Typhoid and paratyphoid fever

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Typhoid fever is estimated to have caused 21.6 million illnesses and 216 500 deaths globally in 2000, affecting all ages. There is also one case of paratyphoid fever for every four of typhoid. The global emergence of multidrug-resistant strains and of strains with reduced susceptibility to fluoroquinolones is of great concern. We discuss the occurrence of poor clinical response to fluoroquinolones despite disc sensitivity. Developments are being made in our understanding of the molecular pathogenesis, and genomic and proteomic studies reveal the possibility of new targets for diagnosis and treatment. Further, we review guidelines for use of diagnostic tests and for selection of antimicrobials in varying clinical situations. The importance of safe water, sanitation, and immunisation in the presence of increasing antibiotic resistance is paramount. Routine immunisation of school-age children with Vi or Ty21a vaccine is recommended for countries endemic for typhoid. Vi vaccine should be used for 2–5 year-old children in highly endemic settings.

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Typhoid fever is a systemic infection caused by *Salmonella enterica* serotype typhi (*S typhi*). A very similar but often less severe disease is caused by *S paratyphi* A, B, and sometimes C. *S typhi*, a highly adapted human-specific pathogen that evolved about 50 000 years ago,¹ has remarkable mechanisms for persistence in its host.²

The incidence of typhoid fever has declined greatly with the provision of clean water and good sewage systems in Europe and the USA since the early 20th century.3 but the disease remains a serious public-health problem in developing countries.4 The advent of chloramphenicol treatment changed the perception of typhoid fever from a severe, often fatal, disease to a common, readily manageable infection.5 Outbreaks of chloramphenicol-resistant typhoid were reported in 1972.6 At this time, the isolates were still sensitive to cotrimoxazole and ampicillin or amoxycillin. In the late 1980s and 1990s, outbreaks of typhoid caused by organisms resistant to chloramphenicol, co-trimoxazole, ampicillin, and amoxicillin were reported.6 Currently, fluoroquinolones and third-generation cephalosporins are the drugs of choice for treatment of typhoid fever, but decreased susceptibility to these antimicrobials has been reported.7-10 There is an urgent need to keep the possible emergence of untreatable strains to a minimum, by prudent use of existing drugs and by resisting the temptation to use yet more antimicrobials.

Disease burden

Typhoid is estimated to have caused 21-6 million illnesses and 216 500 deaths globally in 2000.⁴ The incidence of typhoid was high (>100 cases per 100 000 population per year) in south-central Asia, southeast Asia, and possibly southern Africa, medium (10–100 cases per 100 000) in the rest of Asia, Africa, Latin America, and Oceania, except for Australia and New Zealand, and low in the other parts of the world (<10 cases per 100 000). These estimates are based on blood-culture-positive cases in 22 population-based studies, many of which¹¹⁻²¹ were done after publication of the previous global estimates (table 1).²² These estimates have important limitations, however. They are based on data from only a few countries, with only one study providing data from Africa. Placebo groups from typhoid vaccine trials were included, and vaccine trials are usually done in areas with high disease burden. Some assumptions used also merit consideration. An adjustment factor of two was used to account for the low sensitivity of blood culture. A conservative case-fatality rate of 1% was chosen on the basis of conservative estimates of hospital based typhoid fever studies. Further, there are important consequences of typhoid fever related to absenteeism from work and need for admission, which were not considered.

Where typhoid is endemic, most cases in health facilities are children aged 5–19 years and young adults. Recent population based studies from India, Indonesia, and Vietnam suggest that in some settings typhoid fever is also common in 1–5 year-old children (table 2).^{15,17,19,20} Data from hospital-based studies in Bangladesh and Thailand^{23,24} support these findings. Surveillance data

Search strategy and selection criteria

We searched MEDLINE, PubMed, EMBASE, and the Cochrane Library from 1995 to 2004 to identify recent studies on the causative organism, disease burden, transmission, risk factors, pathogenesis, diagnosis, treatment, and prevention of typhoid and paratyphoid fevers. The main search terms were "typhoid fever", "paratyphoid fever", "enteric fever", and "Salmonella" in combination with "Typhi" or "Paratyphi". No language restrictions were applied. We mainly selected articles in the past 5 years, but did not exclude commonly referenced and highly regarded older reports. We also searched the reference lists of articles identified by this search strategy. Of all articles identified, 1253 were reviewed for inclusion by two of us (RB and SB). Selection criteria included a judgment about the novelty and importance of studies and their relevance for the well-informed general clinician. Several review articles were included because they provide comprehensive overviews that are beyond the scope of this Seminar

	Type of surveillance	Age (years)	n	Incidence per 100 000 per year
Urban				
Chile11	Passive	5-19	21904	107
Chile ¹²	Passive	5-19	27 305	227
Chile ¹³	Passive	5-19	10 302	91
China ¹⁴	Passive	5-19	65 984	22
Indonesia ¹⁵	Passive	3-44	10 268	810
India ¹⁶	Passive	0-40	Not reported	290
India17	Active	0-40	6454	980
South Africa ¹⁸	Active	5-16	11691	850
Rural				
Vietnam ¹⁹	Passive	0->50	28 239	198
Vietnam ²⁰	Active	2-4	6017	414
Nepal ²¹	Active	5-44	3450	655

Table 1: Incidence of culture-proven typhoid fever from population-based studies in urban and rural settings, published after 1986

from the USA indicate that the proportion of cases of typhoid is constant over the first 25 years of life.⁴ Typhoid fever is more common in urban than in rural areas, but studies from two rural settings in Nepal and Vietnam also found a high disease burden.¹⁹⁻²¹ Most cases in developed countries arise in travellers, but domestically acquired disease is still reported.^{25,26} A total of 1393 typhoid cases were reported between 1994 and 1999 in the USA, 74% of which were related to travel and the rest of which were acquired domestically; 7% of total cases were part of recognised outbreaks.²⁷

Paratyphoid fever is estimated to have caused an additional $5 \cdot 4$ million illnesses in 2000.⁴ This number is based on an estimated one case of paratyphoid fever for every four cases of typhoid fever (table 3).^{11,12,14-16,19,28} Studies from India and Nepal suggest that in some settings and times, paratyphoid fever caused by *S paratyphi A* can contribute up to half of all cases of typhoid fever.^{17, 29-31}

About 10% of people recovering from untreated typhoid fever may excrete *S typhi* in the stools for at least 3 months. Between 1% and 5% of typhoid patients become chronic carriers (defined as excretion of *S typhi* in urine or stools for more than one year), and the rate is

	Type of surveillance	Age (years)	n	Incidence per 100 000 per year
Urban				
Indonesia ¹⁵	Passive	3-6	1592	1307
		7-19	4711	1172
		20-44	3965	182
India ¹⁷	Active	0-4	1027	2730
		5-19	2743	1170
		20-40	2684	110
Rural				
Vietnam ¹⁹	Passive	2-4	Not reported	358
		5-9		531
		10-14		429
		15-19		153
		20-29		149
		30-39		51
Vietnam ²⁰	Active	2-4	6017	414

Table 2: Age-specific incidence of culture-proven typhoid fever from population-based studies

higher for women, those older than 50 years, and patients with schistosomiasis, cholelithiasis, carcinoma of the gall bladder, and other gastrointestinal malignancies.³² Most chronic carriers are asymptomatic and almost a quarter may have had no history of typhoid fever.

Transmission and risk factors

People are the only natural host and reservoir for S typhi. The pathogen can survive for days in groundwater, pondwater, or seawater, and for months in contaminated eggs and frozen oysters.^{33–36} The infectious dose is between 1000 and 1 million organisms given orally.37 The infection is transmitted by ingestion of food or water contaminated with faeces. Established risk factors are contaminated water supply, eating ice cream, flavoured iced drinks or food from street vendors, and raw fruit and vegetables grown in fields fertilised with sewage.^{32,38-42} Other reported risk factors include a history of contact with other patients before illness, not using soap for washing hands, poor housing, and past evidence of infection with Helicobacter pylori^{28,42-44} The mechanism of increased risk of typhoid in individuals with chronic H pylori infection is postulated to be reduced gastric acidity. Factors within the household (eg, poor personal hygiene and housing) might be more important risk factors for typhoid, whereas factors outside the household (eg, food from street vendors, flooding) are more important for paratyphoid fever.28 A possible reason proposed for this difference was the higher infective dose necessary for paratyphoid, which is more likely to be present in food from street vendors. Although chronic typhoid carriers are important for survival of the pathogen, they are less important as a direct source of infection in endemic areas than contaminated water or food.³²⁻⁴⁶ In the USA, up to 30% of infections are due to exposure from previously or newly diagnosed chronic carriers.47

Involvement of host genetic factors has also been implicated in the pathogenesis of typhoid fever. Work on typhoid fever patients in Vietnam has suggested an important role of HLA-linked genes in governing susceptibility or resistance to this infection. HLA-DRB1*0301/6/8, HLA-DQB1*0201-3, and tumour necrosis factor a (TNFA*2-308) were associated with susceptibility to typhoid fever, and HLA-DRQB1*04 and HLA-DQB1*0401/2 and TNFA*1(-308) were associated with lower risk.⁴⁸ HLA-DRB1*12 is associated with protection against complicated typhoid fever.⁴⁹ Similar studies in other ethnic groups might help in defining the common HLA genes and the locus of susceptibility.

The pathogen

The causative organism of typhoid fever, belongs to the family Enterobacteriaciae. Most *S typhi* isolates from typhoid fever cases have a polysaccharide capsule (Vi), which is associated with increased infectiousness and virulence, but Vi-negative strains can also cause the

2-4	6017	22	18.7
			10.7
5-19	65 984	5	4.6
3-44	10 268	187	4.3
5-19	21 904	26 (paratyphi B)	4.1
5-19	27 305	37 (paratyphi B)	6-2
1-80	Not reported	Not reported	3.4
0-40	6454	372	2.6
	3-44 5-19 5-19 1-80 0-40	3-44 10 268 5-19 21 904 5-19 27 305 1-80 Not reported 0-40 6454	3-44 10 268 187 5-19 21 904 26 (paratyphi B) 5-19 27 305 37 (paratyphi B) 1-80 Not reported Not reported 0-40 6454 372

disease.⁵⁰ Molecular typing techniques show that many different strains are in circulation in endemic areas but outbreaks are related to a few strains.⁵¹⁻⁵³ Genetic variation between isolates has been related to clinical outcome. Multidrug resistant isolates, for instance, might be more virulent than sensitive strains,⁵⁴ and isolates from fatal cases might be different from background strains.⁵⁵

The complete DNA sequence of a multidrug resistant isolate of S typhi isolate, CT18, shows the number of predicted genes to be 4599.56 The core region conserved between species (70-80% of the chromosome) could represent a gene repertoire associated with basic functions in the Enterobacteriaceae, such as intestinal colonisation, survival in the environment, and transmission. Scattered along this conserved core are single genes or groups of genes that are specific to S enterica or indeed to serotype typhi. Up to ten Salmonella pathogenicity islands have been identified. All types of S enterica have two large clusters of genes known as Salmonella pathogenicity island 1 (SPI-1) and SPI-2, that facilitate invasion of and survival inside host cells. The S typhi genome also contains SPI-7, which has genes that code for Vi polysaccharide production as well as many other genes of unknown function. There are more than 200 pseudogenes that have clearly been inactivated by simple point mutations or frameshifts, 145 of them are present as active genes in S typhimurium. Significantly, S typhimurium causes a different disease in people, and has a wider host range than S typhi.56.57 Two plasmids are present-a larger transmissible one called pHCM1 that encodes several drug-resistance determinants, and a smaller one, pHCM2, which is phenotypically cryptic.57,58

Sequence and microarray analysis of the genome of *S paratypi A* indicates that it is similar to that of *S typhi*, but that it might have a more recent evolutionary origin. It has accumulated 173 pseudogenes by comparison with about 210 of the *S typhi*.⁵⁹

Pathogenesis

S typhi, unlike *S typhimurium*, avoids triggering of an early inflammatory response in the gut of the human host, using a stealth approach to allow colonisation of

deeper tissues of the body.^{2,60} The description of typhoid fever pathogenesis we provide is based largely on the murine model in which *S typhimurium* causes a systemic infection similar to typhoid.

