Characterization of Rotavirus Strains from Newborns in New Delhi, India

BIMAL K. DAS,¹ JON R. GENTSCH,^{2*} HELEN G. CICIRELLO,² PATRICIA A. WOODS,² AARTI GUPTA,¹ MADHUMATI RAMACHANDRAN,¹ RAMESH KUMAR,¹ M. K. BHAN,¹ AND ROGER I. GLASS²

Department of Paediatrics and Microbiology, Division of Gastroenterology and Enteric Infections, All India Institute of Medical Sciences, Ansari Nagar, New Delhi 11029, India,¹ and The Viral Gastroenteritis Section, Division of Viral and Rickettsial Diseases, National Center for Infectious Diseases, Centers for Disease Control and Prevention, Atlanta, Georgia 30333²

Received 8 December 1993/Returned for modification 18 April 1994/Accepted 18 April 1994

Between 1986 and 1993, 72% of rotavirus strains isolated from newborns at five hospitals in New Delhi, India, had long electropherotypes, subgroup II VP6 antigens, and G and P genotypes (G_9P_{11}) identical to those of prototype strain 116E. A novel strain with a G_9P_6 genotype, representing 13% of the isolates, was identified. These results demonstrate that G_9P_{11} and G_9P_6 rotavirus strains are common in nurseries in New Delhi.

Serologic characterization of rotavirus strains indicates that outer capsid proteins VP4 and VP7 independently induce type-specific neutralizing antibodies which have been used to classify rotaviruses into G (VP7) and P (VP4) serotypes (11). Among rotavirus strains isolated from children with diarrhea, four major G serotypes, G_1 to G_4 , have been shown to be epidemiologically important by enzyme-linked immunosorbent assay with VP7-specific monoclonal antibodies, but until recently, analogous methods to study the P serotypes of common human rotavirus strains (i.e., G serotypes 1 to 4) were not available (16, 17). As a result, nucleic acid-based (genotyping) methods that detect genetically distinct VP4 genes and accurately predict P serotypes have been developed (8). VP4 genes that are distinct at the level of nucleotide and deduced amino acid sequences have been referred to as P types (or P genotypes) (6). Although a nomenclature for corresponding P serotypes has not been agreed upon, some investigators use the same numbering system for genotypes and serotypes and that is the convention used in this report (6, 14). The largest survey of rotavirus field isolates indicated that genotypes P8 (strain Wa-like VP4 gene) and P_4 (strain RV5-like VP4 gene) are by far the most common among more than 400 strains from several areas of the world (15), while the largest survey in which both G and P genotypes were determined for the same strain indicated that G_1P_8 (strain Wa-like VP7 and VP4 genes) is the most common in Malaysia (13).

We recently demonstrated that asymptomatic neonatal rotavirus infections of children in New Delhi, India, reduced the frequency of subsequent cases of severe rotavirus diarrhea by 46%, essentially confirming the work of Bishop and coworkers and raising the possibility that strains related to New Delhi isolate 116E may be effective as vaccines (1, 2). Previously isolated neonatal rotaviruses belonged to serotypes G_1P_6 (strain M37), G_2P_6 (strain 1076), G_3P_6 (strain McN13), and G_4P_6 (strain ST3) (9, 12). We subsequently showed by serologic and sequence analyses that prototype strain 116E belongs to serotype G_9 and genotype P_{11} and has not been previously isolated from humans (4). Since the prototype strains for serotypes G_9 (human isolate WI61, serotype G_9P_8) and P_{11} (bovine isolate B223, serotype $G_{10}P_{11}$) have P and G types, respectively, that are different from those of 116E, it was suggested that the latter strain may be a reassortant (7).

We have recently extended these results and shown that strains related to prototype 116E are present in five of six New Delhi hospitals (3). In this report, we describe the complete characterization of rotavirus strains isolated from newborns at six government hospitals in New Delhi between 1986 and 1988 and 1992 and 1993, as well as isolates previously collected in a longitudinal study by using G and P genotyping by reverse transcription (RT)-PCR, electropherotyping, subgrouping analysis, and nucleotide sequencing.

