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Structural characterization of folded and extended conformations $\gamma_{9/2}$ peptides containing γ amino acids with proteinogenic side chains: Crystal structures of γ_n , $\alpha\gamma_n$ and $\gamma\gamma\delta\gamma$ sequences.

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Abstract

The crystal structures of nine peptides containing γ^4 Val and γ^4 Leu are described. The short sequences Boc-[$\gamma^4(R)$ Val]₂-OMe **1**, Boc-[$\gamma^4(R)$ Val]₃-NHMe **2** and Boc- $\gamma^4(S)$ Val- $\gamma^4(R)$ Val-OMe **3** adopt extended apolar, sheet like structures. The tetrapeptide Boc-[$\gamma^4(R)$ Val]₄-OMe **4** adopts an extended conformation, in contrast to the folded C₁₄ helical structure determined previously for Boc-[$\gamma^4(R)$ Leu]₄-OMe. The hybrid $\alpha\gamma$ sequence Boc-[Ala- $\gamma^4(R)$ Leu]₂-OMe **5** adopts an S-shaped structure devoid of intramolecular hydrogen bonds, with both α residues adopting local helical conformations. In sharp contrast, the tetrapeptides Boc-[Aib- $\gamma^4(S)$ Leu]₂-OMe **6** and Boc-[Leu- $\gamma^4(R)$ Leu]₂-OMe **7** adopt folded structures stabilized by two successive C₁₂ hydrogen bonds. γ^4 Val residues have also been incorporated into the strand segments of a crystalline octapeptide, Boc-Leu- $\gamma^4(R)$ Val-Val-^DPro-Gly-Leu- $\gamma^4(R)$ Val-Val-OMe **8**. The $\gamma\gamma\delta\gamma$ tetrapeptide containing γ^4 Val and δ^5 Leu residues adopts an extended sheet like structure. The hydrogen bonding pattern at γ residues corresponds to an apolar sheet, while a polar sheet is observed at the lone δ residue. The transition between folded and extended structures at γ residues involves a change of the torsion angle from the *gauche* to *trans* conformation about the C^β-C^γbond.

Introduction

Hydrogen bonded polypeptide structures have been extensively characterized in peptides and proteins, since the seminal work of Pauling and Corey introduced the α -helix and β -sheet to the literature of protein chemistry.¹ In conventional peptide sequences composed of α amino acids (α -peptides), a limited set of backbone hydrogen bonded motifs characterize the known elements of secondary structure. In the case of helices, the 3₁₀ and α -helical structures, which

are stabilized by $C_{10}(4\rightarrow 1, 10 \text{ atoms in the hydrogen bonded ring)}$ and $C_{13}(5\rightarrow 1, 13 \text{ atoms thick of the hydrogen bonded ring)}$, are the most widely observed structures. The extended segments of α -peptide chains commonly associate into the canonical parallel and anti-parallel β -sheet structures,² although the less common anti-parallel α -sheet structure has also been recently advanced as an important structural element in the case of aggregation prone sequences, implicated in amyloidogenesis.³ A key characteristic of the sheet structures is that the hydrogen bond donors and acceptors may lie far apart in a polypeptide sequence or are derived from different molecules, resulting in intermolecular association. The design of relatively short folded peptide sequences is implicitly based on the recognition that intramolecular hydrogen bonding is a dominant stabilizing interaction.

In recent years, the rapid development of the field of peptide *foldamers*⁴ based largely on sequences containing the higher homologs of the α amino acids, primarily β and γ residues,⁵ has refocused attention on the diversity of hydrogen bonded structures in polypeptides with unnatural backbones.⁶ In the case of helices, hydrogen bonding patterns unprecedented in the structural chemistry of α -peptides have been revealed in the characterization of the C₁₄ helix in β -peptides⁷ and helices with mixed hydrogen bond directionality in hybrid $\alpha\beta$ and $\alpha\gamma$ sequences.⁸ The polar sheet structures in which hydrogen bond acceptors (C=O groups) and donors (NH groups) are aligned in the opposite sides of an extended strand have been observed in β peptide sequences.⁹ As part of a program to systematically explore the conformational space and accessible regular structures for γ residues,¹⁰ derived by backbone homologation of proteinogenic amino acids, we have structurally characterized by X-ray diffraction a number of short γ peptides and $\alpha\gamma$ sequences.