S typhi probably invades the gut mucosa in the terminal ileum through specialised antigen-sampling cells, known as M-cells, which overlie gut-associated tissue,⁶¹ through enterocytes, or via a paracelluar route.⁶² The bacteria adhere to the intestinal mucosa in the terminal ileum through interaction with an epithelial receptor, the cystic fibrosis transmembrane conductance regulator protein.⁶³ A key, early step in the infectious process is the induction of intestinal epithelial cells to increase membrane receptor levels, with enhanced bacterial ingestion and submucosal translocation.⁶⁴

Studies of non-typhoidal *Salmonella* spp indicate that invasion of non-phagocytic epithelial cells occurs through the activity of bacterial proteins delivered into host cells by a type III secretion system, encoded on SPI-1.⁶⁻⁶⁷ The formation of an intracellular replicative niche requires further activity of SPI-1,⁶⁸ which is downregulated after a few hours of invasion,⁶⁹ followed by activity of a second type-III secretion system, encoded on SPI-2.^{70,71}

The bacterial invasion leads to infiltration of peripheral blood leucocytes into the lamina propria. This infiltration is mediated by cytokine secretion from epithelial cells induced by bacterial lipopolysaccharide, a component of the cell wall of gram-negative bacteria. Lipopolysaccharide activates transcription factors in lymphocytes by signalling through a mammalian Toll pathway known as Toll-like receptor 4 complex.^{72,73} Invading bacteria are taken up by macrophages, which undergo salmonella-induced caspase-1 mediated apoptosis.⁵⁷

The bacteria reach the intestinal lymphoid tissue, and are drained into mesenteric nodes, the thoracic duct, and then the general circulation. This primary bacteraemia results in the organism reaching the liver, spleen, bone marrow, and other parts of the reticuloendothelial system within 24 h of their ingestion, where they survive and replicate in cells of monocytic lineage.⁷⁴ Bacteria are shed back into the bloodstream, marking the onset of the clinical illness (after an incubation period of 8–14 days) during which a low level of bacteraemia is sustained.



Figure: Global distribution of antimicrobial resistance in S typhi (1990–2004) Adapted from Parry and colleagues⁵⁶ and updated on basis of data from past 3 years.

A possible underlying molecular mechanism for persistence of salmonella in the host, as seen in typhoid carrier state, has been proposed.² Differences in mouse susceptibility to S typhimurium have been linked to the particular allele of the Nramp1 gene expressed on their macrophages.75 Nramp1 is involved in controlling growth exponential of salmonella in the reticuloendothelial system during the early stages of infection. Mice expressing the wildtype Nramp1 allele did not die after oral inoculation with S typhimurium, but became uniformly persistently infected as did chronic typhoid carriers.76 Salmonella persisted in small numbers, mainly in the macrophages of mesenteric lymph nodes or spleen (or both), despite a robust antibody response. Reactivation of intracellular salmonella and systemic spread could be accomplished by administering antibodies to neutralise interferon γ , suggesting that host cytokine interferon γ is important for suppression of salmonella replication and disease.

Antimicrobial resistance

In late 1987, there was an outbreak of typhoid fever in China caused by strains resistant to all the first line antimicrobials (ampicillin, co-trimoxazole, and chloramphenicol).⁷⁷ During 1989–1990, there were reports of similar *S typhi* strains from India, Pakistan, and the Arabian Gulf.⁷⁸⁻⁸⁰ Such multidrug resistant typhoid is now reported from many parts of the world (figure).^{38,39,53,54,81-94} Multiple antimicrobial resistance is

mediated through pHCM1 plasmid,^{95,96} which has 99% sequence identity with plasmid R27,⁹⁷ an *inc* H1 plasmid.⁹⁵ For at least 74% of the isolates in Vietnam, endemic and epidemic multidrug-resistant typhoid fever was due to one or two clones of *S typhi* carrying a single resistance plasmid.⁹⁸

A decreasing trend has been reported in the isolation of multidrug-resistant *S typhi* strains from southern Asia. In a retrospective analysis of blood cultures in a diarrhoea treatment centre in Bangladesh,⁹⁹ the rate of multidrug-resistant *S typhi* was 0.3% of all blood cultures in 1990, a peak of 3.2% in 1994, and 1.0% in 1996. The isolation rate of susceptible *S typhi* remained remarkably unchanged (3.3%) during the study. More than 90% of *S typhi* strains isolated in 1999 from typhoid patients in a hospital in New Delhi were sensitive to chloramphenicol.¹⁰⁰ In another study from India,¹⁰¹ all *S typhi* isolates were resistant to chloramphenicol in 1991, but chloramphenicol resistance fell to less than 20% by 2000.¹⁰¹

After emergence of multidrug resistance, fluoroquinolones (eg, ciprofloxacin, ofloxacin) became the treatment of choice for typhoid fever. Isolates fully susceptible to ciprofloxacin by disc testing typically have a ciprofloxacin minimum inhibitory concentration of less than 0.03 mg/L and are invariably also susceptible to the first generation quinolone nalidixic acid. A population of isolates exists with a minimum inhibitory concentration of 0.125-1.0 mg/L that seems to be susceptible to ciprofloxacin by disc testing but is associated with clinical failure and is resistant to nalidixic acid (NAR strains); such isolates have been reported from several countries (figure).^{25,83,84,94,102-108} Single point mutations (Ser-83 to Phe, Asp-87 to Asn, Ser-83 to Tyr and Asp-87 to Gly) in the gyrA gene of S typhi and S paratyphi cause this reduced susceptibility to ciprofloxacin. 103,107,109,110 Table 4 summarises key features of NAR isolates.107,108,111,112 The National Committee for Clinical Laboratory Standards currently recommends testing of extraintestinal salmonella isolates for nalidixic acid resistance as a marker for reduced fluoroquinolone susceptibility.113 This testing is, however, not foolproof-in Europe, 11% of S typhi strains with decreased ciprofloxacin susceptibility were sensitive to nalidixic acid.¹¹⁴ Multidrug resistance as well as reduced susceptibility to ciprofloxacin has also been reported for S paratyphi A.¹¹⁵

Laboratory diagnosis

Confirmation of typhoid or paratyphoid fever requires isolation of S typhi or S paratyphi, respectively, from blood, bone marrow, stool, or duodenal fluid. Cultures from skin above rose spots, buffy coats, and blood clots treated with streptokinase have been used.116-119 Bonemarrow aspirate culture is positive in 80-95% of typhoid patients.^{116-118,120} Culturing more than 10 mL of blood is necessary to match the positivity rate with that of 1 mL of bone marrow. Bone-marrow cultures are useful for lengthy illness and antibiotic treatment.118,121,122 Blood culture is the mainstay of diagnosis. Using standard broth cultures, salmonella is isolated in 30-90% of patients with clinical typhoid.^{118,121,122} Sensitivity decreases with increasing duration of fever.¹¹⁸ The volume of blood and the ratio of blood to broth determines blood culture yield: 10-15 mL of blood is necessary to maintain an optimum ratio of 1 to 12. 2-4 mL of blood is sufficient in children who have a greater concentration of bacteria in their blood.^{122,123} Failure to maintain ambient temperatures of 15-40°C during specimen transportation, inappropriate laboratory methods, and antimicrobials compromise the yield.

Stool isolation of *S* typhi alone is insufficient for diagnosis and only marginally improves diagnosis by blood culture. However, it is confirmatory for carrier detection. Serological tests based on agglutination of Vi antigens have 70–80% sensitivity and up to 95% specificity in identifying carriers of *S* typhi.¹²⁴

The Widal test identifies the agglutinating antibodies against the O (somatic) and H (flagellar) *S typhi* antigens, which appear a week to 10 days after disease onset. The sensitivity, specificity, and predictive values reported from different centres vary because of sharing of O and H antigens and cross-reacting epitopes with other Enterobacteriaceae.¹²⁵ The high number of false-positive and false-negative Widal test results limit its clinical usefulness. To make a diagnosis, results from a single acute sample should be interpreted against the

	Proportion of NAR strains	% patients who failed on fluoroquinolone treatment	% cases with faecal carriage during early convalescence
UK ¹¹¹	23% (42/179)	24% (10/42)	Not reported
Canada ¹¹²	33% (7/21)	80% (4/5)	Not reported
India ¹⁰⁷	47% (82/174)	27% (22/82)	Not reported
Vietnam ¹⁰⁸	9% (46/504)	24%* (11/46)	21%† (8/39)

*By comparison, 5% (22/458) of nalidixic-acid-sensitive strains had clinical failure on ofloxacin. †By comparison, 2% (7/383) of nalidixic-acid-sensitive strains had faecal carriage during early convalescence.

Table 4: Key features of nalidixic-acid-resistant (NAR) strains of S typhi

appropriate local cut-off values or there should be a four-fold rise in the antibody titres between convalescent and acute sera. $^{\rm 126,127}$

Some simple inexpensive rapid serological diagnostic tests for typhoid fever are available (table 5).¹²⁸⁻¹³³ In an evaluation of three commercial kits, the sensitivity and specificity for identifying blood-culture-positive cases of typhoid fever was 89% and 53% for multi-test dip-sticks (PANBIO INDX, Baltimore, MD, USA), 79% and 89% for typhidot (Malaysian Biodiagnostic Research SDN BHD, Malaysia), 78% and 89% for tubex test (IDL Bideh, Solletuna, Sweden) as compared with 64% and 76% for Widal.¹²⁸

The understanding of genome sequence of *S typhi* might now result in identification of specific antibodies to fimbrial and other antigens.⁵⁷ Urinary Vi antigen detection by ELISA within the first febrile week shows promise, but positivity in brucellosis patients is an obstacle to further development.¹³⁴ DNA probes and PCR-based tests to detect flagellar genes¹³⁵ are not routinely useful in developing countries, but they are of value in surveillance and research.

Clinical features

The clinical presentation of typhoid fever is very variable, ranging from fever with little other morbidity to marked toxaemia and associated complications involving many systems. In endemic regions, diagnosis can be missed because of non-specific features like diarrhoea and vomiting, or predominant respiratory symptoms.^{17,19} Because of this variable profile, the disease has to be differentiated from tuberculosis, brucellosis, sepsis due to other bacterial pathogens, infectious mononucleosis, anicteric hepatitis, and, infrequently, from leukaemia and lymphoma. In the days when chloramphenicol was clinically effective, typhoid fever was largely treated on an outpatient basis, but with the emergence of multidrugresistant strains and decreased susceptibility to fluoroquinolones, it has acquired a profile of severe infectious disease that often requires inpatient care. In developing countries, admission has substantial consequences for poor families.