To screen for genotype P_{11} rotavirus strains, a specific primer, ND2, that is complementary to nucleotides (nt) 116 to 133 of the strain 116E VP4 gene was synthesized and added to a one-amplification RT-PCR system that detects strains with

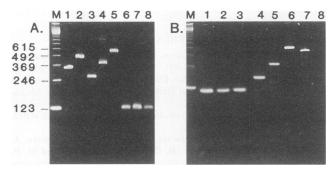


FIG. 1. RT-PCR typing of rotavirus strains. Rotavirus doublestranded RNA was extracted from cell lysates or fecal specimens, and 5 µl of the eluate was analyzed (8). (A) P typing. Lanes: M, markers (123-bp ladder; Gibco BRL, Long Island, N.Y.; marker molecular sizes are indicated on the left in base pairs); 1 to 5, products amplified from double-stranded RNA from human rotavirus strains possessing P types 8 (lane 1, strain Wa), 4 (lane 2, strain DS-1), 6 (lane 3, strain M37), 9 (lane 4, strain K8), 10 (lane 5, strain M37), and 11 (lane 6, strain 116E); 7 to 8, products from double-stranded RNA extracted from cultureadapted strains (lane 7, strain 113E; lane 8, strain 218D). (B) G typing. Lanes: M, markers (123-bp ladder); 1 to 3, products amplified from double-stranded RNA of human rotavirus G serotype 9 (lane 1, strain 116E; lane 2, strain F45; lane 3, strain WI61); 4 to 8, products amplified from field isolates from Costa Rica possessing serotype G1 to G₄ specificities as determined by enzyme-linked immunosorbent assay serotyping with monoclonal antibodies.

^{*} Corresponding author. Phone: (404) 639-3577. Fax: (404) 639-3645.

Hospital	Year(s)	No. of strains tested	No. of strains with the following G genotype, P genotype, and subgroup ^{b}							
			$\overline{G_2P_4SG_1}$	$G_9P_{11}SG_{11}$	$G_9P_{11}SG_2^c$	$G_9P_6SG_{11}$	G ₉ P ₆ SG ₂ ^c	$G_3P_{11}SG_{11}$	DT ¹	Other
Α	1986-1988	21 ^{<i>f</i>}	1	18	0	0	0	2	0	0
А	1992	7	0	6	1	0	0	0	0	1
Α	1993	9	0	0	0	9	1	0	0	0
В	1992	7	0	4	0	0	0	0	3	0
В	1993	3	0	2	1	1	0	0	0	0
С	1993	16	0	12	0	0	0	0	4	0
D	1993	5	0	5	0	0	0	0	0	0
Е		7	0	5	1	0	0	0	1	1
Totals		75	1	49	3	9	1	2	8	2

" Subgroups of rotavirus antigen-positive fecal specimens, as determined by the Rotaclone test (Cambridge Biosciences, Cambridge, Mass.), were determined as described previously, with antibodies provided by H. Greenberg (10).

^b Of the 67 strains tested, 64 had similar long electropherotypes and the lone SG₁ strain had a short electropherotype.

^c SG₂ indicates that the strain could not be subgrouped.

 d DT, dual PCR type, i.e., presence of two DNA products, consistent with the presence of two different genotypes. P₆P₁₁ dual types were verified by restriction endonuclease analysis of both products (data not shown).

^c One strain from hospital A was non-G-nongenotypeable, while the P-typing reaction for one isolate from hospital E was omitted.

 f Culture-adapted strains; all of the other strains tested were isolated directly from fecal specimens.

genotypes P_4 (e.g., strain DS-1 like), P_6 (e.g., strain M37 like), P_8 (e.g., strain Wa like), P_9 (e.g., strain K8 like), and P_{10} (e.g., strain 69M like). The procedures for RNA extraction and amplification were identical to those described previously, except that 40 PCR cycles were used and the extension time was 3 min at 72°C (8).