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In this report, we describe the structural characterization of γ^4 Val and γ^4 Leu residues, (Fig. 1), in extended sheet structures and compare residue conformations in extended and helical structures. We also present the first structural characterization of a hybrid $\gamma\gamma\delta\gamma$ tetrapeptide in an extended sheet conformation.



Fig. 1 Chemical structures of the γ residues, a) γ^4 Val and b) γ^4 Leu.

Results and discussion

The solid state conformations and hydrogen bonding patterns observed in crystals are presented for nine synthetic peptides containing γ residues. The sequences and the observed backbone torsion angles are summarized in Table 1. Hydrogen bond parameters are listed in supplementary Table S1.

Peptides	Residues	φ	$\boldsymbol{\theta}_1$	θ_2	θ_3	ψ
$\mathbf{P}_{\mathbf{a}\mathbf{a}} \left[\mathbf{a}^{4}(\mathbf{P}) \mathbf{V}_{\mathbf{a}} \right] \mathbf{O} \mathbf{M}_{\mathbf{a}} 1$	$n^4(B)$ Vol (1)	122.0	64.3	178 3		1/3.6
$Boc-[\gamma(R) val]_2$ -Owle I	$\gamma(R)$ val (1)	-122.0 147.1	62.2	176.5		-145.0
$\mathbf{D}_{\alpha\alpha} \left[\mathbf{a}^{4}(\mathbf{D}) \mathbf{V}_{\alpha} \right] = \mathbf{N} \mathbf{U} \mathbf{M}_{\alpha} 2$	$\gamma(R)$ val (2)	-14/.1	66.2	170.1		-107.0
$Boc-[\gamma(R) val]_3$ -INHIME 2	$\gamma(R)$ val (1)	-122.0	61.2			-14/.1
	$\gamma(R)$ val (2)	-125.4	58.6	174.4		-125.1
$\mathbf{D}_{a} = \mathbf{A}^{4}(\mathbf{C})\mathbf{V}_{a} + \mathbf{A}^{4}(\mathbf{D})\mathbf{V}_{a} + \mathbf{O}\mathbf{M}_{a} + 2$	$\gamma(R)$ val (3)	-112.0	50.0	173.0		-125.0
$Boc-\gamma(S)$ val- $\gamma(R)$ val-OMe 3	γ (S) val (1)	115./	-07.7	175.0		145.2
$\mathbf{D}_{a} = [\mathbf{A}^{4}(\mathbf{D})\mathbf{X}_{a}]^{1}$ OM \mathbf{A}	$\gamma(R)$ val (2)	-130.5	59.5 68 5	-177.3		-107.0