An average case of acute non-complicated typhoid fever has an incubation period of 10–14 days and is usually associated with prolonged low-grade fever, dull frontal headache, malaise, myalgia, a dry bronchitic

Tubex Styphi 09 antigenImmunoglobulin MVietnam129* N 87% 76% 76% 77% 78% 84% 84% 77% 84% 77% 84% 98% 99% 99% 65% 65% 65% Dipstick StyphiImmunoglobulin MVietnam129* $R M$ 48% 98% 98% 98% 99% 99% 65% 66% Lipopolysacchride antigen GImmunoglobulin GIndonesia139† $R M$ 65.3% $R M$ 100% 100% 61% 61% Immunoglobulin MIndonesia139† $R M$ 65.3% $R M$ 100% 100% 61% 66% Typhidot Dot-ELISA Dot-ELISAImmunoglobulins $G \ r M$ Vietnam128† $R M$ 79% 89% 94% 77% 88% 83.3% 83.3% 66.6% Typhidot M Dot-ELISAImmunoglobulins $G \ r M$ Pakistan 132* 94% 77% 80% 83.3% 83.3% 100% Typhidot M Dot-ELISAImmunoglobulins $G \ r M$ Pakistan 132* 73% 73% 89% 95% 55% Typhidot M Dot-ELISAMPakistan 132* 73% 73% 89% 95% 55%		Antibodies	Country	Sensitivity	Specificity	Positive predictive value	Negative predictive valu
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studies only included where the gold standards were positive blood or bone-marrow culture (*) or positive blood culture (†). OMP=outer membrane protein.	tudies only included where the go	old standards were positive bl	ood or bone-marrow cu	lture (*) or positiv	e blood culture (†). OMP=outer membrane proteir	ı.

cough, anorexia, and nausea. The fever might rise progressively in a stepwise manner to become persistent and high grade by the second week of illness.⁴⁶ Continuous high-grade fever can continue for up to 4 weeks if left untreated, followed by a return to normal temperature. Malaise and lethargy can continue for a couple of months later.

Although not present consistently, relative bradycardia at the peak of high fever is an indicator of typhoid fever. Coated tongue, alteration of bowel habits varying from constipation in adults to diarrhoea in children, tender abdomen, hepatomegaly, and splenomegaly are often present.^{96,136} Small erythematous maculopapular lesions (rose spots) are seen on the back, arms, and legs in up to a quarter of cases late in the first week of fever, particularly in fair-skinned people. Rhonchi and scattered crepts might be heard on chest auscultation. Liver involvement is common with elevated concentrations of serum bilirubin and alanine transferase; in endemic areas, typhoid fever should be a differential diagnosis for a patient with fever and jaundice.¹³⁷

Neonatal typhoid fever resulting from vertical transmission during late pregnancy is a rare but often life-threatening illness. It usually begins during 3 days of delivery with fever, vomiting, diarrhoea, and abdominal distension. There might be significant hepatomegaly and jaundice. Seizures can occur. It could also present as asymptomatic persistent excretion. In children younger than 5 years, typhoid fever can be milder and can mimic a viral syndrome. The rate of severe complications is lower than at later ages.^{17,188}

Overall, about 10–15% of patients develop severe disease. Factors affecting severity include duration of

illness before therapy, choice of antimicrobial therapy, strain virulence, inoculum size, previous exposure or vaccination, and other host factors such as HLA type, AIDS or other immune suppression, or antacid consumption. The commonest complications are gastrointestinal bleeding, intestinal perforation and typhoid encephalopathy.^{46,96} A rapid drop in temperature during the later period of the illness suggests intestinal bleeding or perforation. This drop is usually followed by a rise in temperature after a few hours as peritonitis develops. Gastrointestinal bleeding occurs in 10-20% of cases due to erosion of the Peyer's patch into an intestinal vessel and is usually restricted to either occult blood in stool or malaena. On colonoscopy, terminal ileum is the commonest site involved followed by the ileocaecal valve, the ascending then the transverse colon.139 Multiple punched out ulcers with slightly elevated margins are evident.

Intestinal perforation occurs in 1–3% of cases in hospital; the commonest site is the ileum.¹⁴⁰ Being male, leucopenia, inadequate treatment before admission, and short duration of symptoms are significant predictors of perforation.¹⁴¹ Intermittent confusion, insomnia, and dizziness are reported in 3–10% of cases and these symptoms are associated with high case fatality. Some patients manifest neuropsychiatric symptoms such as picking at bedclothes or imaginary objects, which are described as muttering delirium or coma vigil. Deep coma is rare. Typhoid meningitis, encephalomyelitis, Guillain-Barré syndrome, and cranial or peripheral neuritis have been reported from different regions with incidence of 2–40%. Convulsions occasionally arise in young children.¹⁴² Examination of the cerebrospinal fluid

is recommended in patients with behavioural or neurological manifestations and negative blood cultures to rule out tubercular meningitis.

Severe complications disseminated such as intravascular coagulation or haemorrhages can lead rapidly to death. Severe pneumonia is more frequent in children than adults. The other rare complications reported include hepatic, splenic, and bone-marrow granulomas, splenic and liver abscesses, pleural effusion, multiple organ dysfunction syndrome, haematophagocytic syndrome, pseudotumour cerebri, haemolytic uraemic syndrome, glomerulonephritis or pylonephritis, endocarditis, and pericarditis.143-147 Arrhythmia or cardiogenic shock can be a manifestation of toxic myocarditis with fatty infiltration of the heart.

With the introduction of early and appropriate antibiotic therapy, the average case fatality rates for typhoid are less than 1%; however, mortality as high as 30-50% has been reported from Papua New Guinea and Indonesia for severe typhoid fever.^{148,149} In 552 culture-positive patients with typhoid fever in hospital in Bangladesh, the overall case-fatality rate was 4.3%, with the highest rates for those younger than 1 year (11%) and for adults 31 years or older (10%).142 Further, 5-10% of cases can relapse after 2-3 weeks of resolution of the initial fever, but the clinical severity of this episode is much lower. Relapse can happen without a history of therapeutic intervention but more often it follows antibiotic treatment.150-152 The incidence of relapse after treatment with fluoroquinolones (1.5%) or broad-spectrum cephalosporins (5%) is lower than that after treatment with chloramphenicol, trimethoprim-sulfamethoxazole, and ampicillin.153 Most relapses are caused by the same S typhi strain with the same antibiotic susceptibility patterns as the initial episode. However, some individuals become reinfected with distinct and possibly newly acquired isolates.153

Clinical features of paratyphoid fever are similar to those of typhoid fever but are usually milder with a shorter incubation period. *S paratyphi A* or *paratyphi B* can manifest with jaundice, thrombosis,^{154,155} and systemic infections.¹⁵⁶ *S paratyphi B* might occasionally have an onset similar to non-specific salmonella gastroenteritis. Gastrointestinal symptoms are usually not present with *S paratyphi C* but there have been cases with systemic complications such as septicaemia and arthritis.¹⁵⁷ A relapse rate of 8% has been reported with *S paratyphi A*.¹⁵⁸

Treatment

Important considerations for treatment are the prevention of severe complications and death, and prompt resolution of clinical disease. It is also important to eradicate the organism promptly to prevent relapses and faecal carriage. More than 90% of patients are managed at home with oral antimicrobials, bedrest, and close medical follow up. Patients with persistent vomiting, severe diarrhoea, or abdominal distension need admission to hospital and parenteral antibiotic therapy.⁴⁶

Typhoid fever should be a serious consideration in an endemic area when a fever lasts longer than a week, even by the fifth day if there is severe toxaemia. The decision to send laboratory investigations and initiate empirical antimicrobial therapy depends largely on clinical judgment. Initial choice of antibiotic depends on the sensitivity patterns of *S typhi* and *paratyphi* isolates in the area. The isolates can be broadly classified as sensitive to first-line antimicrobials, multidrug resistant but nalidixic-acid sensitive, and nalidixic-acid resistant (often also multidrug resistant).

Fluoroquinolones (ciprofloxacin, ofloxacin, and pefloxacin) are the most effective drugs for treatment of

	First-line oral drug			Second-line oral drug		
	Antibiotic	Daily dose (mg/kg)	Days	Antibiotic	Daily dose (mg/kg)	Days
Uncomplicated typho	id at home					
Fully susceptible	Fluoroquinolone	15	5-7	Chloramphenicol	50-75	14-21
				Amoxycillin	75-100	14
				Trimethoprim	8 (trimethoprim)	14
				-sulpfamethoxzole	40 (sulfamethoxazole)	
Multidrug-resistant	Fluoroquinolone	15	5-7	Azithromycin	8-10	
				or cefixime	20	7
						7-14
Nalidixic-acid resistant	Azithromycin	8-10	7	Fluoroquinolone	20	10-14
	or cefixime	20	7-14			
Severe typhoid in hos	oital					
Fully susceptible	Fluoroquinolone	15	10-14	Chloramphenicol	100	14-21
				Ampicillin	100	10-14
Multidrug-resistant	Fluoroquinolone	15	10-14	Ceftriaxone or	60	10-14
				cefotaxime	80	
Nalidixic acid resistant	Ceftriaxone or	60	10-14	Fluoroquinolone	20	10-14
	cefotaxime	80				
Adapted from Parry and co	lleagues.96					

typhoid fever caused by isolates that are not quinolone resistant,¹⁵⁹⁻¹⁶³ with a clinical cure rate of about 98%, fever clearance time of about 4 days, and relapse and faecal carriage rates of less than 2%.⁹⁶ Chloramphenicol, the traditional first-line drug of choice, is less effective than fluoroquinolones in all these respects and in terms of persistence of the organism in bone marrow, even for treatment of patients with fully sensitive isolates.^{96,164}

Fluoroquinolones are equally effective for treatment of typhoid fever in children and adults.¹⁶⁵ However, these drugs are not registered for routine use in children because of evidence of articular damage in growing, weight-bearing joints in young beagles.^{166,167} Extensive experience of these drugs in children with typhoid, cystic fibrosis, and dysentery has shown no evidence of bone or joint toxicity, and impairment of linear growth.^{168–172} Where multidrug resistant strains are common and third-generation cephalosporins unavailable, fluoro-quinolones can be used for treatment of typhoid fever in children.

Short courses of ofloxacin given for 2, 3, or 5 days treated more than 90% of patients infected with multidrug resistant but nalidixic-acid susceptible isolates of *S typhi*, with an average fever clearance time of 4 days. Less than 3% of patients relapsed or had a positive posttreatment stool culture. Such short-term regimens are especially useful in control of epidemics. By contrast, the response of NAR isolates to such regimens is poor. Ofloxacin given for 7 days cured only 75% of patients infected with NAR isolates with a fever clearance time of 7 days, and 19% patients had a positive posttreatment stool culture.^{103,108}

Azithromycin, and cefixime, an oral third-generation cepahalosporin, have a clinical cure rate of over 90% with a fever clearance time of 5–7 days and relapse and faecal carriage rates of less than 4% in typhoid fever.^{108,161–163,173–176} These antimicrobials are therefore regarded as acceptable therapy for quinolone-resistant typhoid and paratyphoid fever. Their major limitations are high cost and low availability (table 6).

