattori et al.17 also reported a generally higher level of shikimic acid accumulation in gymnosperms compared to angiosperms.

In summary, the profiling of 210 species in the Western Ghats has led to the identification of newer and promising sources of shikimic acid. Though it would be interesting to address the underlying reasons for the differences in the accumulation of shikimic acid across taxa, unfortunately not much is known. It is often conjectured and also empirically demonstrated in several instances that blockage of the shikimate pathway in plants could result in the accumulation of high levels of shikimic acid 15,18,19. For example, treating plants with the herbicide, glyphosate that inhibits the key enzyme, 5 enol pyruvate shikimate 3phosphate (EPSP) synthase in the shikimic acid pathway has been shown to result in shikimic acid accumulation14.

The leads presented here appear more promising than most others reported in

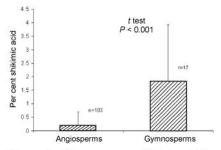


Figure 1. Mean per cent shikimic acid in angiosperms and gymnosperms.

the literature^{2,10,20}. Further, in a few of these species, the estimates are comparable to those reported for star anise² (2.4-7%). However, one of the most significant advantages of the newly identified Indian sources is that the estimates are from leaves and not fruits as is the case star anise. Quite obviously, extraction from leaves will be preferred over that from fruits; moreover, the sheer volume of the biomass offered by the leaves would render it economically feasible. In other words, the finding of the new sources of shikimic acid can potentially be used to meet the emerging needs of both the domestic and the international market.

- Gibson, M. I., Gibson, F., Doy, C. H. and Morgan, P. N., *Nature*, 1962, 195, 1173.
- Payne, R. and Edmonds, M., J. Chem. Edu. (Suppl.), 2005, 82, 599-600.
- Tiedtke, J., Cosmet. Sci. Technol., 2006, 15–21.
- Uma Shaanker, R., Curr. Sci., 2006, 90, 1585–1586.
- Singer, A. C., Nunn, M. A., Gould, E. A. and Johnson, A. C., Environ. Health Perspect., 2007, 115, 102–106.
- 6. Yarnell, A., Chem. Eng. News, 2005, 83, 22-23.
- Wendy, A., The Identification of Medicinal Plants. A Handbook of the Morphology of Botanicals in Commerce, Missouri Botanical Garden Press, 2006, p. 89.

- Ortiz, B. I. and Clauson, K. A., J. Am. Pharm. Assoc., 2006, 46, 161–167.
- Adams, H. et al., Tetrahedron, 1996, 52, 8565–8580.
- Enrich, L. B. et al., Tetrahedron Lett., 2008, 49, 2503–2505.
- Kramer, M. et al., Metab. Eng, 2003, 5, 277–283.
- Frantz, S., Nature Rev. Drug Discovery, 2006, 5, 7–9.
- Myers, N., Mittermeir, R. A., da Fonseca, G. A. B. and Kent, J., *Nature*, 2000, 403, 853–857.
- Harring, T., Streibig, J. C. and Husted, S., J. Agric. Food. Chem., 1998, 46, 4406–4412.
- Lydon, J. and Duke, O. S., J. Agric. Food Chem., 1988, 36, 813–818.
- Bohm, B. A., Chem. Rev., 1965, 65, 435– 466.
- Hattori, S., Yoshida, S. and Hasegawa, M., Physiol. Plant, 1954, 7, 283–289.
- Amrhein, N., Deus, B., Gehrke, P. and Steinrucken, H. C., *Plant Physiol.*, 1980, 66, 830–834.
- Berlin, J. and Witte, L., Z. Naturforsch. C, 1981, 36, 210–214.
- Hudina, M. and Stampar, F., Acta Agricul. Slov., 2005, 85, 179–185.

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