Boc- $[\gamma(R)$ val] ₄ -OMe 4	$\gamma(R)$ val (1)	-12/.4	00.5	-1/9,4		-145.2
	$\gamma(R)$ val (2)	-110.9	00.5	1/0.4		-144.0
	$\gamma(R)$ val (3)	-117.0	05.7	170.9		-145./
	$\gamma(R)$ Val (4)	-140.2	03.9	1/4.0		-1/1.5
Boc-[Ala- γ (<i>R</i>)Leu] ₂ -OMe 5	$\frac{\text{Ala}(1)}{2}$	-04.5	54.2	176 1		-38.2
Molecule-1	γ (R)Let (2)	-141.4	54.2	-1/0.1		-147.0
	Ald (5)	-03.9 122.9	60.8	170.6		-29.7
Malagula 2	γ (R)Leu (4)	-133.0	00.0	170.0		135.5
Molecule-2	$\frac{Ala(1)}{a^4(B)L} = (2)$	-01.0 127.2	55.3	177.6		-43.0
	γ (R)Let (2)	-137.2	55.5	-1//.0		-151.0
	$\frac{Ala(5)}{(4)}$	-07.5	(17	1(5.0		-20.2
$\mathbf{D}_{ab} = [A; \mathbf{h}_{ab}]^{4} (\mathbf{\Omega}) \mathbf{L}_{ab} = \mathbf{O} \mathbf{M}_{ab} \mathbf{C}$	γ (R)Leu (4)	-144.2	01./	105.9		155.0
Boc-[Alb- γ (5)Leu] ₂ -OMe 6	$\frac{AID(1)}{2}$	39.8 122.9	51.0	64.1		40.9
	γ (S)Let (2)	125.0	-51.9	-04.1		20.1
	$\frac{AID(5)}{2}$	03.5	54.0	172 7		29.1
	γ (S)Let (4)	(7.7	-54.9	-1/3./		-104.1
Boc-[Leu- γ (<i>R</i>)Leu] ₂ -OMe 7	$\frac{\text{Leu}(1)}{4}$	-0/./	AC 7	(=)		-33.4
	$\gamma(R)$ Leu (2)	-121.2	40./	07.3		-117.5
	$\frac{\text{Leu}(3)}{4(\mathbf{D})\mathbf{L}(4)}$	-03.1	(0.1	15(0		-33./
	$\gamma(R)$ Leu (4)	-101.7	60.1	-176.8		-175.2
Boc-Leu- $\gamma'(R)$ Val-Val-Pro-	Leu (1)	-156.3	71.0	1(()		144.7
Gly-Leu- $\gamma'(R)$ val-val-OMe 8	$\gamma^{\prime}(R)$ Val (2)	-128.8	71.8	-166.2		-142.8
	Val(3)	-167.1				141.7
	⁵ Pro (4)	58.9				-138.3
	Gly (5)	-83.2				-7.9
	Leu (6)	-78.3	() 0	4=0.0		134.5
	$\gamma^{4}(R)$ Val (7)	-146.1	63.0	-179.9		-125.9
5	Val (8)	-67.4	<i>c</i> a a			-39.2
Boc- $[\gamma^{*}(R)$ Leu] ₂ - $\delta^{3}(R)$ Leu-	$\gamma^{\dagger}(R)$ Leu (1)	-116.5	60.3	172.0		-130.8
$\gamma^*(R)$ Leu-OMe 9	$\gamma^{4}(R)$ Leu (2)	-109.0	60.4	176.5		-134.4
	$\delta^{3}(R)$ Leu (3)	-120.7	65.7	173.6	-64.4	121.2
	$\gamma^4(R)$ Leu (4)	-139.5	57.3	-174.0		174.8