Combinations of antimicrobials are being assessed to provide more affordable options for treatment of quinolone-resistant typhoid fever. Ciprofloxacin proved to be more effective in combination with amoxycillin than ciprofloxacin alone against *S typhi* strains in vitro with a ciprofloxacin minimum inhibitory concentration of 1 mg/L or more.¹⁷⁷ The fractional inhibitory concentration for ciprofloxacin against these isolates was 0.004-0.256 mg/L when the antibiotic combination was used.¹⁷⁷ Similar findings have been reported for a combination of ciprofloxacin and gentamicin.¹⁷⁸

Supportive treatment includes maintenance of hydration, appropriate nutrition, and antipyretics. In a recent study, children with uncomplicated typhoid fever were randomly assigned to receive either ibuprofen or paracetamol every 6 h until 36 h after defervescence.¹⁷⁹ Median fever clearance time was shorter with ibuprofen

as was the area under the temperature–time curve above 37°C. The differences were mainly in children infected with NAR *S typhi* who responded slowly to antibiotic treatment.

Severe typhoid and paratyphoid

Patients with persistent vomiting, severe diarrhoea or abdominal distension requiring admission, and those with complications should be treated for severe typhoid fever. Fluoroquinolones remain the antibiotic of choice in areas where prevalence of quinolone-resistant isolates is low even though there have been no randomised antibiotic trials in patients with severe typhoid.165 Third-generation cephalosporins (ceftriaxone or cefotaxime) are recommended where quinolone resistant isolates are prevalent (table 6). Antimicrobials should be given parenterally for at least 10 days, or for at least for 5 days after defervescence.^{180,181} In Pakistan, clinical and bacteriological cure rates were similar with 7-day or 14-day treatment with intravenous ceftriaxone, but the confirmed bacteriological relapse rates were 14% with the 7-day regimen whereas none in the 14-day treatment group relapsed.187

In Indonesian adults and children with delirium, obtundation, stupor, coma, or shock, mortality was reduced from 50% to 10% for those given dexamethasone as an initial dose of 3 mg/kg by slow intravenous infusion over 30 min followed by 1 mg/kg at the same rate every 6 h for eight additional doses.^{148,183}

Patients with intestinal haemorrhage need intensive care, monitoring, and blood transfusion when blood loss is substantial. Patients with intestinal perforation should be resuscitated with fluids or blood and undergo early surgical intervention within 6 h to prevent death.⁴⁶ Relapses should be treated in the same way as initial infections.

Treatment of chronic carriers

Most carriers (defined as individuals who excrete *S typhi* in their stools or urine for more than a year) without gallstones can be cured by a long course of antimicrobials. Almost 80% of carriers were cured by 750 mg of ciprofloxacin twice daily for 28 days, and 11 out of 12 carriers treated with 400 mg norfloxacin twice daily had negative stool and bile cultures for *S typhi* after 28 days of treatment.^{184,185} In patients with gallstones, cholecystectomy along with antibiotic therapy might be required. Carriers should be excluded from any activities involving food preparation and serving, as should be people recovering from typhoid fever.

Prevention

The key preventive strategies are safe water, safe food, personal hygiene, and appropriate sanitation.⁴⁶ The importance of prevention has greatly increased with the emergence of antibiotic resistance. However,

provision of safe water and appropriate sanitation are expensive and are usually linked with economic development. Vaccination is an additional effective tool for prevention of typhoid fever. Vaccination is useful for prevention of typhoid in travellers from developed countries to typhoid endemic countries, in preventing and controlling epidemics, as well for children in endemic settings aged 2–19 years.⁴⁶

Most cases of typhoid fever in developed countries are a result of travel to endemic disease areas. Six countries—India, Pakistan, Mexico, Bangladesh, the Philippines, and Haiti—accounted for 76% of travel associated cases.¹⁸⁶ 37% of these cases were in people who stayed at their travel destination for 4 weeks or less and 16% were in people who stayed for 2 weeks or less. 80% cases were in people visiting relatives and friends. Travellers to these countries, especially those visiting for 2 weeks or more and those visiting friends and relatives, should be targeted for vaccination.

The old parenteral whole-cell typhoid-paratyphoid A and B vaccine was effective against both typhoid and paratyphoid fevers but has been largely discontinued because of strong side-effects.¹⁸⁷ Two vaccines for typhoid fever, one based on Vi polysaccharide and the other on whole-cell live attenuated bacteria, are currently licensed. A new Vi-conjugate vaccine is highly effective in children younger than 5 years but it has not been tested in infants. Currently, there is no licensed vaccine for paratyphoid fever.

Vi polysaccharide vaccine

This vaccine is licensed for use in individuals older than 2 years and is given in a single subcutaneous or intramuscular dose. The vaccine is moderately effective for about 3 years after vaccination (table 7).^{14,21,188,189} Revaccination is recommended every 3 years. However, 58% of participants in a field trial in South Africa still had protective levels of antibodies 10 years after vaccination.¹⁹² This vaccine has shown about 70% protective efficacy in a population vaccinated before or during an outbreak in China.¹⁸⁹ The Vi vaccine can be given simultaneously with other vaccines relevant for international travellers such as yellow fever and hepatitis A.^{190,191,193}

Ty21a vaccine

This live oral vaccine available in enteric-coated or liquid formulation is approved for use in people 6 years of age and older. The liquid formulation for younger children is currently marketed in only a few countries. Three doses are recommended each given 2 days apart. Antimicrobials should be avoided for 7 days before or after vaccination. The vaccine is moderately effective for up to about 3 years after vaccination (table 7).^{11–13,15,194,195} A booster dose is recommended every 3 years in endemic areas and travellers should be revaccinated annually. Herd immunity was shown during field trials in

	Age range (years)	Follow up (years)	Dose	Protective efficacy (95% CI)
Vi vaccine				
Nepal ²¹	5-44	1.5	Single dose	72% (41-87)
South Africa186	5-16	3	Single dose	55% (30-71)
China ¹⁴	3-19	1.5	Single dose	69% (28-87)
China ¹⁸⁷	12-21	Immunised during	Single dose	71% (34-87)
		epidemic or up to		
		1 year earlier		
Ty21a vaccine				
Egypt ¹⁹⁴	6-7	3	3 doses (enteric coated capsules)	96% (67-99)
Chile ¹¹	6-21	3	3 doses (enteric coated capsules)	67% (47-79)
Chile ¹³	5-19	3	3 doses (enteric coated capsules)	33% (0-57)
			3 doses/liquid	77% (60-87)
Chile ¹²	5-22	2	2 doses (enteric coated capsules)	59% (41-71)
Chile ¹⁹⁵	6-19	5	3 doses (liquid)	78% (65-87)
Chile ¹⁹⁵	6-19	7	3 doses (enteric coated capsules)	62% (48-73)
Indonesia ¹⁵	3-44	2.5	3 doses (enteric coated capsules)	42% (23-57)
			3 doses (liquid)	53% (33-66)

Table 7: Effectiveness of two typhoid vaccines

Chile.^{11,13} The vaccine can be given simultaneously with other vaccines and with antimalarial prophylaxis.¹⁹⁶

The effectiveness of both these licensed vaccines in developing countries is similar. Ty21a has the advantage that it is given orally and therefore might be easier for immunising groups of children, as in schools. The Ty21a vaccine, especially the enteric-coated capsule formulation, is not licensed for use in 2–5 year-old children. Vi vaccine has a relative advantage that it can be used for these preschool children, in settings where typhoid fever is common in this age-group. The vaccine however, is not licensed for use in children younger than 2 years.

Post-marketing surveillance for typhoid fever vaccines from the Vaccine Adverse Effects Reporting System from 1990 to 2002 has shown rare reports of death, admission, permanent disability, or life-threatening illness.¹⁹⁷ Unexpected frequently reported symptoms included dizziness and pruritis for Vi vaccine and fatigue and myalgia for Ty21a. Gastroenteritis for Ty21a and abdominal pain after Vi vaccine are previously recognised events.

Vi-conjugate vaccine

Vi-conjugate vaccine given to 2–5 year-old Vietnamese children had 91·1% protection against typhoid 27 months after vaccination, with geometric mean titres of 7·61 ELISA units for those vaccinated at age 2–3 years.²⁰ The efficacy of this conjugate persisted after 46 months of vaccination: over the entire period the protection was 89% (95% CI 76%–97%).¹⁹⁸ Based on the antibody titres after 46 months of vaccination, the researchers suggest a protective level of antibody to immunoglobulin G to be reduced from 7 to 3·52 ELISA units. This vaccine could be used for children younger than 2 years and be incorporated into the Expanded Programme on Immunization immunisation schedules.

Vaccines under development

Vaccines are under development based on outer membrane proteins known as porins and new live oral vaccines (eg CVD 908-htrA and Ty2 candidate vaccines).¹⁹⁹⁻²⁰¹ A new vaccine against *S paratyphi A* composed of surface-O-specific polysaccharide conjugated with tetanus toxoid has proved safe and immunogenic.²⁰² Live typhoid vaccines are being developed as a vector for immunisation against *H pylori*.²⁰³

Conclusion

Typhoid fever is an important public-health problem in south-central and southeast Asia, the middle east, Africa, and South America, mainly affecting children and young adults. Treatment of typhoid fever is becoming more difficult with multidrug resistant organisms also acquiring some resistance to fluoroquinolones. Fully ciprofloxacin-resistant *S typhi* has not been reported, although full resistance in non-typhoid salmonella has emerged.^{204,205} Effective immunisation and non-vaccine based prevention strategies are available and are becoming more important in the face of increasing antibiotic resistance.

S typhi is restricted to human beings. The organism does not persist long in any environmental reservoir after its elimination from local regions. The analysis of individual pseudogene mutations from *S typhi* isolates shows that different isolates harbour the same mutations. This strongly suggests that the organism has emerged only once and cannot readily evolve from other *S typhi* serotypes.⁵³ Improved sanitation, better diagnostics for early detection and treatment of patients and carriers, and mass vaccination are effective interventions in combating typhoid fever. All these factors indicate that global eradication of typhoid is theoretically possible and merits consideration as a global health issue.

Conflict of interest statement

We declare that we have no conflict of interest.

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References

- Kidgell C, Reichard U, Wain J, et al. Salmonella typhi, the causative agent of typhoid fever, is approximately 50,000 years old. Infect Genet Evol 2002; 2: 39–45.
- Merrell DS, Falkow S. Frontal and stealth attack strategies in microbial pathogenesis. *Nature* 2004; 430: 250–56.
- 3 Osler W. The principles and practice of medicine: designed for the use of practitioners and students of medicine. 8th edn. New York: D Appleton, 1912: 1–46.
- 4 Crump JA, Luby SP, Mintz ED. The global burden of typhoid fever. Bull World Health Organ 2004; 82: 346–53.
- 5 Woodward TE, Smadel JE, Ley HL Jr, Green R, Manikar DS. Preliminary report of the beneficial effect of chloromycetin in the treatment of typhoid fever. Ann Intern Med 1948; 29: 131–34.