For G genotyping (detection of VP7 genes by RT-PCR), a consensus primer, 9 con 1, whose sequence is conserved among the VP7 genes of serotypes G_1 to G_4 and G_9 (GenBank accession numbers, K02033, M11164, U04350, M21650, and L14072) and five genotype-specific primers complementary to variable regions of the VP7 genes of the same serotypes were synthesized and used in an RT-PCR system analogous to that used for P genotyping. The nucleotide positions, strain and serotype specificities, polarities (plus or minus), and sequences (from 5' to 3') of type-specific complementary primers 9T1-1, 9T1-2, 9T-3P, 9T-4, and 9T-9B are as follows: 9 Con 1, nt 37 to 56, Wa, plus sense, and TAGCTCCTTTTAATGTATGG; 9T1-1, nt 176 to 195, Wa, G1, minus sense, and TCTTGT CAAAGCAAATAATG; 9T1-2, nt 262 to 281, S2, G2, minus sense, and GTTAGAAATGATTCTCCACT; 9T-3P, nt 484 to 503, 107E1B, G₃, minus sense, and GTCCAGTTGCAGTGT TAGC; 9T-4, nt 423 to 440, ST3, G₄, minus sense, and GGGTCGATGGAAAATTCT; 9T-9B, nt 131 to 147, 116E, G₉, minus sense, and TATAAAGTCCATTGCAC. The expected molecular sizes of the RT-PCR products of the primerpair consisting of 9 con 1 and 9T1-1, 9T1-2, 9T-3P, 9T-4, or 9T-9B were 158, 244, 466, 403, or 110 bp, respectively. The strain specificities of the P₁₁- and G₉-specific RT-PCR primer pairs are shown in Fig. 1A and B.

A summary of the characteristics of 75 strains from newborns is presented in Table 1. The type designations of the strains identified here by genotyping conform to the suggestions of Estes and Cohen (6). G and/or P serotype designations have only been given to strains analyzed by cross-neutralization tests. Nucleotide sequence analysis of variable regions of the VP4 and VP7 genes of two strains from newborns demonstrated that our RT-PCR typing method accurately predicted their G and P genotypes (7).

Recently, we reported the isolation of novel rotavirus strains from newborns with serotype G_9 and genotype P_{11} specificity (4). These findings have since been extended to five other hospitals in New Delhi where strains related to prototype strain 116E were detected (3). In this report, we more completely characterized all of the strains isolated in the 1993 study, as well as strains isolated from two of the hospital nurseries between 1986 and 1992. Our results strongly suggest that genotype P₁₁ strains are common in New Delhi. A second novel strain from newborns-genotype G₉P₆-identified at two of the hospitals completely replaced the genotype G_9P_{11} strains found in hospital A between 1992 and 1993. Although the genotype P_6 VP4 gene of these strains is related to those of previously identified strains from newborns (e.g., Venezuelan strain M37, serotype G_1P_6), genotype G_9P_6 strains have not been isolated before (9). It is possible that these strains arose by reassortment, since we found that isolates with dual types (types P_6 and P_{11} and G_1 and G_9) were isolated from the same infants in hospital B in 1992, and we subsequently identified genotypes $G_9 P_6$ and $G_9 P_{11}$ cocirculating in 1993 at hospital B. Regardless of their origin, the isolation of three different, unique rotaviruses from newborns in New Delhi and Bangalore suggests that additional surveillance should be conducted to determine the prevalent strains from newborns in other areas of India and to investigate if any of these novel rotaviruses are common in children with diarrhea (3, 5). These studies will be important to assess the possible utility of such strains as vaccine candidates.

We thank Ann Mather for help in editing the manuscript, Brian Holloway of the CDC Molecular Biology Core Facility for primer synthesis, and Taka Hoshino for providing several rotavirus strains.

This study was supported in part by a grant from the INDO-U.S. Vaccine Action Program.