Homooligomeric y sequences

Di- and tri-peptides: Fig. 2 shows the structures of the peptides, Boc- $[\gamma^4(R)Val]_2$ -OMe 1, Boc- $[\gamma^4(R)Val]_3$ -NHMe 2 and Boc- $\gamma^4(S)Val-\gamma^4(R)Val$ -OMe 3. In all three cases the molecules are extended with intermolecular hydrogen bonds linking adjacent chains which are aligned in parallel fashion.



Fig. 2 Molecular conformations observed for short di- and tri-peptides in crystals: a) Boc- $[\gamma^4(R)Val]_2$ -OMe **1**, b) Boc- $[\gamma^4(R)Val]_3$ -NHMe **2** and c) Boc- $\gamma^4(S)Val-\gamma^4(R)Val$ -OMe **3**. Molecules are packed in form an *apolar* sheet like structure in the crystals of d) Boc- $[\gamma^4(R)Val]_2$ -OMe **1**, e) Boc- $[\gamma^4(R)Val]_3$ -NHMe **2** and f) Boc- $\gamma^4(S)Val-\gamma^4(R)Val-OMe$ **3**.

The observed conformational angles for the γ residues establish that both $\phi(N-C^{\gamma})$ and $\psi(C^{\alpha}-C^{\alpha})$ correspond to extended conformations. The torsion angle $\theta_1(C^{\gamma}-C^{\beta})$ corresponds to a folded, *gauche* conformation, while $\theta_2(C^{\beta}-C^{\alpha})$ is *trans* and fully extended. In the observed sheet arrangement, alternating NH and CO groups point on opposite sides of the extended strand resulting in an *apolar* sheet arrangement, analogous to that observed in α polypeptides. *Tetrapeptide*: Fig. 3 shows the observed solid state conformation for Boc-[$\gamma^4(R)$ Val]₄-OMe **4**. The molecule adopts an extended conformation, resulting in an extended version of the parallel *apolar* sheet structure, observed in di- and tripeptide sequences (**1** and **2**). Once again, $\theta_1(C^{\gamma}-C^{\beta})$ adopts a *gauche* conformation, while the other three backbone torsion angles are extended at the γ residues.

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Fig. 3 a) Solid state conformation for the tetrapeptide, Boc- $[\gamma^4(R)Val]_4$ -OMe **4**. b) Solid state conformation of the previously reported helical tetrapeptide, Boc- $[\gamma^4(R)Leu]_4$ -OMe.¹¹ c) Molecules are packed in form an *apolar* sheet like structure in the crystals of Boc- $[\gamma^4(R)Val]_4$ -OMe **4**.

Interestingly, in a previously report, the closely related $\gamma\gamma\gamma\gamma$ sequences, Boc-[$\gamma^4(R)$ Leu]₄-OMe and its corresponding *enantiomer* Boc-[$\gamma^4(S)$ Leu]₄-OMe, were shown to adopt folded helical structure in crystals.¹¹ Fig.3 compares the observed conformations for the two related $\gamma\gamma\gamma\gamma$ sequences. The sole difference observed in backbone torsion angles is that in the extended structure, Boc-[$\gamma^4(R)$ Val]₄-OMe, $\theta_2(C^{\beta}-C^{\alpha})$ adopts a *trans* conformation, while in the folded helical structure $\theta_2(C^{\beta}-C^{\alpha})$ corresponds to a *gauche* conformation. The dramatic structural difference observed between the two closely related γ -tetrapeptides can be realized by a conformational switch (*gauche*=*trans*) about the C^{β}-C^{α}bond. In a previous study using IR and NMR methods in CHCl₃ solution, we had indeed suggested folded C₁₄ helical conformations for Boc-[$\gamma^4(R)$ Val]₄-OMe.¹² Presumably, in this case the tetrapeptide adopts a predominantly folded conformation. Crystals obtained from the more strongly solvating medium, methanol, yield an extended conformation, suggesting that relatively small energy differences, exist between these conformations, resulting in pronounced solvent dependence.

Hybrid αγ sequences

Fig. 4a, 4c and 4d show the observed solid state conformation for three $\alpha\gamma\alpha\gamma$ tetrapeptides, Boc-[Ala- $\gamma^4(R)$ Leu]₂-OMe **5**, Boc-[Aib- $\gamma^4(S)$ Leu]₂-OMe **6** and Boc-[Leu- $\gamma^4(R)$ Leu]₂-OMe **7**.

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Fig. 4 Observed solid state conformation for three $\alpha\gamma\alpha\gamma$ tetrapeptides, a) Boc-[Ala- $\gamma^4(R)$ Leu]₂-OMe **5**, c) Boc-[Aib- $\gamma^4(S)$ Leu]₂-OMe **6** and d) Boc-[Leu- $\gamma^4(R)$ Leu]₂-OMe **7**. b) Packing of molecules in the crystals of Boc-[Ala- $\gamma^4(R)$ Leu]₂-OMe **5**.

In Boc-[Ala- $\gamma^4(R)$ Leu]₂-OMe **5** an 'S' shaped structure is observed for the polypeptide backbone which is devoid of intramolecular hydrogen bonds. This folded arrangement is a consequence of the fact that both the α residues, Ala(1) and Ala(3) adopts helical α_R conformations, while the γ residues retain the conformation observed in the extended structures of homooligomeric γ peptides. Two independent molecules are observed in the crystallographic asymmetric unit, which are linked by intermolecular hydrogen bonds, as shown in Fig. 4b. The structure of tetrapeptide **5** raises the possibility of exploring partially folded states of polypeptide backbones which are not stabilized by intramolecular hydrogen bonds.