- 6 Mirza SH, Beeching NJ, Hart CA. Multi-drug resistant typhoid: a global problem. J Med Microbiol 1996; 44: 317–19.
- 7 Rowe B, Ward LR, Threlfall EJ. Multidrug-resistant Salmonella typhi: a worldwide epidemic. Clin Infect Dis 1997; 24 (suppl 1): \$106–09.
- Mehta G, Randhawa VS, Mohapatra NP. Intermediate susceptibility to ciprofloxacin in Salmonella typhi strains in India. Eur J Clin Microbiol Infect Dis 2001; 20: 760–61.
- 9 Harish BN, Madhulika U, Parija SC. Isolated high-level ciprofloxacin resistance in *Salmonella enterica* subsp. enterica serotype Paratyphi A. J Med Microbiol 2004; 53: 819.
- Saha SK, Talukder SY, Islam M, Saha S. A highly ceftriaxoneresistant Salmonella typhi in Bangladesh. Pediatr Infect Dis J 1999; 18: 387.
- 11 Levine MM, Ferreccio C, Black RE, Germanier R. Large-scale field trial of Ty21a live oral typhoid vaccine in enteric-coated capsule formulation. *Lancet* 1987; 1: 1049–52.
- 12 Black RE, Levine MM, Ferreccio C, et al. Efficacy of one or two doses of Ty21a Salmonella typhi vaccine in enteric-coated capsules in a controlled field trial. Chilean Typhoid Committee. Vaccine 1990; 8: 81–84.
- 13 Levine MM, Ferreccio C, Cryz S, Ortiz E. Comparison of entericcoated capsules and liquid formulation of Ty21a typhoid vaccine in randomised controlled field trial. *Lancet* 1990; 336: 891–94.
- 14 Yang HH, Wu CG, Xie GZ, et al. Efficacy trial of Vi polysaccharide vaccine against typhoid fever in south-western China. Bull World Health Organ 2001; 79: 625–31.
- 15 Simanjuntak CH, Paleologo FP, Punjabi NH, et al. Oral immunisation against typhoid fever in Indonesia with Ty21a vaccine. *Lancet* 1991; 338: 1055–59.
- 16 Bahl R, Sinha A, Poulos C, et al. Costs of illness due to typhoid fever in an Indian urban slum community: implications for vaccination policy. J Health Popul Nutr 2004; 22: 304–10.
- 7 Sinha A, Sazawal S, Kumar R, et al. Typhoid fever in children aged less than 5 years. *Lancet* 1999; 354: 734–37.
- 18 Klugman KP, Gilbertson IT, Koornhof HJ, et al. Protective activity of Vi capsular polysaccharide vaccine against typhoid fever. *Lancet* 1987; 2: 1165–69.
- 19 Lin FY, Vo AH, Phan VB, et al. The epidemiology of typhoid fever in the Dong Thap Province, Mekong Delta region of Vietnam. *Am J Trop Med Hyg* 2000; **62**: 644–48.
- 20 Lin FY, Ho VA, Khiem HB, et al. The efficacy of a Salmonella typhi Vi conjugate vaccine in two-to-five-year-old children. N Engl J Med 2001; 344: 1263–69.
- 21 Acharya IL, Lowe CU, Thapa R, et al. Prevention of typhoid fever in Nepal with the Vi capsular polysaccharide of *Salmonella typhi*. A preliminary report. N Engl J Med 1987; **317**: 1101–04.
- 22 Edelman R, Levine MM. Summary of an international workshop on typhoid fever. *Rev Infect Dis* 1986; **8**: 329–49.
- 23 Thisyakorn U, Mansuwan P, Taylor DN. Typhoid and paratyphoid fever in 192 hospitalized children in Thailand. Am J Dis Child 1987; 141: 862–65.
- 24 Saha SK, Baqui AH, Hanif M, et al. Typhoid fever in Bangladesh: implications for vaccination policy. *Pediatr Infect Dis J* 2001; **20**: 521–24.
- 25 Ackers ML, Puhr ND, Tauxe RV, Mintz ED. Laboratory-based surveillance of *Salmonella* serotype *Typhi* infections in the United States: antimicrobial resistance on the rise. *JAMA* 2000; 283: 2668–73.
- 26 Reller ME, Olsen SJ, Kressel AB, et al. Sexual transmission of typhoid fever: a multistate outbreak among men who have sex with men. *Clin Infect Dis* 2003; 37: 141–44.
- 27 Olsen SJ, Bleasdale SC, Magnano AR, et al. Outbreaks of typhoid fever in the United States, 1960–99. *Epidemiol Infect* 2003; 130: 13–21.
- 28 Vollaard AM, Ali S, van Asten HA, et al. Risk factors for typhoid and paratyphoid fever in Jakarta, Indonesia. JAMA 2004; 291: 2607–15.
- 29 Shlim DR, Schwartz E, Eaton M. Clinical importance of *Salmonella paratyphi A* infection to enteric fever in Nepal. *J Travel Med* 1995; 2: 165–68.
- 30 Sood S, Kapil A, Dash N, Das BK, Goel V, Seth P. Paratyphoid Fever in India: an emerging problem. *Emerg Infect Dis* 1999; 5: 483–84.

- 31 Tankhiwale SS, Agrawal G, Jalgaonkar SV. An unusually high occurrence of Salmonella enterica serotype paratyphi A in patients with enteric fever. Indian J Med Res 2003; 117: 10–12.
- 32 Levine MM, Black RE, Lanata C. Precise estimation of the numbers of chronic carriers of Salmonella typhi in Santiago, Chile, an endemic area. J Infect Dis 1982; 146: 724–26.
- 33 Cho JC, Kim SJ. Viable, but non-culturable, state of a green fluorescence protein-tagged environmental isolate of *Salmonella typhi* in groundwater and pond water. *FEMS Microbiol Lett* 1999; 170: 257–64.
- 34 Wait DA, Sobsey MD. Comparative survival of enteric viruses and bacteria in Atlantic Ocean seawater. Water Sci Technol 2001; 43: 139–42.
- 35 Nishio T, Nakamori J, Miyazaki K. Survival of Salmonella typhi in oysters. Zentralbl Bakteriol Mikrobiol Hyg [B] 1981; 172: 415–26.
- 36 Elsarnagawy D. Viability of some Salmonella strains in Algerian eggs. Arch Inst Pasteur Alger 1978–79; 53: 282–90.
- 37 Hornick RB, Greisman SE, Woodward TE, DuPont HL, Dawkins AT, Snyder MJ. Typhoid fever: pathogenesis and immunologic control. N Engl J Med 1970; 283: 686–91 and 739–46.
- 38 Swaddiwudhipong W, Kanlayanaphotporn J. A common-source water-borne outbreak of multidrug-resistant typhoid fever in a rural Thai community. J Med Assoc Thai 2001; 84: 1513–17.
- 39 Mermin JH, Villar R, Carpenter J, et al. A massive epidemic of multidrug-resistant typhoid fever in Tajikistan associated with consumption of municipal water. J Infect Dis 1999; 179: 1416–22.
- 40 Black RE, Cisneros L, Levine MM, Banfi A, Lobos H, Rodriguez H. Case-control study to identify risk factors for paediatric endemic typhoid fever in Santiago, Chile. *Bull World Health Organ* 1985; 63: 899–904.
- 41 Luby SP, Faizan MK, Fisher-Hoch SP, et al. Risk factors for typhoid fever in an endemic setting, Karachi, Pakistan. *Epidemiol Infect* 1998; 120: 129–38.
- 42 Bhan MK, Bahl R, Sazawal S, et al. Association between *Helicobacter pylori* infection and increased risk of typhoid fever. J Infect Dis 2002; 186: 1857–60.
- 43 Luxemburger C, Chau MC, Mai NL, et al. Risk factors for typhoid fever in the Mekong delta, southern Viet Nam: a case-control study. *Trans R Soc Trop Med Hyg* 2001; 95: 19–23.
- 44 Gasem MH, Dolmans WM, Keuter MM, Djokomoeljanto RR. Poor food hygiene and housing as risk factors for typhoid fever in Semarang, Indonesia. *Trop Med Int Health* 2001; 6: 484–90.
- 45 Morris JG Jr, Ferreccio C, Garcia J, et al. Typhoid fever in Santiago, Chile: a study of household contacts of pediatric patients. *Am J Trop Med Hyg* 1984; 33: 1198–202.
- 46 World Health Organization. Background document: The diagnosis, treatment and prevention of typhoid fever. WHO/V&B/03.07. Geneva: World Health Organization, 2003.
- 47 Ryan C, Salmonella typhi infections in the United States, 1975–1984: increasing role of foreign travel. Rev Infect Dis 1989; 11: 1–8.
- 48 Dunstan SJ, Stephens HA, Blackwell JM, et al. Genes of the class II and class III major histocompatibility complex are associated with typhoid fever in Vietnam. J Infect Dis 2001; 183: 261–68.
- 49 Dharmana E, Joosten I, Tijssen HJ, et al. HLA-DRB1*12 is associated with protection against complicated typhoid fever, independent of tumour necrosis factor alpha. *Eur J Immunogenet* 2002; 29: 297–300.
- 50 Jegathesan M. Phage types of *Salmonella typhi* isolated in Malaysia over the 10-year period 1970–1979. *J Hyg (Lond)* 1983; **90**: 91–97.
- 51 Thong KL, Bhutta ZA, Pang T. Multidrug-resistant strains of Salmonella enterica serotype typhi are genetically homogenous and coexist with antibiotic-sensitive strains as distinct, independent clones. Int J Infect Dis 2000; 4: 194–97.
- 52 Connerton P, Wain J, Hien TT, et al. Epidemic typhoid in vietnam: molecular typing of multiple-antibiotic-resistant Salmonella enterica serotype typhi from four outbreaks. J Clin Microbiol 2000; 38: 895–97.
- 53 Mirza S, Kariuki S, Mamun KZ, Beeching NJ, Hart CA. Analysis of plasmid and chromosomal DNA of multidrug-resistant *Salmonella enterica* serovar typhi from Asia. *J Clin Microbiol* 2000; 38: 1449–52.
- 54 Bhutta ZA, Naqvi SH, Razzaq RA, Farooqui BJ. Multidrug-resistant typhoid in children: presentation and clinical features. *Rev Infect Dis* 1991; 13: 832–36.