REFERENCES

- Bhan, M. K., J. F. Lew, S. Sazawal, B. K. Das, J. R. Gentsch, and R. I. Glass. 1993. Protection conferred by neonatal rotavirus infection against subsequent diarrhea. J. Infect. Dis. 168:282–287.
- Bishop, R. F., G. L. Barnes, E. Cipriani, and J. S. Lund. 1983. Clinical immunity after neonatal rotavirus infection: a prospective longitudinal study in young children. N. Engl. J. Med. 309:72–76.
- Cicirello, H., B. K. Das, A. Gupta, M. K. Bhan, J. R. Gentsch, R. Kumar, and R. I. Glass. High prevalence of rotavirus infection among neonates born at hospitals in Delhi, India: predisposition of newborns for infection with unusual rotavirus strains. Pediatr. Infect. Dis. J., in press.
- Das, B. K., J. R. Gentsch, Y. Hoshino, S.-I. Ishida, O. Nakagomi, M. K. Bhan, R. Kumar, and R. I. Glass. 1993. Characterization of

the G serotype and genogroup of New Delhi newborn rotavirus strain 116E. Virology **197:**99–107.

- Das, M., S. J. Dunn, G. N. Woode, H. B. Greenberg, and C. D. Rao. 1993. Both surface proteins (VP4 and VP7) of an asymptomatic neonatal rotavirus strain (I321) have high levels of sequence identity with the homologous proteins of a serotype 10 bovine rotavirus. Virology 194:374–379.
- 6. Estes, M. K., and J. Cohen. 1989. Rotavirus gene structure and function. Microbiol. Rev. 53:410-449.
- 7. Gentsch, J., B. K. Das, B. Jiang, M. K. Bhan, and R. I. Glass. 1993. Similarity of the VP4 protein of human rotavirus strain 116E to that of the bovine B223 strain. Virology **194**:424–430.
- Gentsch, J. R., R. I. Glass, P. Woods, V. Gouvea, M. Gorziglia, J. Flores, B. K. Das, and M. K. Bhan. 1992. Identification of group A rotavirus gene 4 types by polymerase chain reaction. J. Clin. Microbiol. 30:1365–1373.
- Gorziglia, M., G. Larralde, A. Z. Kapikian, and R. M. Chanock. 1990. Antigenic relationships among human rotaviruses as determined by outer capsid protein VP4. Proc. Natl. Acad. Sci. USA 87:7155-7159.
- Greenberg, H., V. McAuliffe, J. Valdesuso, R. Wyatt, J. Flores, A. Kalica, Y. Hoshino, and N. Singh. 1983. Serological analysis of the subgroup protein of rotavirus using monoclonal antibodies. Infect. Immun. 39:91–99.
- 11. Hoshino, Y., M. M. Sereno, K. Midthun, J. Flores, A. Z. Kapikian, and R. M. Chanock. 1985. Independent segregation of two anti-

genic specificities (VP3 and VP7) involved in neutralization of rotavirus infectivity. Proc. Natl. Acad. Sci. USA 82:8701-8704.

- Hoshino, Y., R. G. Wyatt, J. Flores, K. Midthun, and A. Z. Kapikian. 1985. Serotypic characterization of rotaviruses derived from asymptomatic human neonatal infections. J. Clin. Microbiol. 21:425–430.
- Rasool, N. B. G., G. Larralde, and M. I. Gorziglia. 1993. Determination of human rotavirus VP4 using serotype-specific cDNA probes. Arch. Virol. 133:275–282.
- Snodgrass, D. R., Y. Hoshino, T. A. Fitzgerald, M. Smith, G. F. Browning, and M. Gorziglia. 1992. Identification of four VP4 serological types (P serotypes) of bovine rotavirus using viral reassortants. J. Gen. Virol. 73:2319–2325.
- Steele, A., D. Garcia, J. Sears, G. Gerna, O. Nakagomi, and J. Flores. 1993. Distribution of VP4 gene alleles in human rotaviruses by using probes to the hyperdivergent region of the VP4 gene. J. Clin. Microbiol. 31:1735–1740.
- Taniguchi, K., T. Urasawa, Y. Morita, H. B. Greenberg, and S. Urasawa. 1987. Direct serotyping of human rotavirus in stools using serotype 1-, 2-, 3-, and 4-specific monoclonal antibodies to VP7. J. Infect. Dis. 155:1159–1166.
- Woods, P. A., J. Gentsch, V. Gouvea, L. Mata, A. Simhon, M. Santosham, Z.-S. Bai, S. Urasawa, and R. I. Glass. 1992. Distribution of serotypes of human rotavirus in different populations. J. Clin. Microbiol. 30:781–785.