The peptides Boc-[Aib- $\gamma^4(S)$ Leu]₂-OMe **6** and Boc-[Leu- $\gamma^4(R)$ Leu]₂-OMe **7** fold into helical conformations stabilized by two successive C₁₂ hydrogen bonds. These two structures may be classified as the backbone expanded analogues of the incipient 3₁₀ helix, first characterized in the 1970s for tetrapeptides containing the conformationally constrained Aib (α -aminoisobutyric acid) residue.¹³ Comparison of the observed structures of peptides **5** and **7** is instructive. In both cases, α and γ residues bear only a single substituent and may be considered as largely unconstrained. The dramatic difference in structures for the two tetrapeptides suggests that in these short sequences conformational choice may be mainly

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determined by subtle environmental factors, primarily solvent interactions in solutions^{VientAdicle Online} crystal contact forces in the solid state. The sole difference in backbone conformational angles is observed for $\theta_2(C^{\beta}-C^{\alpha})$, which is *trans* for the residue 2, $\gamma^4(R)$ Leu(2) and *gauche* for the γ^4 Leu(2) residue in peptides 5 and 7, respectively. Interestingly, in both the helical structures, 6 and 7, θ_2 at the C-terminus γ residue is *trans*. In the case of this residue only the NH group contributes to the intramolecular hydrogen bonded pattern, leaving the conformational choice at this terminal residue open.

Facing γ residue in an antiparallel strands of a β hairpin

Previous studies on designed peptide hairpins have established that β residues can be readily accommodated at facing positions of antiparallel strands.¹⁴



Fig. 5 a) The molecular conformation observed in crystals for Boc-Leu- $\gamma^4(R)$ Val-Val-^DPro-Gly-Leu- $\gamma^4(R)$ Val-Val-OMe **8**. b) Packing of molecules reveals an anti-parallel sheet motif for the octapeptide **8**.

 OMe 8. Fig.5 shows the molecular conformation observed in crystals. The observed packing icle online arrangement reveals an anti-parallel sheet motif. In the structure of the hairpin the ^DPro-Gly segment adopt the hairpin facilitating type II' β -turn¹⁶, while the α residues with the exception of the C terminus Val(8) adopt extended conformation in the P_{II} region of ϕ - ψ space. The $\gamma^4(R)$ Val(2) and $\gamma^4(R)$ Val(7)have conformations identical to that observed in the short extended peptides, 1, 2, 3 and 4 with θ_1 adopting *gauche* conformation, while θ_2 is *trans*.

A hybrid γγδγ tetrapeptide

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In an attempt to extend our study to the higher ω amino acids we determined the crystal structure of the tetrapeptide, Boc- $\gamma^4(R)$ Leu- $\gamma^4(R)$ Leu- $\delta^5(R)$ Leu- $\gamma^4(R)$ Leu-OMe 9. The observed molecular conformation corresponds to an extended polypeptide with molecules arranged parallel to one another linked by hydrogen bonds into sheet like structures.



Fig. 6 a) Chemical structure of the δ residue, δ^5 Leu. b) Molecular conformations observed for tetrapeptide, Boc- $\gamma^4(R)$ Val- $\gamma^4(R)$ Val- $\delta^5(R)$ Leu- $\gamma^4(R)$ Val-OMe **9**, in crystals. c) Packing of molecules in crystals for the tetrapeptide reveals an overall apolar sheet like structure, while at the $\delta^5(R)$ Leu(3) a polar sheet hydrogen bonding pattern was observed.