- 55 Thong KL, Passey M, Clegg A, Combs BG, Yassin RM, Pang T. Molecular analysis of isolates of *Salmonella typhi* obtained from patients with fatal and nonfatal typhoid fever. *J Clin Microbiol* 1996; 34: 1029–33.
- 56 Parkhill J, Dougan G, James KD, et al. Complete genome sequence of a multiple drug resistant Salmonella enterica serovar Typhi CT18. Nature 2001; 413: 848–52.
 - 7 Wain J, House D, Parkhill J, Parry C, Dougan G. Unlocking the genome of the human typhoid bacillus. *Lancet Infect Dis* 2002; 2: 163–70.
- 58 Sherburne CK, Lawley TD, Gilmour MW, et al. The complete DNA sequence and analysis of R27, a large IncHI plasmid from Salmonella typhi that is temperature sensitive for transfer. Nucleic Acids Res 2000; 28: 2177–86.
- 59 McClelland M, Sanderson KE, Clifton SW, et al. Comparison of genome degradation in Paratyphi A and Typhi, human-restricted serovars of *Salmonella enterica* that cause typhoid. *Nat Genet* 2004; 36: 1268–74.
- 60 Weinstein DL, O'Neill BL, Hone DM, Metcalf ES. Differential early interactions between *Salmonella enterica* serovar Typhi and two other pathogenic Salmonella serovars with intestinal epithelial cells. *Infect Immun* 1998; 66: 2310–18.
- 61 Pier GB, Grout M, Zaidi T, et al. *Salmonella typhi* uses CFTR to enter intestinal epithelial cells. *Nature* 1998; **393**: 79–82.
- 62 Kops SK, Lowe DK, Bernent WM, West AB. Migration of Salmonella typhi through intestinal epithelial monolayers: an in vitro study. Microbiol Immunol 1996; 40: 799–811.
- 53 Lyczak JB, Zaidi TS, Grout M, Bittner M, Contreras I, Pier GB. Epithelial cell contact-induced alterations in *Salmonella enterica* serovar Typhi lipopolysaccharide are critical for bacterial internalization. *Cell Microbiol* 2001; 3: 763–72.
- 14 Lyczak JB, Pier GB. Salmonella enterica serovar typhi modulates cell surface expression of its receptor, the cystic fibrosis transmembrane conductance regulator, on the intestinal epithelium. Infect Immun 2002; 70: 6416–23.
- 65 Hardt WD, Chen LM, Schuebel KE, Bustelo XR, Galan JE. S. typhimurium encodes an activator of Rho GTPases that induces membrane ruffling and nuclear responses in host cells. Cell 1998; 93: 815–26.
- 66 Galan JE, Zhou D. Striking a balance: modulation of the actin cytoskeleton by Salmonella. *Proc Natl Acad Sci USA* 2000; 97: 8754–61.
- 67 Galan JE. Salmonella interactions with host cells: type III secretion at work. *Annu Rev Cell Dev Biol* 2001; **17**: 53–86.
- 68 Hernandez LD, Hueffer K, Wenk MR, Galan JE. Salmonella modulates vesicular traffic by altering phosphoinositide metabolism. *Science* 2004; 304: 1805–07.
- 69 Boddicker JD, Jones BD. Lon protease activity causes downregulation of Salmonella pathogenicity island 1 invasion gene expression after infection of epithelial cells. *Infect Immun* 2004; 72: 2002–13.
- 70 Waterman SR, Holden DW. Functions and effectors of the Salmonella pathogenicity island 2 type III secretion system. *Cell Microbiol* 2003; 5: 501–11.
- 71 Beuzon CR, Meresse S, Unsworth KE, et al. Salmonella maintains the integrity of its intracellular vacuole through the action of SifA. *EMBO J* 2000; **19**: 3235–49.
- 72 Chen LM, Bagrodia S, Cerione RA, Galan JE. Requirement of p21-activated kinase (PAK) for Salmonella typhimurium-induced nuclear responses. J Exp Med 1999; 189: 1479–88.
- 73 Vazquez-Torres A, Vallance BA, Bergman MA, et al. Toll-like receptor 4 dependence of innate and adaptive immunity to Salmonella: importance of the Kupffer cell network. *J Immunol* 2004; 172: 6202–08.
- 74 House D, Bishop A, Parry C, Dougan G, Wain J. Typhoid fever: pathogenesis and disease. *Curr Opin Infect Dis* 2001; 14: 573–78.
- 75 Vidal S, Tremblay ML, Govoni G, et al. The Ity/Lsh/Bcg locus: natural resistance to infection with intracellular parasites is abrogated by disruption of the Nramp1 gene. J Exp Med 1995; 182: 655–66.
- 76 Monack DM, Bouley DM, Falkow S. Salmonella typhimurium persists within macrophages in the mesenteric lymph nodes of chronically infected Nramp1+/+ mice and can be reactivated by IFNgamma neutralization. J Exp Med 2004; 199: 231–41.

- 77 Wang F, Gu XJ, Zhang MF, Tai TY. Treatment of typhoid fever with ofloxacin. J Antimicrob Chemother 1989; 23: 785–88.
- 78 Jesudasan M, John TJ. Multiresistant Salmonella typhi in India. Lancet 1990; 336: 252.
- 79 Rowe B, Ward LR, Threlfall EJ. Spread of a multiresistant Salmonella typhi. Lancet 1990; **336**: 1065.
- 80 Wallace MR, Yousif AA, Mahroos GA, Mapes T, Threlfall EJ, Rowe B. Ciprofloxacin versus ceftriaxone in the treatment of multiresistant typhoid fever. *Eur J Clin Microbiol Infect Dis* 1993; 12: 907–10.
- 81 Karmaker S, Biswas D, Shaikh NM, Chatterjee SK, Kataria VK, Kumar R. Role of a large plasmid of *Salmonella typhi* encoding multiple drug resistance. *J Med Microbiol* 1991; 34: 149–51.
- 82 Yoo S, Pai H, Byeon JH, Kang YH, Kim S, Lee BK. Epidemiology of *Salmonella enterica* serotype typhi infections in Korea for recent 9 years: trends of antimicrobial resistance. *J Korean Med Sci* 2004; 19: 15–20.
- 83 Akinyemi KO, Coker AO, Olukoya DK, Oyefolu AO, Amorighoye EP, Omonigbehin EO. Prevalence of multi-drug resistant Salmonella typhi among clinically diagnosed typhoid fever patients in Lagos, Nigeria. Z Naturforsch [C] 2000; 55: 489–93.
- 84 Hoa NT, Diep TS, Wain J, et al. Community-acquired septicaemia in southern Viet Nam: the importance of multidrugresistant Salmonella typhi. Trans R Soc Trop Med Hyg 1998; 92: 503–08.
- 85 Mills-Robertson F, Addy ME, Mensah P, Crupper SS. Molecular characterization of antibiotic resistance in clinical Salmonella typhi isolated in Ghana. FEMS Microbiol Lett 2002; 215: 249–53.
- 86 Hermans PW, Saha SK, van Leeuwen WJ, Verbrugh HA, van Belkum A, Goessens WH. Molecular typing of Salmonella typhi strains from Dhaka (Bangladesh) and development of DNA probes identifying plasmid-encoded multidrug-resistant isolates. J Clin Microbiol 1996; 34: 1373–79.
- 87 Kariuki S, Gilks C, Revathi G, Hart CA. Genotypic analysis of multidrug-resistant Salmonella enterica Serovar typhi, Kenya. Emerg Infect Dis 2000; 6: 649–51.
- 88 Bahrmand AR, Velayati AA. Antimicrobial resistance pattern and plasmid profile of *Salmonella typhi* isolated from an outbreak in Tehran province. *Scand J Infect Dis* 1997; 29: 265–69.
- 89 El-Sherbini A. An outbreak of typhoid fever resistant to chloramphenicol and other drugs in Gharbeya Governorate in Egypt. J Trop Pediatr 1991; 38: 331–34.
- 90 Watson JP. Multi-resistant typhoid fever in Nepal. Trop Doct 1992; 22: 172.
- 91 Coovadia YM, Gathiram V, Bhamjee A, et al. An outbreak of multiresistant *Salmonella typhi* in South Africa. *Q J Med* 1992; 82: 91–100.
- 92 Panigrahi D, al-Aneziz AH, West PW. Plasmid-mediated multidrug resistance in *Salmonella typhi* in Kuwait. *Trop Med Int Health* 1996; 1: 439–42.
- 93 Panhotra BR, Saxena AK, Al-Ghamdi AM. Typhoid fever due to multiresistant Salmonella enterica serovar typhi having reduced susceptibility to ciprofloxacin and nalidixic acid resistance. Saudi Med J 2004; 25: 1509–11.
- 94 Battikhi MN. Occurrence of Salmonella typhi and Salmonella paratyphi in Jordan. New Microbiol 2003; 26: 363–73.
- 95 Taylor DE, Chumpitaz JC, Goldstein F. Variability of IncH11 plasmids from Salmonella typhi with special reference to Peruvian plasmids encoding resistance to trimethoprim and other antibiotics. Antimicrob Agents Chemother 1985; 28: 452–55.
- 96 Parry CM, Hien TT, Dougan G, White NJ, Farrar JJ. Typhoid fever. N Engl J Med 2002; 347: 1770–82.
- 97 Sherburne CK, Lawley TD, Gilmour MW, et al. The complete DNA sequence and analysis of R27, a large IncHI plasmid from *Salmonella typhi* that is temperature sensitive for transfer. *Nucleic Acids Res* 2000 May 15; 28: 2177–86.
- 98 Le TA, Lejay-Collin M, Grimont PA, et al. Endemic, epidemic clone of *Salmonella enterica* serovar typhi harboring a single multidrug-resistant plasmid in Vietnam between 1995 and 2002. *J Clin Microbiol* 2004; **42**: 3094–99.
- 99 Rahman M, Ahmad A, Shoma S. Decline in epidemic of multidrug resistant *Salmonella typhi* is not associated with increased incidence of antibiotic-susceptible strain in Bangladesh. *Epidemiol Infect* 2002; **129**: 29–34.