Inspection of the packing diagrams in Fig.6 reveals that at the γ residues an *apolar* sheet pattern is observed, while at the lone δ residue a *polar* sheet hydrogen bonding pattern is seen. At the δ residue, adjacent NH groups point on one side of the sheet, while CO groups are oriented on the opposite side. The $\delta^5(R)$ Leu residue adopts a conformation in which ϕ and ψ are extended. The torsion angles about the central C-C bonds are 65.7°, 173.6° and -64.4°,

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corresponding to a g^+ , t, g^- conformation about the θ_1 , θ_2 and θ_3 torsional variables. Very Article Online previously reported structure containing the unconstrained δ residue, δ Ava (δ -aminovaleric acid), the observed torsion angles are, $\phi = -127^\circ$, $\theta_1 = 53^\circ$, $\theta_2 = 64^\circ$, $\theta_3 = 167^\circ$ and $\psi = 77^\circ$.¹⁷ In that example, the δ Ava residue occupied the i+2 position in an expanded ^DPro- δ Ava C₁₃ turn which nucleated a hairpin structure, Boc-Leu-Val-Val-^DPro- δ Ava-Leu-Val-Val-OMe. These observations suggest that δ residues can also be considered as monomers of choice in the design of new *foldamer* structures.

Conclusions

The results presented above, together with the growing number of studies on peptides containing unconstrained γ residues with proteinogenic sidechains¹⁸ provides an opportunity to understand the nature of the conformational transitions that occurs upon the conversion of folded helices into extended sheet like structures. Fig.7 presents a cluster plot of the observed θ_1 , θ_2 values for γ^4 residues with proteinogenic sidechains (primarily γ^4 Val, γ^4 Leu and γ^4 Phe) in peptide crystal structures. In both helices and sheets, the ϕ , ψ values lie in the same region of conformational space ($\phi \approx -125^\circ$, $\psi \approx -128^\circ$ for helices; $\phi \approx -129^\circ$, $\psi \approx -154^\circ$ for sheets).



Fig. 7 A θ_1 - θ_2 scatter plot for γ^4 residues with proteinogenic sidechains.

Two distinct clusters are observed in the θ_1 - θ_2 plot corresponding to a transition of θ_2 from *gauche* ($\approx 60^\circ$) to *trans* ($\approx 180^\circ$). The use of γ residues, synthetically accessible from readily available proteinogenic α residues, may be exploited in the design of *foldamers* which mimic structures observed in proteins.

Experimental

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Synthesis

General: The γ amino acids, Boc- $\gamma^4(R)$ Val-OH, Boc- $\gamma^4(S)$ Val-OH, Boc- $\gamma^4(R)$ Leu-OH, and Boc- $\delta^5(R)$ Leu-OH were synthesized by previously described procedures.¹⁹ Peptide synthesis procedures.²⁰ solution-phase The conventional was carried out using tertbutyloxycarbonyl(Boc) group was used for N terminal protection and C terminus was protected using a methyl ester. Deprotections were performed using 98% formic acid to remove the Boc group, whereas the methyl ester was removed by alkaline hydrolysis. The details of the synthesis are discussed below. The final peptides were purified by reversedphase high-performance liquid chromatography (HPLC) on Jupitor Proteo C₁₂ column (10-250 mm, 4 µ particle size) using methanol/water systems and monitored at 226 nm. Melting points were determined using Stuart melting point apparatus SMP10. Electrospray ionization mass spectra (ESI-MS) were obtained on a Bruker Daltonics Esquire-3000 instrument. ¹H NMR spectra were recorded on Bruker 500/700 MHz spectrometers. Characterization data for the peptides 1-9 are given in the Electronic Supplementary Information (ESI).†

X-ray crystallography

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Crystals for the peptides, Boc- $[\gamma^4(R)Val]_2$ -OMe 1, Boc- $[\gamma^4(R)Val]_3$ -NHMe 2, Boc- $[\gamma^4(R)Val]_4$ -OMe 4, Boc-[Leu- $\gamma^4(R)Leu]_2$ -OMe 7 and Boc-Leu- $\gamma^4(R)Val$ -Val-DPro-Gly-Leu- $\gamma^4(R)Val$ -Val-OMe 8 were assigned to the monoclinic space group *P*2₁ from systematic absences (0k0, k = odd). Boc-[Ala- $\gamma^4(R)Leu]_2$ -OMe 5, Boc-[Aib- $\gamma^4(S)Leu]_2$ -OMe 6 and Boc- $[\gamma^4(R)Leu]_2$ - $\delta^5(R)Leu-\gamma^4(R)Leu$ -OMe 9 crystallized in the triclinic space group *P*1. The dipeptide, Boc- $\gamma^4(S)Val-\gamma^4(R)Val$ -OMe 3 crystallized in the monoclinic space group *C*2 and the space group assignment was done from the systematic absences (hkl, h+k = odd). Peptides 1, 2, 3, 4 and 9 crystallized with one peptide molecule in the asymmetric unit and were free of solvent. The tetrapeptide 5 crystallized with two peptide molecules in the asymmetric unit. The peptides, 6, 7 and 8 crystallized with one peptide molecule); 7 (two water molecules); 8 (one water molecule)] in the crystallographic asymmetric unit.