- 100 Sood S, Kapil A, Das B, Jain Y, Kabra SK. Re-emergence of chloramphenicol-sensitive Salmonella typhi. Lancet 1999; 353: 1241–42.
- 101 Mandal S, Mandal MD, Pal NK. Reduced minimum inhibitory concentration of chloramphenicol for Salmonella enterica serovar typhi. Indian J Med Sci 2004; 58: 16–23.
- 102 Threlfall EJ, Ward LR, Skinner JA, Smith HR, Lacey S. Ciprofloxacin-resistant Salmonella typhi and treatment failure. Lancet 1999; 353: 1590–91.
- 103 Wain J, Hoa NT, Chinh NT, et al. Quinolone-resistant Salmonella typhi in Viet Nam: molecular basis of resistance and clinical response to treatment. Clin Infect Dis 1997; 25: 1404–10.
- 104 Murdoch DA, Banatvala N, Bone A, Shoismatulloev BI, Ward LR, Threlfall EJ. Epidemic ciprofloxacin-resistant Salmonella typhi in Tajikistan. Lancet 1998; 351: 339.
- 105 Asna SM, Haq JA, Rahman MM. Nalidixic acid-resistant Salmonella enterica serovar Typhi with decreased susceptibility to ciprofloxacin caused treatment failure: a report from Bangladesh. Jpn J Infect Dis 2003; 56: 32–33.
- 106 Kariuki S, Revathi G, Muyodi J, et al. Characterization of multidrugresistant typhoid outbreaks in Kenya. J Clin Microbiol 2004; 42: 1477–82.
- 107 Renuka K, Kapil A, Kabra SK, et al. Reduced susceptibility to ciprofloxacin and gyra gene mutation in North Indian strains of *Salmonella enterica* serotype Typhi and serotype Paratyphi A. *Microb Drug Resist* 2004; **10**: 146–53.
- 108 Parry CM. The treatment of multidrug-resistant and nalidixic acidresistant typhoid fever in Viet Nam. *Trans R Soc Trop Med Hyg* 2004; 98: 413–22.
- 109 Brown JC, Shanahan PM, Jesudason MV, Thomson CJ, Amyes SG. Mutations responsible for reduced susceptibility to 4-quinolones in clinical isolates of multi-resistant *Salmonella typhi* in India. *J Antimicrob Chemother* 1996; **37**: 891–900.
- 110 Walker RA, Saunders N, Lawson AJ, et al. LightCycler gyrA mutation assay (GAMA) identifies heterogeneity in GyrA in *Salmonella enterica* serotypes Typhi and Paratyphi A with decreased susceptibility to ciprofloxacin. *Int J Antimicrob Agents* 2003; **22**: 622–25.
- 111 Threlfall EJ, Skinner JA, Ward LR. Detection of decreased in vitro susceptibility to ciprofloxacin in *Salmonella enterica* serotypes Typhi and Paratyphi A. J Antimicrob Chemother 2001; **48**: 740–41.
- 112 Slinger R, Desjardins M, McCarthy AE, et al. Suboptimal clinical response to ciprofloxacin in patients with enteric fever due to *Salmonella* spp. with reduced fluoroquinolone susceptibility: a case series. *BMC Infect Dis* 2004; **4**: 36.
- 113 National Committee on Clinical Laboratory Standards. (NCCLS). Performance standards for antimicrobial susceptibility testing; Fourteenth informational supplement M100-S14. PA: NCCLS, 2004.
- 114 Threlfall EJ, Fisher IS, Berghold C, et al. Trends in antimicrobial drug resistance in *Salmonella enterica* serotypes Typhi and Paratyphi A isolated in Europe, 1999–2001. *Int J Antimicrob Agents* 2003; 22: 487–91.
- 115 Chandel DS, Chaudhry R, Dhawan B, Pandey A, Dey AB. Drugresistant Salmonella enterica serotype paratyphi A in India. Emerg Infect Dis 2000; 6: 420–21.
- 116 Gilman RH, Terminel M, Levine MM, Hernandez-Mendoza P, Hornick RB. Relative efficacy of blood, urine, rectal swab, bone marrow, and rose-spot cultures for recovery of *Salmonella typhi* in typhoid fever. *Lancet* 1975; 1: 1211–13.
- 117 Vallenas C, Hernandez H, Kay B, Black R, Gotuzzo E. Efficacy of bone marrow, blood, stool and duodenal contents cultures for bacteriologic confirmation of typhoid fever in children. *Pediatr Infect Dis* 1985; 4: 496–98.
- 118 Hoffman SL, Edman DC, Punjabi NH, et al. Bone marrow aspirate culture superior to streptokinase clot culture and 8 ml 1:10 blood-tobroth ratio blood culture for diagnosis of typhoid fever. *Am J Trop Med Hyg* 1986; **35**: 836–39.
- 119 Rubin FA, McWhirter PD, Burr D, et al. Rapid diagnosis of typhoid fever through identification of *Salmonella typhi* within 18 hours of specimen acquisition by culture of the mononuclear cell-platelet fraction of blood. *J Clin Microbiol* 1990; 28: 825–27.
- 120 Farooqui BJ, Khurshid M, Ashfaq MK, Khan MA. Comparative yield of *Salmonella typhi* from blood and bone marrow cultures in patients with fever of unknown origin. *J Clin Pathol* 1991; 44: 258–59.

- 121 Gasem MH,. Dolmans WM, Isbandrio BB, Wahyono H, Keuter M, Djokomoeljanto R. Culture of Salmonella typhi and Salmonella paratyphi from blood and bone marrow in suspected typhoid fever. Trop Geogr Med 1995; 47: 164–67.
- 122 Wain J, Diep TS, Ho VA, et al. Quantitation of bacteria in blood of typhoid fever patients and relationship between counts and clinical features, transmissibility, and antibiotic resistance. *J Clin Microbiol* 1998; **36**: 1683–87.
- 123 Wain J, Bay PV, Vinh H, et al. Quantitation of bacteria in bone marrow from patients with typhoid fever; relationship between counts and clinical features. *Vaccine* 2001; **39**: 1571–76.
- 124 Lanata CF, Levine MM, Ristori C, et al. Vi serology in detection of chronic Salmonella typhi carriers in an endemic area. Lancet 1983; 322: 441–43.
- 125 Olopoenia LA, King AL. Widal agglutination test: 100 years later: still plagued by controversy *Postgrad Med J* 2000; 76: 80–84.
- 126 Pang T, Puthucheary SD. Significance and value of the Widal test in the diagnosis of typhoid fever in an endemic area. J Clin Pathol 1983; 36: 471–75.
- 127 Parry CM, Hoa NT, Diep TS, et al. Value of a single-tube Widal test in diagnosis of typhoid fever in Vietnam. J Clin Microbiol 1999; 37: 2882–86.
- 128 Olsen SJ, Pruckler J, Bibb W, et al. Evaluation of rapid diagnostic tests for typhoid fever. *J Clin Microbiol* 2004; **42**: 1885–89.
- 129 House D, Wain J, Ho VA, et al. Serology of typhoid fever in an area of endemicity and its relevance to diagnosis. *J Clin Microbiol* 2001; **39**: 1002–07.
- 130 Hatta M, Goris MG, Heerkens E, Gooskens J, Smits HL. Simple dipstick assay for the detection of *Salmonella typhis* specific IgM antibodies and the evolution of the immune response in patients with typhoid fever. *Am J Trop Med Hyg* 2002; 66: 416–21.
- 131 Gasem MH, Smits HL, Nugroho N, Goris MA, Dolmans WMV. Evaluation of a simple and rapid dipstick assay for the diagnosis of typhoid fever in Indonesia. *J Med Microbiol* 2002; **51**: 173–77.
- 132 Bhutta ZA, Mansurali N. Rapid serologic diagnosis of pediatric typhoid fever in an endemic area: a prospective comparative evaluation of two dot-enzyme immunoassays and the Widal test. *Am J Trop Med Hyg* 1999; 61: 654–57.
- 133 Jesudason M, Esther E, Mathai E. Typhidot test to detect IgG and IgM antibodies in typhoid fever. *Indian J Med Res* 2002; 116: 70–72.
- 134 Fadeel MA, Crump JA, Mahoney FJ, et al. Rapid diagnosis of typhoid fever by enzyme-linked immunosorbent assay detection of *Salmonella* serotype Typhi antigens in urine. *Am J Trop Med Hyg* 2004; **70**: 323–28.
- 135 Hirose K, Itoh K, Nakajima H, et al. Selective amplification of tyv (rfbE), prt (rfbS), viaB, and fliC genes by multiplex PCR for identification of *Salmonella enterica* serovars Typhi and Paratyphi A. J Clin Microbiol 2002; 40: 633–36.
- 136 Vinh H, Wain J, Vo TN, et al. Two or three days of ofloxacin treatment for uncomplicated multidrug-resistant typhoid fever in children. *Antimicrob Agents Chemother* 1996; 40: 958–61.
- 137 Shetty AK, Mital SR, Bahrainwala AH, Khubchandani RP, Kumta NB. Typhoid hepatitis in children. J Trop Pediatr 1999; 45: 287–90.
- 138 Chiu C-H, Lin T-Y. Typhoid fever in children. *Lancet* 1999; **354**: 2001–02.
- 139 Lee JH, Kim JJ, Jung JH, et al. Colonoscopic manifestations of typhoid fever with lower gastrointestinal bleeding. *Dig Liver Dis* 2004; 36: 141–46.
- 140 Van Basten JP, Stockenbrugger R. Typhoid perforation. A review of the literature since 1960. *Trop Geogr Med* 1994; 46: 336–39.
- 141 Hosoglu S, Aldemir M, Akalin S, Geyik MF, Tacyildiz IH, Loeb M. Risk factors for enteric perforation in patients with typhoid Fever. Am J Epidemiol 2004; 160: 46–50.
- 142 Butler T, Islam A, Kabir I, Jones PK. Patterns of morbidity and mortality in typhoid fever dependent on age and gender: review of 552 hospitalized patients with diarrhea. *Rev Infect Dis* 1991; 13: 85–90.

- 143 Chaudhry R, Mahajan RK, Diwan A, et al. Unusual presentation of enteric fever: three cases of splenic and liver abscesses due to *Salmonella typhi* and *Salmonella paratyphi A*. *Trop Gastroenterol* 2003; 24: 198–99.
- 144 Mert A, Tabak F, Ozaras R, Ozturk R, Aki H, Aktuglu Y. Typhoid fever as a rare cause of hepatic, splenic, and bone marrow granulomas. *Intern Med* 2004; 43: 436–39.
- 145 Snyder GE, Shaps HJ, Nelson M. Multiple organ dysfunction syndrome associated with *Salmonella typhi* infection. *Am J Emerg Med* 2004; 22: 138–39.
- 146 Albaqali A, Ghuloom A, Al Arrayed A, et al. Hemolytic uremic syndrome in association with typhoid fever. *Am J Kidney Dis* 2003; 41: 709–13.
- 147 Balasubramanian S, Shivbalan S, Miranda PK. Pseudotumour cerebri as an unusual manifestation of typhoid. Ann Trop Paediatr 2003; 23: 223–24.
- 148 Punjabi NH, Hoffman SL, Edman DC, et al. Treatment of severe typhoid fever in children with high dose dexamethasone. *Pediatr Infect Dis J* 1988; 7: 598–600.
- 149 Rogerson SJ, Spooner VJ, Smith TA, Richens J. Hydrocortisone in chloramphenicol-treated severe typhoid fever in Papua New Guinea. *Trans R Soc Trop Med Hyg* 1991; 85: 113–16.
- 150 Gotuzzo, E, Morris JG Jr, Benavente L, et al. Association between specific plasmids and relapse in typhoid fever. J Clin Microbiol 1987; 25: 1779–81.
- 151 Smith MD, Duong NM, Hoa NT, et al. Comparison of ofloxacin and ceftriaxone for short-course treatment of enteric fever. Antimicrob Agents Chemother 1994; 38: 1716–20.
- 52 Yew FS, Chew SK, Goh KT, Monteiro EH, Lim YS. Typhoid fever in Singapore: a review of 370 cases. J Trop Med Hyg 1991; 94: 352–57.
- 153 Wain J, Hien TT, Connerton P, et al. Molecular typing of multipleantibiotic-resistant *Salmonella enterica* serovar Typhi from Vietnam: application to acute and relapse cases of typhoid fever. *J Clin Microbiol* 1999; **37**: 2466–72.
- 154 Rajagopal A, Ramasamy R, Mahendran G, Thomas M. Hepatic abscess complicating paratyphoid infection. *Trop Gastroenterol* 2002; 23: 181–82.
- 155 Mohanty S, Bakshi S, Gupta AK, Kapil A, Arya LS, Das BK. Venous thrombosis associated with Salmonella: report of a case and review of literature. *Indian J Med Sci* 2003; 57: 199–203.
- 156 Lee WS, Puthucheary SD, Parasakthi N. Extra-intestinal nontyphoidal Salmonella infections in children. *Ann Trop Paediatr* 2000; 20: 125–29.
- 157 Lang R, Maayan MC, Lidor C, Savin H, Kolman S, Lishner M. Salmonella paratyphi C osteomyelitis: report of two separate episodes 17 years apart. Scand J Infect Dis 1992; 24: 793–96.
- 158 Goh KT. An outbreak of paratyphoid A in Singapore: clinical and epidemiological studies. *Southeast Asian J Trop Med Public Health* 1981; 12: 55–62.
- 159 Gotuzzo E, Echevarria J, Carrillo C, et al. Randomized comparison of aztreonam and chloramphenicol in treatment of typhoid fever. *Antimicrob Agents Chemother* 1994; 38: 558–62.
- 160 White NJ, Dung NM, Vinh H, Bethell D, Hien IT. Fluoroquinolone antibiotics in children with multidrug resistant typhoid. *Lancet* 1996; 348: 547.
- 161 Cao XT, Kneen R, Nguyen TA, Truong DL, White NJ, Parry CM. A comparative study of ofloxacin and cefixime for treatment of typhoid fever in children. The Dong Nai Pediatric Center Typhoid Study Group. *Pediatr Infect Dis J* 1999; 18: 245–48.
- 162 Girgis NI, Butler T, Frenck RW, et al. Azithromycin versus ciprofloxacin for treatment of uncomplicated typhoid fever in a randomized trial in Egypt that included patients with multidrug resistance. Antimicrob Agents Chemother 1999; 43: 1441–44.
- 163 Chinh NT, Parry CM, Ly NT, et al. A randomized controlled comparison of azithromycin and ofloxacin for treatment of multidrug-resistant or nalidixic acid-resistant enteric fever. *Antimicrob Agents Chemother* 2000; 44: 1855–59.
- 164 Gasem MH, Keuter M, Dolmans WM, et al. Persistence of Salmonellae in blood and bone marrow: randomized controlled trial comparing ciprofloxacin and chloramphenicol treatments against enteric fever. Antimicrob Agents Chemother 2003; 47: 1727–31.
- 165 Dutta P, Rasaily R, Saha MR, et al. Ciprofloxacin for treatment of severe typhoid fever in children. *Antimicrob Agents Chemother* 1993; 37: 1197–99.