For peptides 1, 3, 4 and 8 X-ray data were collected on BRUKER AXS SMART APEXII ULTRA CCD (rotating anode X-ray generator) with CuK α (1.54178 Å) radiation and for peptides 2, 5, 6, 7 and 9 X-ray data were collected on BRUKER AXS KAPPA APEXII CCD with MoK α (0.71073 Å) radiation. All the data sets were collected at room temperature (296K), in phi and omega scan type mode.

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All peptides structures were solved by using iterative dual space direct methodisw price Online SHELXD.²¹ After the initial solution methods, all the structures were refined against F^2 isotropically followed by full matrix anisotropic least-squares refinement using SHELXL-97.²² In the case of peptide 1, only two hydrogen atoms attached to N2 and C2G were located from the difference Fourier map and rest of the hydrogen atoms were fixed geometrically in idealized positions and allowed to ride with the C or N atoms to which they were bonded, in the final cycles of refinement. In the case of peptides 2 and 4, all the hydrogen atoms were fixed geometrically in the idealized position and allowed to ride with the non-hydrogen atoms to which they were bonded. For peptides 3, 5, 6, 7, 8 and 9 a few hydrogen atoms bonded to respective C or N atoms were located from difference Fourier maps [3 (N1, N2, C1A, C1B, C1G, C1D, C2B, C2G and C2D); 5 (N1, N2, N3, N4, C1A, C1G, C2A, C2B, C2G, C2D, C2E, C3A, C3B, C3G, C4G, C4D); 6 (N1, N3, N4); 7 (N1, N2, N3, N4, C1A, C1G, C2A, C2B, C2G, C2D, C2E, C3A, C3B, C3G, C4G, C4D); 8 (N1, N2, N3, N5, N6, N7, N8, C1A, C2A, C2B, C2G, C2D, C3A, C3B, C4A, C6A, C6B, C7A, C7B, C7G, C7D, C8A, C8B); 9 (N1, N2, N3, N4, C1B, C1G, C1D, C2G, C3A, C3B, C3G, C3D, C4G, C4D, C4E)] and rest of the hydrogen atoms were fixed geometrically in the idealized position and allowed to ride with the C or N atoms to which they were bonded, in the final cycles of refinement. For the octapeptide, 8, positional disorder was observed at three atomic sites. One was at the site of C^{γ} position (C4G) of the ^DPro (4) residue and other two at the $C^{\delta 1}$ (C6D1) and $C^{\delta 2}$ (C6D2) positions of Leu (6) residue sidechain. Positional disorder at different atomic sites of the crystal structure mentioned above was treated with partial occupancy and also with proper restraints and constraints to get chemically meaningful geometry of the disordered groups. Non-hydrogen atoms of the solvent molecules in peptides, 6, 7 and 8 were also located from difference Fourier map. The details of the crystal data and structure refinement parameters for all the peptides mentioned above are given as the Supplementary Tables, ST1, ST2 and ST3.

CCDC deposition numbers for the peptides are 1038826 (1), 1038825 (2), 1038828 (3), 1038827 (4), 1038821 (5), 1038829 (6), 1038822 (7), 1038823 (8) and 1038824 (9) which contain the supplementary crystallographic data for this paper. These data can be obtained free of charge from the Cambridge Crystallographic Data Centre *via* www.ccdc.cam.ac.uk/data_request/cif.

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