- 166 Burkhardt JE, Hill MA, Carlton WW, Kesterson JW. Histologic and histochemical changes in articular cartilages of immature beagle dogs dosed with difloxacin, a fluoroquinolone. *Vet Pathol* 1990; 27: 162–70.
- 167 Stahlmann R, Kuhner S, Shakibaei M, et al. Chondrotoxicity of ciprofloxacin in immature beagle dogs: immunohistochemistry, electron microscopy and drug plasma concentrations. *Arch Toxicol* 2000; 73: 564–72.
- 168 Schaad UB, abdus Salam M, Aujard Y, et al. Use of fluoroquinolones in pediatrics: consensus report of an International Society of Chemotherapy commission. *Pediatr Infect Dis J* 1995; 14: 1–9.
- 169 Pradhan KM, Arora NK, Jena A, Susheela AK, Bhan MK. Safety of ciprofloxacin therapy in children: magnetic resonance images, body fluid levels of fluoride and linear growth. Acta *Paediatr* 1995; 84: 555–60.
- 170 Bethell DB, Hien TT, Phi LT, et al. Effects on growth of single short courses of fluoroquinolones. Arch Dis Child 1996; 74: 44–46.
- 171 Doherty CP, Saha SK, Cutting WA. Typhoid fever, ciprofloxacin and growth in young children. *Ann Trop Paediatr* 2000; **20**: 297–303.
- 172 Gendrel D, Chalumeau M, Moulin F, Raymond J. Fluoroquinolones in paediatrics: a risk for the patient or for the community? *Lancet Infect Dis* 2003; 3: 537–46.
- 173 Frenck RW Jr, Mansour A, Nakhla I, et al. Short-course azithromycin for the treatment of uncomplicated typhoid fever in children and adolescents. *Clin Infect Dis* 2004; 38: 951–57.
- 174 Butler T, Sridhar CB, Daga MK, et al. Treatment of typhoid fever with azithromycin versus chloramphenicol in a randomized multicentre trial in India. J Antimicrob Chemother 1999; 44: 243–50.
- 175 Frenck RW Jr, Nakhla I, Sultan Y, et al. Azithromycin versus ceftriaxone for the treatment of uncomplicated typhoid fever in children. *Clin Infect Dis* 2000; **31**: 1134–38.
- 176 Memon IA, Billoo AG, Memon HI. Cefixime: an oral option for the treatment of multidrug-resistant enteric fever in children. *South Med J* 1997; **90**: 1204–07.
- 177 Mandal S, Mandal M, Pal NK. In vitro efficacy of ciprofloxacin alone and in combination with amoxycillin against Salmonella typhi isolates. Indian J Exp Biol 2003; 41: 360–62.
- 178 Mandal S, Mandal MD, Pal NK. Combination effect of ciprofloxacin and gentamicin against clinical isolates of *Salmonella enterica* serovar typhi with reduced susceptibility to ciprofloxacin. *Jpn J Infect Dis* 2003; 56: 156–57.
- 179 Vinh H, Parry CM, Hanh VT, et al. Double blind comparison of ibuprofen and paracetamol for adjunctive treatment of uncomplicated typhoid fever. *Pediatr Infect Dis J* 2004; 23: 226–30.
- 180 Dutta P, Mitra U, Dutta S, De A, Chatterjee MK, Bhattacharya SK. Ceftriaxone therapy in ciprofloxacin treatment failure typhoid fever in children. *Indian J Med Res* 2001; 113: 210–13.
- 181 Tatli MM, Aktas G, Kosecik M, Yilmaz A. Treatment of typhoid fever in children with a flexible-duration of ceftriaxone, compared with 14day treatment with chloramphenicol. *Int J Antimicrob Agents* 2003; 21: 350–53.
- 182 Bhutta ZA, Khan IA, Shadmani M. Failure of short-course ceftriaxone chemotherapy for multidrug-resistant typhoid fever in children: a randomized controlled trial in Pakistan. *Antimicrob Agents Chemother* 2000; 44: 450–52.
- 183 Hoffman SL, Punjabi NH, Kumala S, et al. Reduction of mortality in chloramphenicol-treated severe typhoid fever by high-dose dexamethasone. N Engl J Med 1984; 310: 82–88.
- 184 Ferreccio C, Morris JG Jr, Valdivieso C, et al. Efficacy of ciprofloxacin in the treatment of chronic typhoid carriers. J Infect Dis 1988; 157: 1235–39.
- 185 Gotuzzo E, Guerra JG, Benavente L, et al. Use of norfloxacin to treat chronic typhoid carriers. J Infect Dis 1988; 157: 1221–25.
- 186 Steinberg EB, Bishop R, Haber P, et al. Typhoid fever in travelers: who should be targeted for prevention? *Clin Infect Dis* 2004; 39: 186–91.
- 187 Engels EA, Falagas ME, Lau J, Bennish ML. Typhoid fever vaccines: a meta-analysis of studies on efficacy and toxicity. *BMJ* 1998; 316: 110–16.

- 188 Klugman KP, Koornhof HJ, Robbins JB, Le Cam NN. Immunogenicity, efficacy and serological correlate of protection of *Salmonella typhi* Vi capsular polysaccharide vaccine three years after immunization. *Vaccine* 1996; 14: 435–38.
- 189 Yang HH, Kilgore PE, Yang LH, et al. An outbreak of typhoid fever, Xing-An County, People's Republic of China, 1999: estimation of the field effectiveness of Vi polysaccharide typhoid vaccine. J Infect Dis 2001; 183: 1775–80.
- 190 Proell S, Maiwald H, Nothdurft HD, et al. Combined vaccination against hepatitis A, hepatitis B, and typhoid fever: safety, reactogenicity, and immunogenicity. J Travel Med 2002; 9: 122–26.
- 191 Loebermann M, Kollaritsch H, Ziegler T, et al. A randomized, openlabel study of the immunogenicity and reactogenicity of three lots of a combined typhoid fever/hepatitis A vaccine in healthy adults. *Clin Ther* 2004; 26: 1084–91.
- 192 Keddy KH, Klugman KP, Hansford CF, Blondeau C, Bouveret le Cam NN. Persistence of antibodies to the Salmonella typhi Vi capsular polysaccharide vaccine in South African school children ten years after immunization. Vaccine 1999; 17: 110–13.
- 193 Jong EC, Kaplan KM, Eves KA, Taddeo CA, Lakkis HD, Kuter BJ. An open randomized study of inactivated hepatitis A vaccine administered concomitantly with typhoid fever and yellow fever vaccines. J Travel Med 2002; 9: 66–70.
- 194 Wahdan MH, Serie C, Cerisier Y, Sallam S, Germanier R. A controlled field trial of live *Salmonella typhi* strain Ty 21a oral vaccine against typhoid: three-year results. *J Infect Dis* 1982; 145: 292–95.
- 195 Levine MM, Ferreccio C, Abrego P, Martin OS, Ortiz E, Cryz S. Duration of efficacy of Ty21a, attenuated *Salmonella typhi* live oral vaccine. *Vaccine* 1999; 17 (suppl 2): S22–27.
- 196 Faucher JF, Binder R, Missinou MA, et al. Efficacy of atovaquone/proguanil for malaria prophylaxis in children and its effect on the immunogenicity of live oral typhoid and cholera vaccines. *Clin Infect Dis* 2002; **35**: 1147–54.
- 197 Begier EM, Burwen DR, Haber P, Ball R. Vaccine Adverse Event Reporting System Working Group. Postmarketing safety surveillance for typhoid fever vaccines from the Vaccine Adverse Event Reporting System, July 1990 through June 2002. *Clin Infect Dis* 2004; **38**: 771–79.
- 198 Mai NL, Phan VB, Vo AH, et al. Persistent efficacy of Vi conjugate vaccine against typhoid fever in young children. N Engl J Med 2003; 349: 1390–91.
- 199 Singh M, Ganguly NK, Kumar L, Vohra H. Protective efficacy and immunogenicity of Vi-porin conjugate against Salmonella typhi. Microbiol Immunol 1999; 43: 535–42.
- 200 Salazar-Gonzalez RM, Maldonado-Bernal C, Ramirez-Ciuz NE, et al. Induction of cellular immune response and anti-Salmonella enterica serovar typhi bactericidal antibodies in healthy volunteers by immunization with a vaccine candidate against typhoid fever. Immunol Lett 2004; 93: 115–22.
- 201 Tacket CO, Pasetti MF, Sztein MB, Livio S, Levine MM. Immune responses to an oral typhoid vaccine strain that is modified to constitutively express Vi capsular polysaccharide. J Infect Dis 2004; 190: 565–70.
- 202 Konadu EY, Lin FY, Ho VA, et al. Phase 1 and phase 2 studies of Salmonella enterica serovar paratyphi A O-specific polysaccharidetetanus toxoid conjugates in adults, teenagers and 2-4 year old children in Viet Nam. Infect Immun 2000; 68: 1529–34.
- 203 Metzger WG, Mansouri E, Kronawitter M, et al. Impact of vectorpriming on the immunogenicity of a live recombinant *Salmonella enterica* serovar typhi Ty21a vaccine expressing urease A and B from *Helicobacter pylori* in human volunteers. *Vaccine* 2004; 22: 2273–77.
- 204 Chiu CH, Wu TL, Su LH, et al. The emergence in Taiwan of fluoroquinolone resistance in *Salmonella enterica* serotype choleraesuis. N Engl J Med 2002; 34: 413–19.
- 205 Nakaya H, Yasuhara A, Yoshimura K, Oshihoi Y, Izumiya H, Watanabe H. Life-threatening infantile diarrhea from fluoroquinolone resistant to *Salmonella enterica typhimurium* with mutations in both gyrA and parC. *Emerg Infect Dis* 2003; 9: 255–57.